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APPLICATION FOR LETTERS PATENT

for

STREPTOCOCCUS SUIS VACCINES AND DIAGNOSTIC TESTS

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STREPTOCOCCUS SUIS VACCINES AND DIAGNOSTIC TESTS

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority to, and is a continuation of, International Application No. PCT/NL99/00460, filed on July 19, 1999, designating the United States of America, the contents of which are incorporated herein by this reference, the PCT International Patent Application itself claiming priority from European Patent Office Application Serial No. 98202465.5 filed July 22, 1998 and European Patent Office Application Serial No. 98202467.1 filed July 22, 1998.

JUN 29 2004

TECHNICAL FIELD

[0002] The invention relates to *Streptococcus* infections in pigs, vaccines directed against those infections, tests for diagnosing *Streptococcus* infections and bacterial vaccines. More particularly, the invention relates to vaccines directed against *Streptococcus* infections. TECH CENTER 1600/2900

BACKGROUND OF THE INVENTION

[0003] *Streptococcus* species, of which a large variety cause infections in domestic animals and man, are often grouped according to Lancefield's groups. Typing according to Lancefield occurs on the basis of serological determinants or antigens that are, among others, present in the capsule of the bacterium, and allows for only an approximate determination. Often, bacteria from different groups show cross-reactivity with each other, while other Streptococci cannot be assigned a group-determinant at all. Within groups, further differentiation is often possible on the basis of serotyping. These serotypes further contribute to the large antigenic variability of Streptococci, a fact that creates an array of difficulties within diagnosis of and vaccination against Streptococcal infections.

[0004] Lancefield group A *Streptococcus* species (Group A *Streptococci* "GAS," *Streptococcus pyogenes*) are common in children, causing nasopharyngeal infections and complications thereof. Among animals, cattle are especially susceptible to GAS infection which can cause mastitis.

[0005] Group A streptococci are the etiologic agents of streptococcal pharyngitis and impetigo, two of the most common bacterial infections in children, as well as a variety of less common, but potentially life-threatening, infections including soft tissue infections, bacteremia, and pneumonia. In addition, GAS are uniquely associated with the post-infectious autoimmune syndromes of acute rheumatic fever and post streptococcal glomerulonephritis.

[0006] Several recent reports suggest that the incidence of both serious infections due to GAS and acute rheumatic fever has increased during the past decade, focusing renewed interest on defining the attributes or virulence factors of the organism that may play a role in the pathogenesis of these diseases.

[0007] GAS produce several surface components and extracellular products that may be important in virulence. The major surface protein, M protein, has been studied in the most detail and has been convincingly shown to play a role in both virulence and immunity. Isolates rich in M protein are able to grow in human blood, a property thought to reflect the capacity of N protein to interfere with phagocytosis, and these isolates tend to be virulent in experimental animals.

[0008] Lancefield group B *Streptococcus* (“GBS”) are most often seen in cattle, causing mastitis; however, human infants are susceptible as well, often with fatal consequences. Group B streptococci (GBS) constitute a major cause of bacterial sepsis and meningitis among human neonates born in the United States and Western Europe and are emerging as significant neonatal pathogens in developing countries as well.

[0009] It is estimated that GBS strains are responsible for 10,000 to 15,000 cases of invasive infection in neonates in the United States alone. Despite advances in early diagnosis and treatment, neonatal sepsis due to GBS continues to carry a mortality rate of 15 to 20%. In addition, survivors of GBS meningitis have 30 to 50% incidence of long-term neurologic sequelae. Over the past two decades, increasing recognition of GBS as an important pathogen for human infants has generated renewed interest in defining the bacterial and host factors important in virulence of GBS and in the immune response to GBS infection.

[0010] Particular attention has focused on the capsular polysaccharide as the predominant surface antigen of the organisms. In a modification of the system originally developed by Rebecca Lancefield, GBS strains are serotyped on the basis of antigenic differences in their capsular

polysaccharides and the presence or absence of serologically defined C proteins. While GBS isolated from non-human sources often lack a serologically detectable capsular, a large majority of strains associated with neonatal infection belong to one of four major capsular serotypes, 1a, 1b, II or III. The capsular polysaccharide forms the outermost layer around the exterior of the bacterial cell, superficial to the cell wall. The capsule is distinct from the cell wall-associated group B carbohydrate. It has been suggested that the presence of sialic acid, in the capsule of bacteria that causes meningitis, is important for allowing these bacteria to breach the blood-brain barrier. Indeed, in *S. agalactiae*, sialic acid has been shown to be critical for the virulence function of the type III capsule. The capsule of *S. suis* serotype is composed of glucose, galactose, N-acetylglucosamine, rhamnose and sialic acid.

[0011] The group B polysaccharide, in contrast to the type-specific capsule, is present on all GBS strains and is the basis for serogrouping the organisms into Lancefield's group B. Early studies by Lancefield and co-workers showed that antibodies raised in rabbits against whole GBS organisms protected mice against challenge with strains of homologous capsular type, demonstrating the central role of the capsular polysaccharide as a protective antigen. Studies in the 1970s by Baker and Kasper demonstrated that cord blood of human infants with type III GBS sepsis uniformly had low or undetectable levels of antibodies directed against the type III capsule, suggesting that a deficiency of anticapsular antibody was a key factor in susceptibility of human neonates to GBS disease.

[0012] Lancefield group C infections, such as those with *S. equi*, *S. zooepidemicus*, *S. dysgalactiae*, and others, are mainly seen in horses, cattle and pigs, but can also cross the species barrier to humans. Lancefield group D (*S. bovis*) infections are found in all mammals and some birds, sometimes resulting in endocarditis or septicemia.

[0013] Lancefield groups E, G, L, P, U and V (*S. porcinus*, *S. canis*, *S. dysgalactiae*) are found in various hosts, causing neonatal infections, nasopharyngeal infections or mastitis.

[0014] Within Lancefield groups R, S and T (and with ungrouped types), *Streptococcus suis* is an important cause of meningitis, septicemia, arthritis and sudden death in young pigs (4, 46). Incidentally, it can also cause meningitis in man (1). *S. suis* strains are usually identified and classified by their morphological, biochemical and serological characteristics (58, 59, 46).

Serological classification is based on the presence of specific antigenic polysaccharides. So far, 35 different serotypes have been described (9, 56, 14). In several European countries, *S. suis* serotype 2 is the most prevalent type isolated from diseased pigs, followed by serotypes 9 and 1. Serological typing of *S. suis* is performed using different types of agglutination tests. In these tests, isolated and biochemically characterized *S. suis* cells are agglutinated with a panel of 35 specific sera. These methods are very laborious and time-consuming.

[0015] Little is known about the pathogenesis of the disease caused by *S. suis*, let alone about its various serotypes such as type 2. Various bacterial components, such as extracellular and cell-membrane associated proteins, fimbriae, hemagglutinins, and hemolysis, have been suggested as virulence factors (9, 10, 11, 15, 16, 47, 49). However, the precise role of these protein components in the pathogenesis of the disease remains unclear (37). It is well known that the polysaccharide capsule of various Streptococci and other gram-positive bacteria plays an important role in pathogenesis (3, 6, 35, 51, 52). The capsule enables these microorganisms to resist phagocytosis and is therefore regarded as an important virulence factor. Recently, a role of the capsule of *S. suis* in the pathogenesis was suggested as well (5). However, the structure, organization and function of the genes responsible for capsule polysaccharide synthesis ("cps") in *S. suis* is unknown. Within *S. suis*, serotype 1 and 2 strains can differ in virulence for pigs (41, 45, 49). Some type 1 and 2 strains are virulent, other strains are not. Because both virulent and non-virulent strains of serotype 1 and 2 strains are fully encapsulated, it may even be that the capsule is not a relevant factor required for virulence.

[0016] Attempts to control *S. suis* infections or disease are still hampered by the lack of knowledge about the epidemiology of the disease and the lack of effective vaccines and sensitive diagnostics. It is well known and generally accepted that the polysaccharide capsule of various Streptococci and other gram-positive bacteria plays an important role in pathogenesis. The capsule enables these microorganisms to resist phagocytosis and is therefore regarded as an important virulence factor.

[0017] Compared to encapsulated *S. suis* strains, non-encapsulated *S. suis* strains are phagocytosed by murine polymorphonuclear leucocytes to a greater degree. Moreover, an increase in thickness of capsule was noted for *in vivo* grown virulent strains while no increase was observed for

avirulent strains. Therefore, these data again demonstrate the role of the capsule in the pathogenesis for *S. suis* as well.

[0018] Ungrouped *Streptococcus* species, such as *S. mutans*, causing caries in humans, *S. uberis*, causing mastitis in cattle, and *S. pneumonia*, causing major infections in humans, and *Enterococcus faecalis* and *E. faecium*, further contribute to the large group of Streptococci.

[0019] *Streptococcus pneumoniae* (the pneumococcus) is a human pathogen causing invasive diseases, such as pneumonia, bacteremia, and meningitis. Despite the availability of antibiotics, pneumococcal infections remain common and can still be fatal, especially in high-risk groups, such as young children and elderly people. Particularly in developing countries, many children under the age of five years die each year from pneumococcal pneumonia. *S. pneumoniae* is also the leading cause of otitis media and sinusitis. These infections are less serious, but nevertheless incur substantial medical costs, especially when leading to complications, such as permanent deafness. The normal ecological niche of the pneumococcus is the nasopharynx of man. The entire human population is colonized by the pneumococcus at one time or another, and at a given time, up to 60% of individuals may be carriers. Nasopharyngeal carriage of pneumococci by man is often accompanied by the development of protection against infection by the same serotype. Most infections do not occur after prolonged carriage but follow exposure to recently acquired strains. Many bacteria contain surface polysaccharides that act as a protective layer against the environment. Surface polysaccharides of pathogenic bacteria usually make the bacteria resistant to the defense mechanisms of the host, for example, the lytic action of serum or phagocytosis. In this respect, the serotype-specific capsular polysaccharide ("CP") of *Streptococcus pneumoniae*, is an important virulence factor. Unencapsulated strains are avirulent, and antibodies directed against the CP are protective. Protection is serotype specific; each serotype has its own, specific CP structure. Ninety different capsular serotypes have been identified. Currently, CPs of 23 serotypes are included in a vaccine.

[0020] Vaccines directed against *Streptococcus* infections typically aim to utilize an immune response directed against the polysaccharide capsule of the various *Streptococcus* species, especially since the capsule is considered a primary virulence factor for these bacteria. During

infection, the capsule provides resistance against phagocytosis and thus protects the bacteria from the immune system of the host, and from elimination by macrophages and neutrophils.

[0021] The capsule particularly confers the bacterium resistance to complement-mediated opsonophagocytosis. In addition, some bacteria express capsular polysaccharides (CPs) that mimic host molecules, thereby avoiding the immune system of the host. Also, even when the bacteria have been phagocytosed, intracellular killing is hampered by the presence of a capsule.

[0022] It is generally thought that the bacterium will get recognized by the immune system through the anticapsular-antibodies or serum-factors bound to its capsule and will, through opsonization, get phagocytosed and killed only when the host has antibodies or other serum factors directed against capsule antigens.

[0023] However, these antibodies are serotype-specific, and will often only confer protection against only one of the many serotypes known within a group of *Streptococci*.

[0024] For example, current commercially available *S. suis* vaccines, which are generally based on whole-cell-bacterial preparations, or on capsule-enriched fractions of *S. suis*, confer only limited protection against heterologous strains. Also, the current pneumococcal vaccine that was licensed in the United States in 1983, consists of purified CPs of 23 pneumococcal serotypes whereas at least 90 CP types exist.

[0025] The composition of this pneumococcal vaccine was based, on the frequency of the occurrence of disease isolates in the US and cross-reactivity between various serotypes. Although this vaccine protects healthy adults against infections caused by serotypes included in the vaccine, it fails to raise a protective immune response in infants younger than 18 months and it is less effective in elderly people. In addition, the vaccine confers only limited protection in patients with immunodeficiencies and hematology malignancies.

[0026] Thus, improved vaccines are needed against *Streptococcus* infections. Much attention is directed toward producing CP vaccines by producing the relevant polysaccharides via chemical or recombinant means. However, chemical synthesis of polysaccharides is costly, and capsular polysaccharide synthesis by recombinant means necessitates knowledge about the relevant genes, which is not always available, and needs to be determined for every relevant serotype.

DISCLOSURE OF THE INVENTION

[0027] The invention provides an isolated or recombinant nucleic acid encoding a capsular (*cps*) gene cluster of *Streptococcus suis*. Biosynthesis of capsule polysaccharides has generally been studied in a number of Gram-positive and Gram-negative bacteria (32). In Gram-negative bacteria, but also in a number of Gram-positive bacteria, genes which are involved in the biosynthesis of polysaccharides are clustered at a single locus.

[0028] *Streptococcus suis* capsular genes, as provided by the invention, show a common genetic organization involving three distinct regions. The central region is serotype specific and encodes enzymes responsible for the synthesis and polymerization of the polysaccharides. The central region is flanked by two regions conserved in *Streptococcus suis* which encode proteins for common functions, such as transport of the polysaccharide across the cellular membrane. However, between species, only low homologies exist, hampering easy comparison and detection of seemingly similar genes. Knowing the nucleic acid encoding the flanking regions allows type-specific determination of nucleic acid of the central region of *Streptococcus suis* serotypes as, for example, described herein.

[0029] The invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* or a gene or gene fragment derived thereof. Such a nucleic acid is, for example, provided by hybridizing chromosomal DNA derived from any one of the *Streptococcus suis* serotypes to a nucleic acid encoding a gene derived from a *Streptococcus suis* serotype 1, 2 or 9 capsular gene cluster, as provided by the invention (see, for example, Tables 4 and 5) and cloning of (type-specific) genes as, for example, described herein. At least 14 open reading frames are identified. Most of the genes belong to a single transcriptional unit, identifying a coordinate control of these genes. The genes, and the enzymes and proteins they encode, act in concert to provide the capsule with the relevant polysaccharides.

[0030] The invention provides *cps* genes and proteins encoded thereof involved in regulation (CpsA), chain length determination (CpsB, C), export (CpsC) and biosynthesis (CpsE, F, G, H, J, K). Although, at first glance, the overall organization seemed to be similar to that of the *cps* and *eps* gene clusters of a number of Gram-positive bacteria (19, 32, 42), overall homologies are low

(*see*, table 3). The region involved in biosynthesis is located at the center of the gene cluster and is flanked by two regions containing genes with more common functions.

[0031] The invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* serotype 2, or a gene or gene fragment derived thereof, preferably as identified in FIG. 3. Genes in this gene cluster are involved in polysaccharide biosynthesis of capsular components and antigens. For a further description of such genes *see*, for example, Table 2. For example, a *cpsA* gene is provided functionally encoding regulation of capsular polysaccharide synthesis, whereas *cpsB* and *cpsC* are functionally involved in chain-in-chain length determination. Other genes, such as *cpsD*, E, F, G, H, I, J, K and related genes, are involved in polysaccharide synthesis, functioning, for example, as glucosyl- or glycosyltransferase. The *cpsF*, G, H, I, J genes encode more type-specific proteins than the flanking genes which are found more-or-less conserved throughout the species and can serve as a base for selection of primers or probes in PCR-amplification or cross-hybridization experiments for subsequent cloning.

[0032] The invention further provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* serotype 1 or a gene or gene fragment derived thereof, preferably as identified in FIG. 4.

[0033] In addition, the invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* serotype 9 or a gene or gene fragment derived thereof, preferably as identified in FIG. 5.

[0034] Furthermore, the invention provides, for example, a fragment of the *cps* locus, or parts thereof, involved in the capsular polysaccharide biosynthesis of *S. suis* exemplified herein for serotypes 1, 2 or 9, and allows easy identification or detection of related fragments derived of other serotypes of *S. suis*.

[0035] The invention provides a nucleic acid probe or primer derived from a nucleic acid according to the invention allowing species or serotype-specific detection of *Streptococcus suis*. Such a probe or primer (used interchangeably herein) is, for example, a DNA, RNA or PNA (peptide nucleic acid) probe hybridizing with capsular nucleic acid as provided by the invention. Species-specific detection is provided preferably by selecting a probe or primer sequence from a species-specific region (*e.g.* flanking region) whereas serotype-specific detection is provided

preferably by selecting a probe or primer sequence from a type-specific region (e.g. central region) of a capsular gene cluster as provided by the invention. Such a probe or primer can be used in a further unmodified form, for example, in cross-hybridization or polymerase-chain reaction (PCR) experiments as, for example described in the experimental part herein. The invention provides the isolation and molecular characterization of additional type-specific *cps* genes of *S. suis* types 1 and 9. In addition, we describe the genetic diversity of the *cps* loci of serotypes 1, 2 and 9 among the 35 *S. suis* serotypes known. Type-specific probes are identified. Also, a type-specific PCR, for example, for serotype 9, is provided, being a rapid, reliable and sensitive assay used directly on nasal or tonsillar swabs or other samples of infected or carrier animals.

[0036] The invention also provides a probe or primer according to the invention with at least one reporter molecule. Examples of reporter molecules are manifold and known in the art; for example, a reporter molecule can include additional nucleic acid provided with a specific sequence (e.g. oligo-dT) hybridizing to a corresponding sequence in which hybridization can easily be detected, for example, because it has been immobilized to a solid support.

[0037] Yet other reporter molecules include chromophores, e.g. fluorochromes for visual detection, for example, by light microscopy or fluorescent *in situ* hybridization (“FISH”) techniques, or include an enzyme such as horseradish peroxidase for enzymatic detection, e.g. in enzyme-linked assays (“EIA”). Yet other reporter molecules include radioactive compounds for detection in radiation-based assays.

[0038] In a preferred embodiment of the invention, at least one probe or primer according to the invention is provided (labeled) with a reporter molecule and a quencher molecule, together with an unlabeled probe or primer in a PCR-based test allowing rapid detection of specific hybridization.

[0039] The invention further provides a diagnostic test or test kit including a probe or primer as provided by the invention. Such a test or test kit is, for example, a cross-hybridization test or PCR-based test advantageously used in rapid detection and/or serotyping of *Streptococcus suis*.

[0040] The invention further provides a protein or fragment thereof encoded by a nucleic acid according to the invention. Examples of such a protein or fragment are proteins described in Table 2. For example, a *cpsA* protein is provided that functionally encodes regulation of capsular

polysaccharide synthesis, whereas *cpsB* and *cpsC* are functionally involved in chain-in-chain length determination. Other proteins or functional fragments thereof, as provided by the invention, such as *cpsD*, *E*, *F*, *G*, *H*, *I*, *J*, *K* and related proteins, are involved in polysaccharide biosynthesis, functioning, for example, as glucosyl- or glycosyltransferase in polysaccharide biosynthesis of *Streptococcus suis* capsular antigen.

[0041] The invention also provides a method of producing a *Streptococcus suis* capsular antigen including using a protein or functional fragment thereof as provided by the invention, and provides therewith a *Streptococcus suis* capsular antigen obtainable by such a method.

[0042] A comparison of the predicted amino acid sequences of the *cps2* genes with sequences found in the databases allowed the assignment of functions to the open reading frames. The central region contains the type-specific glycosyltransferases and the putative polysaccharide polymerase. This region is flanked by two regions encoding for proteins with common functions, such as regulation and transport of polysaccharide across the membrane. Biosynthesis of *Streptococcus* capsular polysaccharide antigen using a protein or functional fragment thereof is advantageously used in chemo-enzymatic synthesis and the development of vaccines which offer protection against serotype-specific *Streptococcal* disease, and is also advantageously used in the synthesis and development of multivalent vaccines against *Streptococcal* infections. Such vaccines elicit anticapsular antibodies which confer protection.

[0043] Furthermore, the invention provides an acapsular *Streptococcus* mutant for use in a vaccine, a vaccine strain derived thereof and a vaccine derived thereof. Surprisingly, and against the grain of common doctrine, the invention provides use of a *Streptococcus* mutant deficient in capsular expression in a vaccine.

[0044] Acapsular *Streptococcus* mutants have long been known in the art and can be found in nature. Griffith (*J. Hyg.* 27:113-159, 1928) demonstrated that pneumococci could be transformed from one type to another. If he injected live rough (acapsular or unencapsulated) type 2 pneumococci into mice, the mice would survive. If, however, he injected the same dose of live rough type 2 mixed with heat-killed smooth (encapsulated) type 1 into a mouse, the mouse would die, and, from the blood, he could isolate live smooth type 1 pneumococci. At that time, the significance of this transforming principle was not understood. However, understanding came when

it was shown that DNA constituted the genetic material responsible for phenotypic changes during transformation.

[0045] *Streptococcus* mutants deficient in capsular expression are found in several forms. Some are fully deficient and have no capsule at all, others form a deficient capsule, characterized by a mutation in a capsular gene cluster. Deficiency can, for instance, include capsular formation wherein the organization of the capsular material has been rearranged as, for example, demonstrable by electron microscopy. Yet others have a nearly fully developed capsule which is only deficient in a particular sugar component.

[0046] Now, after much advance of biotechnology and despite the fact that little is still known about the exact localization and sequence of genes involved in capsular synthesis in Streptococci, it is possible to create mutants of Streptococci, for example, by homologous recombination or transposon mutagenesis, which has, for example, been done for GAS (Wessels et al., *PNAS* 88:8317-8321, 1991), for GBS (Wessels et al., *PNAS* 86: 8983-8987, 1989), for *S. suis* (Smith, ID-DLO Annual report 1996, page 18-19; Charland et al., *Microbiol.* 144:325-332, 1998) and *S. pneumoniae* (Kolkman et al., *J. Bact.* 178:3736-3741, 1996). Such recombinant derived mutants, or isogenic mutants, can easily be compared with the wild-type strains from which they have been derived.

[0047] In a preferred embodiment, the invention provides use of a recombinant-derived *Streptococcus* mutant deficient in capsular expression in a vaccine. Recombinant techniques useful in producing such mutants are, for example, homologous recombination, transposon mutagenesis, and others, wherein deletions, insertions or (point) mutations are introduced in the genome. Advantages of using recombinant techniques include the stability of the obtained mutants (especially with homologous recombination and double cross-over techniques), and the knowledge about the exact site of the deletion, mutation or insertion.

[0048] In another embodiment, the invention provides a stable mutant deficient in capsular expression obtained, for example, through homologous recombination or cross-over integration events. Examples of such a mutant can be found herein, such as mutants 10cpsB or 10cpsEF are stable mutants as provided by the invention.

[0049] The invention also provides a *Streptococcus* vaccine strain and vaccine that has been derived from a *Streptococcus* mutant deficient in capsular expression. In general, the strain or vaccine is applicable within the whole range of Streptococcal infections, including animals or man or with zoonotic infections. It is, of course, now possible to first select a common vaccine strain and derive a *Streptococcus* mutant deficient in capsular expression thereof for the selection of a vaccine strain and use in a vaccine according to the invention.

[0050] In a preferred embodiment, the invention provides use of a *Streptococcus* mutant deficient in capsular expression in a vaccine wherein the *Streptococcus* mutant is selected from the group composed of *Streptococcus* group A, *Streptococcus* group B, *Streptococcus suis* and *Streptococcus pneumoniae*. Herewith the invention provides vaccine strains and vaccines for use with these notoriously heterologous Streptococci, of which a multitude of serotypes exist. With a vaccine, as provided by the invention, that is derived from a specific *Streptococcus* mutant that is deficient in capsular expression, the difficulties relating to lack of heterologous protection can be circumvented since these mutants do not rely on capsular antigens, *per se*, to induce protection.

[0051] In a preferred embodiment, the vaccine strain is selected for its ability to survive, or even replicate, in an immune-competent host or host cells and thus can persist for a certain period, varying from 1-2 days to more than one or two weeks, in a host, despite its deficient character.

[0052] Although an immunodeficient host will support replication of a wide range of bacteria that are deficient in one or more virulence factors, in general, it is considered a characteristic of pathogenicity of Streptococci that they can survive for certain periods or replicate in a normal host or host cells such as macrophages. For example, Williams and Blakemore (*Neuropath. Appl. Neurobiol.* 16, 345-356, 1990; *Neuropath. Appl. Neurobiol.* 16, 377-392, 1990; *J. Infect. Dis.* 162, 474-481, 1990) show that both polymorphonuclear cells and macrophage cells are capable of phagocytosing pathogenic *S. suis* in pigs lacking anti-*S. suis* antibodies; only pathogenic bacteria could survive and multiply inside macrophages and the pig.

[0053] In a preferred embodiment, the invention, however, provides a deficient or avirulent mutant or vaccine strain which is capable of surviving at least 4-5 days, preferably at least 8-10 days in the host, thereby allowing the development of a solid immune response to subsequent *Streptococcus* infection,

[0054] Due to its persistent but avirulent character, a *Streptococcus* mutant or vaccine strain, as provided by the invention, is well suited to generate specific and/or long-lasting immune responses against Streptococcal antigens. Moreover, possible specific immune responses of the host directed against a capsule are relatively irrelevant because a vaccine strain, as provided by the invention, is typically not recognized by such antibodies.

[0055] In addition, the invention provides a *Streptococcus* vaccine strain, according to the invention, which strain includes a mutant capable of expressing a *Streptococcus* virulence factor or antigenic determinant.

[0056] In a preferred embodiment, the invention provides a *Streptococcus* vaccine strain, according to the invention, which includes a mutant capable of expressing a *Streptococcus* virulence factor wherein the virulence factor or antigenic determinant is selected from a group of cellular components, such as muramidase-released protein (“MRP”), extracellular factor (“EF”), and cell-membrane associated proteins, 60kDa heat shock protein, pneumococcal surface protein A (Psp A), pneumolysin, C protein, protein M, fimbriae, hemagglutinins and hemolysis or components functionally related thereto.

[0057] In a preferred embodiment, the invention provides a *Streptococcus* vaccine strain including a mutant capable of over-expressing the virulence factor. In this way, the invention provides a vaccine strain for incorporation in a vaccine which specifically causes a host immune response directed against antigenically important determinants of virulence (listed above), thereby providing specific protection against the determinants. Over-expression can, for example, be achieved by cloning the gene involved behind a strong promoter, which is, for example, constitutionally expressed in a multicopy system, either in a plasmid or via integration in a genome.

[0058] In yet another embodiment, the invention provides a *Streptococcus* vaccine strain, according to the invention, including a mutant capable of expressing a non-*Streptococcus* protein. Such a vector-*Streptococcus* vaccine strain allows, when used in a vaccine, protection against pathogens other than *Streptococcus*.

[0059] Due to its persistent but avirulent character, a *Streptococcus* vaccine strain or mutant as provided by the invention is well suited to generate specific and long-lasting immune responses, not only against Streptococcal antigens, but also against other antigens expressed by the

strain. Specifically, antigens derived from another pathogen are now expressed without the detrimental effects of the antigen or pathogen which would otherwise have harmed the host.

[0060] An example of such a vector is a *Streptococcus* vaccine strain or mutant wherein the antigen is derived from a pathogen, such as *Actinobacillus pleuropneumonia*, *Mycoplasmatae*, *Bordetella*, *pasteurella*, *E. coli*, *Salmonella*, *campylobacter*, *Serpulina* and others.

[0061] The invention also provides a vaccine including a *Streptococcus* vaccine strain or mutant according to the invention and a pharmaceutically acceptable carrier or adjuvant. Carriers or adjuvants are well known in the art; examples are phosphate buffered saline, physiological salt solutions, (double-) oil-in-water emulsions, aluminumhydroxide, Specol, block- or co-polymers, and others.

[0062] A vaccine according to the invention can include a vaccine strain either in a killed or live form. For example, a killed vaccine including a strain having (over)expressed a Streptococcal or heterologous antigen or virulence factor is very well suited for eliciting an immune response. In a preferred embodiment, the invention provides a vaccine wherein the strain is live, due to its persistent but avirulent character; a *Streptococcus* vaccine strain, as provided by the invention, is well suited to generate specific and long-lasting immune responses.

[0063] The invention also provides a method for controlling or eradicating a Streptococcal disease in a population comprising vaccinating subjects in the population with a vaccine according to the invention.

[0064] In a preferred embodiment, a method for controlling or eradicating a Streptococcal disease is provided including testing a sample, such as a blood sample, or nasal or throat swab, feces, urine, or other samples such as can be sampled at or after slaughter, collected from at least-one subject, such as an infant or a pig, in a population partly or wholly vaccinated with a vaccine according to the invention for the presence of encapsulated Streptococcal strains or mutants. Since a vaccine strain or mutant according to the invention is not pathogenic, and can be distinguished from wild-type strains by capsular expression, the detection of (fully) encapsulated Streptococcal strains indicates that wild-type infections are still present. Such wild-type infected subjects can then be isolated from the remainder of the population until the infection has passed. With domestic animals, such as pigs, it is even possible to remove the infected subject from the population as a whole by

culling. Detection of wild-type strains can be achieved via traditional culturing techniques, or by rapid detection techniques such as PCR detection.

[0065] In yet another embodiment, the invention provides a method for controlling or eradicating a Streptococcal disease including testing a sample collected from at least one subject in a population partly or wholly vaccinated with a vaccine according to the invention for the presence of capsule-specific antibodies directed against Streptococcal strains. Capsule-specific antibodies can be detected with classical techniques known in the art, such as used for Lancefield's group typing or serotyping.

[0066] A preferred embodiment for controlling or eradicating a Streptococcal disease in a population includes vaccinating subjects in the population with a vaccine according to the invention and testing a sample collected from at least one subject in the population for the presence of encapsulated Streptococcal strains and/or for the presence of capsule-specific antibodies directed against Streptococcal strains.

[0067] For example, a method is provided wherein the Streptococcal disease is caused by *Streptococcus suis*.

[0068] The invention also provides a diagnostic assay for testing a sample for use in a method according to the invention including at least one means for the detection of encapsulated Streptococcal strains and/or for the detection of capsule-specific antibodies directed against Streptococcal strains.

[0069] The invention further provides a vaccine including an antigen according to the invention and a suitable carrier or adjuvant. The immunogenicity of a capsular antigen provided by the invention is, for example, increased by linking to a carrier (such as a carrier protein), allowing the recruitment of T-cell help in developing an immune response.

[0070] The invention further provides a recombinant microorganism provided with at least a part of a capsular gene cluster derived from *Streptococcus suis*. The invention provides for example a lactic acid bacterium provided with at least a part of a capsular gene cluster derived from *Streptococcus suis*. Various food-grade lactic acid bacteria (*Lactococcus lactis*, *Lactobacillus casei*, *Lactobacillus plantarum* and *Streptococcus gordonii*) have been used as delivery systems for mucosal immunization. It has now been shown that oral (or mucosal) administration of recombinant

L. lactis, *Lactobacillus*, and *Streptococcus gordonii* can elicit local IgA and/or IgG antibody responses to an expressed antigen. The use of oral routes for immunization against infective diseases is desirable because oral vaccines are easier to administer and have higher compliance rates, and because mucosal surfaces are the portals of entry for many pathogenic microbial agents. It is within the skill of the artisan to provide such microorganisms with (additional) genes.

[0071] The invention further provides a recombinant *Streptococcus suis* mutant provided with a modified capsular gene cluster. It is within the skill of the artisan to swap genes within a species. In a preferred embodiment, an avirulent *Streptococcus suis* mutant is selected to be provided with at least a part of a modified capsular gene cluster according to the invention.

[0072] The invention further provides a vaccine including a microorganism or a mutant provided by the invention. An advantage of such a vaccine over currently used vaccines is that they include accurately defined microorganisms and well-characterized antigens, allowing accurate determination of immune responses against various antigens of choice.

[0073] The invention is further explained in the experimental part of this description without limiting the invention thereto.

DESCRIPTION OF THE DRAWINGS

[0074] FIG. 1 illustrates the organization of the cps2 gene cluster of *S. suis* type 2.

[0075] (A) Genetic map of the cps2 gene cluster. The shadowed arrows represent potential ORFs. Interrupted ORFs indicate the presence of stop codons or frame-shift mutations. Gene designations are indicated below the ORFs. The closed arrows indicate the position of the potential promoter sequences. | indicates the position of the potential transcription regulator sequence. ||| indicates the position of the 100-bp repeated sequence.

[0076] (B) Physical map of the cps2 locus. Restriction sites are as follows: A: *Alu*I; C: *Clal*; E: *Eco*RI; H: *Hind*III; K: *Kpn*I; M: *Mlu*I; N: *Nsi*I; P: *Pst*I; S: *Sna*BI; Sa: *Sac*I; X: *Xba*I.

[0077] (C) The DNA fragments cloned in the various plasmids.

[0078] FIG. 2 illustrates ethidium bromide stained agarose gel showing PCR products obtained with chromosomal DNA of *S. suis* strains belonging to the serotypes 1,2, 1/2, 9 and 14 and *cps2J*, *cps1I*, and *cps9H* primer sets as described herein.

[0079] (A) *cpsII* primers; (B) *cps2J* primers and (C) *cps9H* primers.

[0080] Lanes 1-3: serotype 1 strains; lanes 4-6: serotype 2 strains; lanes 7-9: serotype ½ strains; lanes 10-12: serotype 9 strains and lanes 13-15: serotype 14 strains.

[0081] (B) Ethidium bromide stained agarose gel showing PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 2, type 1 or type 9 strains and *cps2J*, *cpsII* and *cpsH* primer sets as described in Materials and Methods. Bacterial DNA suitable for PCR was prepared by using the multiscreen methods as described previously (20).

[0082] (C) *cpsII* primers. (B) *cps2J* primers and (C) *cps9H* primers.

[0083] Lanes 1-3: PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 1 strains; lanes 4-6: PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 2 strains; lanes 7-9: PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 9 strains; lanes 10-12: PCR products obtained with chromosomal DNA from serotype 9, 2 and 1 strains respectively; lane 13: negative control, no DNA present.

[0084] FIG. 3 illustrates the CPS2 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0085] FIG. 4 illustrates the CPS1 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0086] FIG. 5 illustrates the CPS9 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0087] FIG. 6 illustrates the CPS7 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0088] FIG. 7 illustrates alignment of the N-terminal parts of Cps2J and Cps2K.

[0089] Identical amino acids are marked by bars. The amino acids shown in bold are also conserved in Cps14I, Cps24J of *S. pneumoniae* and several other glycosyltransferases (19). The aspartate residues marked by asterisks are strongly conserved.

[0090] FIG. 8 illustrates transmission electron micrographs of thin sections of various *S. suis* strains.

[0091] (A) wild-type strain 10;

[0092] (B) mutant strain 10cpsB;

[0093] (C) mutant strain 10cpsEF.

[0094] Bar = 100 nm

[0095] FIG. 9 illustrates the kinetics of phagocytosis of wild-type and mutant *S. suis* strains.

[0096] (A) Kinetics of phagocytosis of wild-type and mutant *S. suis* strains by porcine alveolar macrophages. Phagocytosis was determined as described herein. The Y-axis represents the number of CFU per milliliter in the supernatant fluids as determined by plate counting, the X-axis represents time in minutes.

[0097] □ wild-type strain 10;

[0098] ○ mutant strain 10cpsB;

[0099] Δ mutant strain 10cpsEF.

[00100] (B) Kinetics of intracellular killing of wild-type and mutant *S. suis* strains by porcine AM. The intracellular killing was determined as described herein. The Y-axis represents the number of CFU per ml in the supernatant fluids after lysis of the macrophages as determined by plate counting, the X-axis represents time in minutes.

[00101] □ wild-type strain 10;

[00102] ○ mutant strain 10cpsB;

[00103] Δ mutant strain 10cpsEF.

[00104] FIG. 10 illustrates the nucleotide sequence alignment of the highly conserved 100-bp repeated element.

[00105] 1) 100-bp repeat between cps2G and cps2H

[00106] 2) 100-bp repeat within “cps2M”

[00107] 3) 100-bp repeat between cps20 and cps2P

[00108] FIG. 11 illustrates the cps2, cps9 and cps7 gene clusters of *S. suis* serotypes 2, 9 and 7.

[00109] (A) Genetic organization of the cps2 gene cluster [84]. The large arrows represent potential ORFs. Gene designations are indicated below the ORFs. Identically filled arrows represent ORFs which showed homology. The small closed arrows indicate the position of the potential promoter sequences. | indicates the position of the potential transcription regulator sequence.

[00110] (B) Physical map and genetic organization of the *cps9* gene cluster [15]. Restriction sites are as follows: B: *Bam*HI; P: *Pst*I; H: *Hind*III; X: *Xba*I. The DNA fragments cloned in the various plasmids are indicated. The open arrows represent potential ORFs.

[00111] (C) Physical map and genetic organization of the *cps7* gene cluster. Restriction sites are as follows: C: *Clal*; P: *Pst*I; Sc: *Scal*. The DNA fragments cloned in the various plasmids are indicated. The open arrows represent potential ORFs.

[00112] FIG. 12 illustrates ethidium bromide stained agarose gel showing PCR products.

[00113] (A) Ethidium bromide stained agarose gel showing PCR products obtained with chromosomal DNA of *S. suis* strains belonging to the serotypes 1, 2, 9 and 7 and the *cps7H* primer set. Strain designations are indicated above the lanes. C: negative control, no DNA present. M: molecular size marker (lambda digested with *Eco*RI and *Hind*III).

[00114] (B) Ethidium bromide stained agarose gel showing PCR products obtained with serotype 7 strains collected in different countries and from different organs. Bacterial DNA suitable for PCR was prepared by using the multiscreen method as described herein [89]. Strain designations are indicated above the lanes. M: molecular size marker (lambda digested with *Eco*RI and *Hind*III).

DETAILED DESCRIPTION OF THE INVENTION

Experimental part

MATERIAL AND METHODS

Bacterial strains and growth conditions.

[00115] The bacterial strains and plasmids used in this study are listed in Table 1. *S. suis* strains were grown in Todd-Hewitt broth (code CM189, Oxoid), and plated on Columbia agar blood base (code CM331, Oxoid) containing 6% (v/v) horse blood. *E.coli* strains were grown in Luria broth (28) and plated on Luria broth containing 1.5% (w/v) agar. If required, antibiotics were added to the plates at the following concentrations: spectinomycin: 100 µg/ml for *S. suis* and 50 µg/ml for *E. coli* and ampicillin, 50 µg/ml.

[00116] **Serotyping.** The *S. suis* strains were serotyped by the slide agglutination test with serotype-specific antibodies (44).

[00117] DNA techniques. Routine DNA manipulations were performed as described by Sambrook et al. (36).

[00118] Alkaline phosphatase activity. To screen for PhoA fusions in *E. coli*, plasmid libraries were constructed. Therefore, chromosomal DNA of *S. suis* type 2 was digested with *Alu*I. The 300-500-bp fragments were ligated to *Sma*I-digested pPHOS2. Ligation mixtures were transformed to the PhoA⁻ *E. coli* strain CC118. Transformants were plated on LB media supplemented with 5- Bromo-4-chloro-3-indolylfosfaat (BCIP, 50 µg/ml, Boehringer, Mannheim, Germany). Blue colonies were purified on fresh LB/BCIP plates to verify the blue phenotype.

[00119] DNA sequence analysis. DNA sequences were determined on a 373A DNA Sequencing System (Applied Biosystems, Warrington, GB). Samples were prepared by using an ABI/PRISM dye terminator cycle sequencing ready reaction kit (Applied Biosystems). Sequencing data were assembled and analyzed using the MacMollyTetra program. Custom-made sequencing primers were purchased from Life Technologies. Hydrophobic stretches within proteins were predicted by the method of Klein et al. (17). The BLAST program available on Netscape NavigatorTM was used to search for protein sequences related to the deduced amino acid sequences.

[00120] Construction of gene-specific knock-out mutants of *S. suis*. To construct the mutant strains 10cpsB and 10cpsEF, we electrotransformed the pathogenic serotype 2 strain 10 (45, 49) of *S. suis* with pCPS11 and pCPS28 respectively. In these plasmids, the *cpsB* and *cpsEF* genes were disturbed by the insertion of a spectinomycin-resistance gene. To create pCPS11, the internal 400 bp *Pst*I-*Bam*HI fragment of the *cpsB* gene in pCPS7 was replaced by the Spc^R gene. For this purpose, pCPS7 was digested with *Pst*I and *Bam*HI and ligated to the 1,200-bp *Pst*I-*Bam*HI fragment, containing the Spc^R gene, from pIC-spc. To construct pCPS28, we have used pIC20R. In this plasmid we inserted the *Kpn*I-*Sa*II fragment from pCPS17 (resulting in pCPS25) and the *Xba*I-*Cla*I fragment from pCPS20 (resulting in pCPS27). pCPS27 was digested with *Pst*I and *Xho*I and ligated to the 1,200-bp *Pst*I-*Xho*I fragment, containing the Spc^R gene of pIC-spc. The electrotransformation to *S. suis* was carried out as described before (38).

[00121] Southern blotting and hybridization. Chromosomal DNA was isolated as described by Sambrook et al. (36). DNA fragments were separated on 0.8% agarose gels and transferred to Zeta-Probe GT membranes (Bio-Rad) as described by Sambrook et al. (36). DNA

probes were labeled with [$\text{-}^{32}\text{P}$]dCTP (3000 Ci mmol $^{-1}$ Amersham) by use of a random primed labeling kit (Boehringer). The DNA on the blots was hybridized at 65°C with appropriate DNA probes as recommended by the supplier of the Zeta-Probe membranes. After hybridization, the membranes were washed twice with a solution of 40 mM sodium phosphate, pH 7.2, 1 mM EDTA, 5% SDS for 30 min at 65°C and twice with a solution of 40 mM sodium phosphate, pH 7.2, 1 mM EDTA, 1% SDS for 30 min at 65°C.

[00122] PCR. The primers used in the *cps2*JPCR correspond to the positions 13791-13813 and 14465-14443 in the *S. suis* *cps2* locus. The sequences were: 5'-CAAACGCAAGGAATTACGGTATC-3' (SEQ ID NO:1) and 5'-GAGTATCTAAAGAATGCCTATTG-3' (SEQ ID NO:2). The primers used for the *cps1*IPCR correspond to the positions 4398-4417 and 4839-4821 in the *S. suis* *cps1* sequence. The sequences were: 5'-GGCGGTCTAGCAGATGCTCG-3' (SEQ ID NO:3) and 5'-GCGAACTGTTAGCAATGAC-3' (SEQ ID NO:4). The primers used in the *cps9*H PCR correspond to the positions 4406-4126 and 4494-4475 in the *S. suis* *cps9* sequence. The sequences were: 5'-GGCTACATATAATGGAAGCCC3' (SEQ ID NO:5) and 5'-CGGAAGTATCTGGGCTACTG-3' (SEQ ID NO:6).

[00123] Construction of gene-specific knock-out mutants of *S. suis*. To construct the mutant strains 10cpsB and 10cpsEF, we electrotransformed the pathogenic serotype 2 strain 10 of *S. suis* with pCPS11 and pCPS28 respectively. In these plasmids, the *cpsB* and *cpsEF* genes were disturbed by the insertion of a spectinomycin-resistance gene. To create pCPS11, the internal 400 bp *Pst*I-*Bam*HI fragment of the *cpsB* gene in pCPS7 was replaced by the *Spc*^R gene. For this purpose, pCPS7 was digested with *Pst*I and *Bam*HI and ligated to the 1,200-bp *Pst*I-*Bam*HI fragment, containing the *Spc*^R gene, from pIC-spc. To construct pCPS28, we have used pIC20R. In this plasmid, we inserted the *Kpn*I-*Sal*I fragment from pCPS17 (resulting in pCPS25) and the *Xba*I-*Cl*I fragment from pCPS20 (resulting in pCPS27). pCPS27 was digested with *Pst*I and *Xho*I and ligated to the 1,200-bp *Pst*I-*Xho*I fragment, containing the *spc*^R gene of pIC-Spc. The electrotransformation to *S. suis* was carried out as described before (38).

[00124] Phagocytosis assay. Phagocytosis assays were performed as described by Leij et al. (23). Briefly, to opsonize the cells, 10^7 *S. suis* cells were incubated with 6% SPF-pig serum for 30

min at 37°C in a head-over-head rotor at 6 rpm. 10^7 AM and 10^7 opsonized *S. suis* cells were combined and incubated at 37°C under continuous rotation at 6 rpm. At 0, 30, 60 and 90 min, 1-ml samples were collected and mixed with 4 ml of ice-cold EMEM to stop phagocytosis. Phagocytes were removed by centrifugation for 4 min at 110 x g and 4°C. The number of colony-forming units, ("CFU") in the supernatants was determined. Control experiments were carried out simultaneously by combining 10^7 opsonized *S. suis* cells with EMEM (without AM).

[00125] Killing assays. AM (10^7 /ml) and opsonized *S. suis* cells (10^7 /ml) were mixed 1 : 1 and incubated for 10 min at 37°C under continuous rotation at 6 rpm. Ice-cold EMEM was added to stop further phagocytosis and killing. To remove extracellular *S. suis* cells, phagocytes were washed twice (4 min, 110 x g, 4°C) and resuspended in 5 ml EMEM containing 6% SPF serum. The tubes were incubated at 37°C under rotation at 6 rpm. After 0, 15, 30, 60 and 90 min, samples were collected and mixed with ice-cold EMEM to stop further killing. The samples were centrifuged for 4 min at 110 x g at 4°C and the phagocytic cells were lysed in EMEM containing 1% saponine for 20 min at room temperature. The number of CFU in the suspensions was determined.

[00126] Pigs. Germfree pigs, crossbreeds of Great Yorkshire and Dutch Landrace, were obtained from sows by caesarian sections. The surgery was performed in sterile flexible film isolators. Pigs were allotted to groups, each consisting of 4 pigs, and were housed in sterile stainless steel incubators.

[00127] Experimental infections. Pigs were inoculated intranasally with *S. suis* type 2 as described before. To predispose the pigs for infection with *S. suis*, five-day old pigs were inoculated intranasally with about 10^7 CFU of *Bordetella bronchiseptica* strain 92932. Two days later, the pigs were inoculated intranasally with *S. suis* type 2 (10^6 CFU). Pigs were monitored twice daily for clinical signs of disease, such as fever, nervous signs and lameness. Blood samples were collected three times a week from each pig. White blood cells were counted with a cell counter. To monitor infection with *S. suis* and *B. bronchiseptica* and to check for absence of contaminants, we collected swabs of nasopharynx and feces daily. The swabs were plated directly onto Columbia agar containing 6% horse blood. After three weeks, the pigs were killed and examined for pathological changes. Tissue specimens from the central nervous system, serosae, and joints were examined bacteriologically and histologically as described herein (45, 49). Colonization of the serosae was

scored positively when *S. suis* was isolated from the pericardium, thoracal pleura or the peritoneum. Colonization of the joints was scored positively when *S. suis* was isolated from one or more joints (12 joints per animal were scored).

[00128] Vaccination and challenge. One week old pigs were vaccinated intravenously with a dosage of 106 cfu of the *S. suis* strains 10cpsEF or 10cpsB. Three weeks later, the pigs were challenged intravenously with the pathogenic serotype 2 strain 10 (107 cfu). Disease monitoring, hematological, serological and bacteriological examinations as well as post-mortum examinations were as described before under experimental infections.

[00129] Electron Microscopy. Bacteria were prepared for electron microscopy as described by Wagenaar et al. (50). Shortly, bacteria were mixed with agarose MP (Boehringer) of 37°C to a concentration of 0.7%. The mixture was immediately cooled on ice. Upon gelyfying, samples were cut into 1 to 1.5 mm slices and incubated in a fixative containing 0.8% glutaraldehyde and 0.8% osmiumtetraoxide. Subsequently, the samples were fixed and stained with uranyl acetate by microwave stimulation, dehydrated and imbedded in eponaraldite resin. Ultra-thin sections were counterstained with lead citrate and examined with a Philips CM 10 electron microscope at 80 kV. (FIG. 8.)

[00130] Isolation of porcine alveolar macrophages (AM). Porcine AM were obtained from the lungs of specific pathogen-free (“SPF”) pigs. Lung lavage samples were collected as described by van Leengoed et al. (43). Cells were suspended in EMEM containing 6% (v/v). SPF-pig serum and adjusted to 10 cells per ml.

RESULTS

Identification of the *cps* locus.

[00131] The *cps* locus of *S. suis* type 2 was identified through a strategy developed for the genetic identification of exported proteins (13, 31). In this system, a plasmid (pPHOS2) containing a truncated alkaline phosphatase gene (13) was used. The gene lacked the promoter sequence, the translational start site and the signal sequence. The truncated gene is preceded by a unique *Sma*I restriction site. Chromosomal DNA of *S. suis* type 2, digested with *Alu*I, was randomly cloned in this restriction site. Because translocation of PhoA across the cytoplasmic membrane of *E. coli* is

required for enzymatic activity, the system can be used to select for *S. suis* fragments containing a promoter sequence, a translational start site and a functional signal sequence. Among 560 individual *E. coli* clones tested, 16 displayed a dark blue phenotype when plated on media containing BCIP. DNA sequence analysis of the inserts from several of these plasmids was performed (results not shown) and the deduced amino acid sequences were analyzed. The hydrophobicity profile of one of the clones (pPHOS7, results not shown) showed that the N-terminal part of the sequence resembled the characteristics of a typical signal peptide: a short hydrophilic N-terminal region is followed by a hydrophobic region of 38 amino acids. These data indicate that the phoA system was successfully used for the selection of *S. suis* genes encoding exported proteins. Moreover, the sequences were analyzed for similarities present in the databases. The sequence of pPHOS7 showed a high similarity (37% identity) with the protein encoded by the *cps14C* gene of *Streptococcus pneumoniae* (19). This strongly suggests that pPHOS7 contains a part of the *cps* operon of *S. suis* type 2.

[00132] Cloning of the flanking *cps* genes. In order to clone the flanking *cps* genes of *S. suis* type 2, the insert of pPHOS7 was used as a probe to identify chromosomal DNA fragments which contain flanking *cps* genes. A 6-kb *Hind*III fragment was identified and cloned in pKUN19. This yielded clone pCPS6 (FIG. 1, part C). Sequence analysis of the insert of pCPS6 revealed that pCPS6 most probably contained the 5'-end of the *cps* locus, but still lacked the 3'-end. Therefore, sequences of the 3'-end of pCPS6 were in turn used as a probe to identify chromosomal fragments containing *cps* sequences located further downstream. These fragments were also cloned in pKUN19, resulting in pCPS17. Using the same system of chromosomal walking, plasmids pCPS18, pCPS20, pCPS23 and pCPS26, containing downstream *cps* sequences were subsequently generated.

[00133] Analysis of the *cps* operon. The complete nucleotide sequence of the cloned fragments was determined (FIG. 4). Examination of the compiled sequence revealed the presence of at least 13 potential open reading frames (Orfs), which were designated as Orf 2Y, Orf2X and Cps2A-Cps2K (FIG. 1, part A; FIG. 11, part A). Moreover, a 14th, incomplete Orf (Orf 2Z) was located at the 5'-end of the sequence. Two potential promoter sequences were identified. One was located 313 bp (locations 1885-1865 and 1884-1889) upstream of Orf2X. The other potential promoter sequence was located 68 bp upstream of Orf2Y (locations 2241-2236 and 2216-2211). Orf2Y is expressed in opposite orientation. Between Orfs 2Y and 2Z, the sequence contained a

potential stem-loop structure, which could act as a transcription terminator. Each Orf is preceded by a ribosome-binding site and the majority of the Orfs are very closely linked. The only significant intergenic gap was found between Cps2G and Cps2H (389 nucleotides). However, no obvious promoter sequences or potential stem-loop structures were found in this region. These data suggest that Orf2X and Cps2A-Cps2K are arranged as an operon.

[00134] An overview of all Orfs with their properties is shown in Table 2. The majority of the predicted gene products is related to proteins involved in polysaccharide biosynthesis. Orf2Z showed some similarity with the YitS protein of *Bacillus subtilis*. YitS was identified during the sequence analysis of the complete genome of *B. subtilis*. The function of the protein is unknown.

[00135] Orf2Y showed similarity with the YcxD protein of *B. subtilis* (53). Based on the similarity between YcxD and MocR of *Rhizobium meliloti* (33), YcxD was suggested to be a regulatory protein.

[00136] Orf2X showed similarity with the hypothetical YAAA proteins of *Haemophilus influenzae* and *E. coli*. The function of these proteins is unknown.

[00137] The gene products encoded by the *cps2A*, *cps2B*, *cps2C* and *cps2D* genes showed approximate similarity to the CpsA, CpsC, CpsD and CpsB proteins of several serotypes of *Streptococcus pneumoniae* (19), respectively. This suggests similar functions for these proteins. Hence, Cps2A may have a role in the regulation of the capsular polysaccharide synthesis. Cps2B and Cps2C could be involved in the chain length determination of the type 2 capsule and Cps2C can play an additional role in the export of the polysaccharide. The Cps2D protein of *S. suis* is related to the CpsB protein of *S. pneumoniae* and to proteins encoded by genes of several other Gram-positive bacteria involved in polysaccharide or exopolysaccharide synthesis, but their function is unknown (19).

[00138] The protein encoded by the *cps2E* gene showed similarity to several bacterial proteins with glycosyltransferase activities: Cps14E and Cps19fE of *S. pneumoniae* serotypes 14 and 19F (18, 19, 29), CpsE of *Streptococcus salvarius* (X94980) and CpsD of *Streptococcus agalactiae* (34). Recently, Kolkman et al. (18) showed that Cps14E is a glucosyl-l-phosphate transferase that links glucose to a lipid carrier, the first step in the biosynthesis of the *S. pneumoniae* type 14 repeating unit. Based on these data, a similar function may be fulfilled by Cps2E of *S. suis*.

[00139] The protein encoded by the *cps2F* gene showed similarity to the protein encoded by the *rfbU* gene of *Salmonella enteritica* (25). This similarity is most pronounced in the C-terminal regions of these proteins. The *rfbU* gene was shown to encode mannosyltransferase activity (25).

[00140] The *cps2G* gene encoded a protein that showed moderate similarity with the *rfbF* gene product of *Campylobacter hyoilei* (22), the *epsF* gene product of *S. thermophilus* (40) and the *capM* gene product of *S. aureus* (24). On the basis of similarity, the *rfbF*, *epsF* and *capM* genes are suggested to encode galactosyltransferase activities. Hence, a similar glycosyltransferase activity could be fulfilled by the *cps2G* gene product.

[00141] The *cps2H* gene encodes a protein that is similar to the N-terminal region of the *lgtD* gene product of *Haemophilus influenzae* (U32768). Moreover, the hydrophobicity plots of Cps2H and LgtD looked very similar in these regions (data not shown). Based on sequence similarity, the *lgtD* gene product was suggested to have glycosyltransferase activity (U32768).

[00142] The gene product encoded by the *cps2I* gene showed some similarity with a protein of *Actinobacillus actinomycetemcomitans* (AB002668). This protein is part of the gene cluster responsible for the serotype-b-specific antigen of *A. actinomycetemcomitans*. The function of the protein is unknown.

[00143] The gene products encoded by the *cps2J* and *cps2K* genes showed significant similarities to the Cps14J protein of *S. pneumoniae*. The *cps14J* gene of *S. pneumoniae* was shown to encode a β -1,4-galactosyltransferase activity. In *S. pneumoniae*, CpsJ is responsible for the addition of the fourth (*i.e.* last) sugar in the synthesis of the *S. pneumoniae* serotype 14 polysaccharide (20). Even some similarity was found between Cps2J and Cps2K (FIG. 2, 25.5% similarity). This similarity was most pronounced in the N-terminal regions of the proteins (FIG. 7). Recently, two small conserved regions were identified in the N-terminus of Cps14J and Cps14I and their homologues (20). These regions were predicted to be important for catalytic activity. Both regions, DXS and DXDD (FIG. 2), were also found in Cps2J and Cps2K.

[00144] **Distribution of the *cps2* genes in other *S. suis* serotypes.** To examine the relationship between the *cps2* genes and *cps* genes in the other *S. suis* serotypes, we performed cross-hybridization experiments. DNA fragments of the individual *cps2* genes were amplified by PCR, labeled with ^{32}P , and used to probe Southern blots of chromosomal DNA of the reference

strains of the 35 different *S. suis* serotypes. Large variations in the hybridization patterns were observed (Table 4). As a positive control, we used a probe specific for 16S rRNA. The 16S rRNA probe hybridized with all serotypes tested. However, none of the other genes tested were common in all serotypes. Based on the genetic organization of the genes, it was previously suggested that *orfX* and *cpsA-cpsK* genes are part of one operon and that the proteins encoded by these genes are all involved in polysaccharide biosynthesis. OrfY and OrfZ are not a part of this operon, and their role in the polysaccharide biosynthesis is unclear. Based on sequence similarity data, OrfY may be involved in regulation of the *cps2* genes. OrfZ is proposed to be unrelated to polysaccharide biosynthesis. Probes specific for the *orfZ*, *orfY*, *orfX*, *cpsA*, *cpsB*, *cpsC* and *cpsD* genes hybridized with most other serotypes. This suggests that the proteins encoded by these genes are not type-specific, but may perform more common functions in biosynthesis of the capsular polysaccharide. This confirms previous data which showed that the *cps2A-cps2D* genes showed strong similarity to *cps* genes of several serotypes of *Streptococcus pneumoniae*. Based on this similarity, Cps2A is possibly a regulatory protein, whereas Cps2B and Cps2C may play a role in length determination and export of polysaccharide. The *cps2E* gene hybridized with DNA of serotypes 1, 2, 14 and 1/2. The *cps2E* gene showed a strong similarity to the *cps14E* gene of *S. pneumoniae* (18). This enzyme was shown to have a glucosyl-1-phosphate activity and catalyzed the transfer of glucose to a lipid carrier (18). These data indicate that a glycosyltransferase closely related to Cps14E may be responsible for the first step in the biosynthesis of polysaccharide in the *S. suis* serotypes 1, 2, 14 and 1/2. The *cps2F*, *cps2G*, *cps2H*, *cps2I* and *cps2J* genes hybridized with chromosomal DNA of serotypes 2 and 1/2 only. The *cps2G* gene showed an additional weak hybridization signal with DNA of serotype 34. In agglutination tests, serotype 1/2 showed agglutination with sera specific for serotype 2 as well as with sera specific for serotype 1. This suggests that serotype 1/2 shares antigenic determinants with both types 1 and 2. The hybridization data confirmed these data. All putative glycosyltransferases present in serotype 2 are also present in serotype 1/2. The *cps2K* gene showed a hybridization pattern similar to the *cps2E* gene. Hybridization was observed with DNA of serotypes 1, 2, 14 and 1/2. Taken together, these hybridization data show that the *cps2* gene cluster can be divided into three regions: a central region

containing the type-specific genes is flanked by two regions containing common genes for various serotypes.

[00145] Cloning of the type-specific *cps* genes of serotypes 1 and 9. To clone the type-specific *cps* genes of *S. suis* serotype 1, the *cps2E* gene was used as a probe to identify chromosomal DNA fragments of type 1 which contain flanking *cps* genes. A 5 kb *EcoRV* fragment was identified and cloned in pKUN19. This yielded pCPS1-1 (FIG. 1, part B). This fragment was in turn used as a probe to identify an overlapping 2.2 kb *HindIII* fragment. pKUN19 containing this *HindIII* fragment was designated pCPS1-2. The same strategy was followed to identify and clone the type-specific *cps* genes of serotype 9. In this case, we used the *cps2D* gene as a probe. A 0.8 kb *HindIII-XbaI* fragment was identified and cloned, yielding pCPS9-1 (FIG. 1, part C). This fragment was in turn used as a probe to identify a 4 kb *XbaI* fragment. pKUN19 containing this 4 kb *XbaI* fragment was designated pCPS9-2.

[00146] Analysis of the cloned *cps1* genes. The complete nucleotide sequence of the inserts of pCPS1-1 and pCPS1-2 was determined (FIG. 5). Examination of the sequence revealed the presence of five complete and two incomplete Orfs (FIG. 1, part B). Each Orf is preceded by a ribosome-binding site. In accord with data obtained for the *cps2* genes of serotype 2, the majority of the Orfs is very closely linked. The only significant gap (718 bp) was found between Cps1G and Cps1H. No obvious promoter sequences or potential stem-loop structures could be found in this region. This suggests that, as in serotype 2, the *cps* genes in serotype 1 are arranged in an operon.

[00147] An overview of the Orfs and their properties is shown in Table 2. As expected on the basis of the hybridization data (Table 4), the protein encoded by the *cps1E* gene was related to Cps2E of *S. suis* type 2 (identity of 86%). The fragment cloned in pCPS1-1 lacked the coding region for the first 7 amino acids of the *cps1E* gene.

[00148] The protein encoded by the *cps1F* and *cps1G* genes showed strong similarity to the Cps14F and Cps14G proteins of *Streptococcus pneumoniae* serotype 14, respectively (20). The function of the Cps14F is not completely clear, but it has been suggested that Cps14F has a role in glycosyltransferase activity. The *cps14G* gene of *S. pneumoniae* was shown to encode β -1,4-galactosyltransferase activity. In *S. pneumoniae* type 14, this activity is required for the second step in the biosynthesis of the oligosaccharide subunit (20). Based on the similarity of the data,

similar glycosyltransferase and enhancing activities are suggested for the *cps1G* and *cps1F* genes of *S. suis* type 1.

[00149] The protein encoded by the *cps1H* gene showed similarity to the Cps14H protein of *S. pneumoniae* (20). Based on sequence similarity, Cps14H was proposed to be the polysaccharide polymerase (20).

[00150] The protein encoded by the *cps1I* gene showed some similarity with the Cps14J protein of *S. pneumoniae* (19). The *cps14J* gene was shown to encode a β -1,4-galactosyltransferase activity, responsible for the addition of the fourth (*i.e.* last) sugar in the synthesis of the *S. pneumoniae* serotype 14 polysaccharide.

[00151] Between Cps1G and Cps1H, a gap of 718 bp was found. This region revealed three small Orfs. The three Orfs were expressed in three different reading frames and were not preceded by potential ribosome binding sites, nor contained potential start sites. However, the three potential gene products encoded by this region showed some similarity with three successive regions of the C-terminal part of the EpsK protein of *Streptococcus thermophilus* (27% identity, 40). The region related to the first 82 amino acids is lacking.

[00152] **Analysis of the cloned *cps9* genes.** We also determined the complete nucleotide sequence of the inserts of pCPS9-1 and pCPS9-2 (FIG. 6). Examination of the sequence revealed the presence of three complete and two incomplete Orfs (FIG. 1, part C). As in serotypes 1 and 2, all Orfs are preceded by a ribosome-binding site and are very closely coupled. As suggested by the hybridization data (Table 4), the Cps2D and Cps9D proteins were highly related (Table 2). Based on sequence comparisons, pCPS9-1 lacked the first 27 amino acids of the Cps9D protein.

[00153] The protein encoded by the *cps9E* gene showed some similarity with the CapD protein of *Staphylococcus aureus* serotype 1 (24). Based on sequence similarity data, the Cap1D protein was suggested to be an epimerase or a dehydratase involved in the synthesis of N-acetylfructosamine or N- acetylgalactosamine (63).

[00154] Cps9F showed some similarity to the CapM proteins of *S. aureus* serotypes 5 and 8 (61, 64, 65). Based on sequence similarity data, Cap5M and Cap8M are proposed to be glycosyltransferases (63).

[00155] The protein encoded by the *cps9G* gene showed some similarity to a protein of *Actinobacillus actinomycetemcomitans* (AB002668_4). This protein is part of a gene cluster responsible for the serotype b-specific antigens of *Actinobacillus actinomycetemcomitans*. The function of the protein is unknown.

[00156] The protein encoded by the *cps9H* gene showed some similarity to the *rfbB* gene of *Yersinia enterolitica* (68). The RfbB protein was shown to be essential for O-antigen synthesis, but the function of the protein in the synthesis of the O:3 lipopolysaccharide is unknown.

[00157] **Serotype 1 and serotype 9-specific *cps* genes.** To determine whether the cloned fragments in pCPS1-1, pCPS1-2, pCPS9-1 and pCPS9-2 contained the type-specific genes for serotype 1 and 9, respectively, cross-hybridization experiments were performed. DNA fragments of the individual *cps1* and *cps9* genes were amplified by PCR, labeled with ³²P, and used to probe Southern blots of chromosomal DNA of the reference strains of the 35 different *S. suis* serotypes. The results are shown in Table 5. Based on the data obtained with the *cps2E* probe (Table 4), the *cps1E* probe was expected to hybridize with chromosomal DNA of *S. suis* serotypes 1, 2, 14, 27 and 1/2. The *cps1H*, *cps9E* and *cps9F* probes hybridized with most other serotypes. However, the *cps1F* and *cps1G* and *cps1I* probes hybridized with chromosomal DNA of serotypes 1 and 14 only. The *cps9G* and *cps9H* probes hybridized with serotype 9 only. These data suggest that the *cps9G* and *cps9H* probes are specific for serotype 9 and, therefore, could be useful tools for the development of rapid and sensitive diagnostic tests for *S. suis* type 9 infections.

[00158] **Type-specific PCR.** So far, the probes were tested on the 35 different reference strains only. To test the diagnostic value of the type-specific *cps* probes further, several other *S. suis* serotype 1, 2, 1/2, 9 and 14 strains were used. Moreover, since a PCR-based method would be even more rapid and sensitive than a hybridization test, we tested whether we could use a PCR for the serotyping of the *S. suis* strains. The oligonucleotide primer sets were chosen within the *cps2J*, *cps1I* and *cps9H* genes. Amplified fragments of 675 bp, 380 bp and 390 bp were expected, respectively. The results show that 675 bp fragments were amplified on type 2 and 1/2 strains using *cps2J* primers; 380 bp fragments were amplified on type 1 and 14 strains using *cps1I* primers and 390 bp fragments were amplified on type 9 strains using *cps9H* primers.

[00159] Construction of mutants impaired in capsule production. To evaluate the role of the capsule of *S. suis* type 2 in pathogenesis, we constructed two isogenic mutants in which capsule production was disturbed. To construct mutant 10cpsB, pCPS11 was used. In this plasmid, a part of the *cps2B* gene was replaced by the spectinomycin-resistance gene. To construct mutant strain 10cpsEF, the plasmid pCPS28 was used. In pCPS28, the 3'-end of *cps2E* gene, as well as the 5'-end of *cps2F* gene, were replaced by the spectinomycin-resistance gene. pCPS11 and pCPS28 were used to electrotransform strain 10 of *S. suis* type 2 and spectinomycin-resistant colonies were selected. Southern blotting and hybridization experiments were used to select double cross-over integration events (results not shown). To test whether the capsular structure of the strains 10cpsB and 10cpsEF was disturbed, we used a slide agglutination test using a suspension of the mutant strains in hyperimmune anti-*S. suis* type 2 serum (44). The results showed that even in the absence of serotype-specific antisera, the bacteria agglutinated. This indicates that, in the mutant strains, the capsular structure was disturbed. To confirm this, thin sections of wild-type and mutant strains were compared by electron microscopy. The results showed that, compared to the wild-type (FIG. 3, part A), the amount of capsule produced by the mutant strains was greatly reduced (FIG. 3, parts B and C). Almost no capsular material could be detected on the surface of the mutant strains.

Capsular mutants are sensitive to phagocytosis and killing by porcine alveolar macrophages (“PAM”).

[00160] The capsular mutants were tested for their ability to resist phagocytosis by PAM in the presence of porcine SPF serum. The wild-type strain 10 seemed to be resistant to phagocytosis under these conditions (FIGS. 9A and 9B). In contrast, the mutant strains were efficiently ingested by macrophages (FIGS. 9A and 9B). After 90 minutes, more than 99.7% (strain 10cpsB) and 99.8% (strain 10cpsEF) of the mutant cells were ingested by the macrophages. Moreover, as shown in FIGS. 9A and 9B, the ingested strains were efficiently killed by the macrophages. 90-98% of all ingested cells were killed within 90 min. No differences could be observed between wild-type and mutant strains. These data indicate that the capsule of *S. suis* type 2 efficiently protects the organism from uptake by macrophages *in vitro*.

[00161] Capsular mutants are less virulent for germfree piglets. The virulence properties of the wild-type and mutant strains were tested after experimental infection of newborn germfree pigs (45, 49). Table 1 shows that specific and nonspecific signs of disease could be observed in all pigs inoculated with the wild-type strain. Moreover, all pigs inoculated with the wild-type strain died during the course of the experiment or were killed because of serious illness or nervous disorders (Table 3). In contrast, the pigs inoculated with strains 10cpsB and 10cpsEF showed no specific signs of disease and all pigs survived until the end of the experiment (Table 6). The temperature of the pigs inoculated with the wild-type strain increased 2 days after inoculation and remained high until day 5 (Table 3). The temperature of the pigs inoculated with the mutant strains sometimes exceeded 40°C, however, we could observe significant differences in the fever index (*i.e.*, percent of observations in an experimental group during which pigs showed fever ($>40^{\circ}\text{C}$)) between pigs inoculated with wild-type and mutant strains. All pigs showed increased numbers of polymorphonuclear leucocytes (PMLs) ($>10 \times 10^9$ PMLs per liter) (Table 3). However, in pigs inoculated with the mutant strains, the percentage of samples with increased numbers of PMLs was considerably lower. *S. suis* strains and *B. bronchiseptica* could be isolated from the nasopharynx and feces swab samples of all pigs from 1 day post-infection until the end of the experiment (Table 3). Postmortem, the wild-type strain could frequently be isolated from the central nervous system (“CNS”), kidney, heart, liver, spleen, serosae, joints and tonsils. Mutant strains could easily be recovered from the tonsils, but were never recovered from the kidney, liver or spleen. Interestingly, low numbers of the mutant strains were isolated from the CNS, the serosae, the joints, the lungs and the heart. Taken together, these data strongly indicated that mutant *S. suis* strains, impaired in capsule production, are not virulent for young germfree pigs.

[00162] We describe the identification and the molecular characterization of the *cps* locus, involved in the capsular polysaccharide biosynthesis, of *S. suis*. Most of the genes seemed to belong to a single transcriptional unit, suggesting a coordinate control of these genes. Functions to most of the gene products were assigned. Regions involved in regulation (Cps2A), chain length determination (Cps2B, C), export (Cps2C) and biosynthesis (Cps2E, F, G, H, J, K) were identified. The region involved in biosynthesis is located at the center of the gene cluster and is flanked by two regions containing genes with more common functions. The incomplete *orf2Z* gene was located at

the 5'-end of the cloned fragment. Orf2Z showed some similarity with the YitS protein of *B. subtilis*. However, because the function of the YitS protein is unknown, this did not give us any information about the possible function of Orf2Z. Because the *orf2Z* gene is not a part of the *cps* operon, a role of this gene in polysaccharide biosynthesis is not expected. The Orf2Y protein showed some similarity with the YcxD protein of *B. subtilis* (53). The YcxD protein was suggested to be a regulatory protein. Similarly, Orf2Y may be involved in the regulation of polysaccharide biosynthesis. The Orf2X protein showed similarity with the YAAA proteins of *H. influenzae* and *E. coli*. The function of these proteins is unknown. In *S. suis* type 2, the *orf2X* gene seemed to be the first gene in the *cps2* operon. This suggests a role of Orf2X in the polysaccharide biosynthesis. In *H. influenzae* and *E. coli*, however, these proteins are not associated with capsular gene clusters. The analysis of isogenic mutants impaired in the expression of Orf2X should give more insight in the presumed role of Orf2X in the polysaccharide biosynthesis of *S. suis* type 2.

[00163] The gene products encoded by the *cps2E*, *cps2F*, *cps2G*, *cps2H*, *cps2J* and *cps2K* genes showed little similarity with glycosyltransferases of several Gram-positive or Gram-negative bacteria (18, 19, 20, 22, 25). The *cps2E* gene product shows some similarity with the Cps14E protein of *S. pneumoniae* (18, 19). Cps14E is a glucosyl-1-phosphate transferase that links glucose to a lipid carrier (18). In *S. pneumoniae* this is the first step in the biosynthesis of the oligosaccharide repeating unit. The structure of the *S. suis* serotype 2 capsule contains glucose, galactose, rhamnose, N-acetyl glucosamine and sialic acid in a ratio of 3:1:1:1:1 (7). Based on these data, we conclude that Cps2E of *S. suis* has glycosyltransferase activity and is involved in the linkage of the first sugar to the lipid carrier.

[00164] The C-terminal region of the *cps2F* gene product showed some similarity with the RfbU of *Salmonella enteritica*. RfbU was shown to have mannosyltransferase activity (24). Because mannosyl is not a component of the *S. suis* type 2 polysaccharide, a mannosyltransferase activity is not expected in this organism. Nevertheless, *cps2F* encodes a glycosyltransferase with another sugar specificity.

[00165] Cps2G showed moderate similarity to a family of gene products suggested to encode galactosyltransferase activities (22, 24, 40). Hence, a similar activity is shown for Cps2G.

[00166] Cps2H showed some similarity with LgtD of *H. influenzae* (U32768). Because LgtD was proposed to have glycosyltransferase activity, a similar activity is fulfilled by Cps2H.

[00167] Cps2J and Cps2K showed similarity to Cps14J of *S. pneumoniae* (20). Cps2J showed similarity with Cps14I of *S. pneumoniae* as well. Cps14I was shown to have N-acetyl glucosaminyltransferase activity, whereas Cps14J has a β -1,4-galactosyltransferase activity (20). In *S. pneumoniae*, Cps14I is responsible for the addition of the third sugar and Cps14J for the addition of the last sugar in the synthesis of the type 14 repeating unit (20). Because the capsule of *S. suis* type 2 contains galactose as well as N-acetyl glucosamine components, galactosyltransferase as well as N-acetyl glucoaminyltransferase activities could be envisaged for the *cps2J* and *cps2K* gene products, respectively. As was observed for Cps14I and Cps14J, the N-termini of Cps2J and Cps2K showed a significant degree of sequence similarity. Within the N-terminal domains of Cps14I and Cps14J, two small regions were identified, which were also conserved in several other glycosyltransferases (22). Within these two regions, two Asp residues were proposed to be important for catalytic activity. The two conserved regions, DXS and DXDD, were also found in Cps2J and Cps2K.

[00168] The function of Cps2I remains unclear. Cps2I showed some similarity with a protein of *A. actinomycetemcomitans*. Although this protein part is of the gene cluster responsible for the serotype-B-specific antigens, the function of the protein is unknown.

[00169] We further describe the identification and characterization of the *cps* genes specific for *S. suis* serotypes 1, 2 and 9. After the entire *cps2* locus of *S. suis* serotype 2 was cloned and characterized, functions for most of the *cps2* gene products could be assigned by sequence homologies. Based on these data, the glycosyltransferase activities, required for type specificity, could be located in the center of the operon. Cross-hybridization experiments, using the individual *cps2* genes as probes on chromosomal DNAs of the 35 different serotypes, confirmed this idea. The regions containing the type-specific genes of serotypes 1 and 9 could be cloned and characterized, showing that an identical genetic organization of the *cps* operons of other *S. suis* serotypes exists. The *cps1E*, *cps1F*, *cps1G*, *cps1H*, and *cps1I* genes revealed a striking similarity with *cps14E*, *cps14F*, *cps14G*, *cps14H* and *cps14J* genes of *S. pneumoniae*. Interestingly, *S. pneumoniae* serotype 14 is the serotype most commonly associated with pneumococcal infections in young children (54),

whereas *S. suis* serotype 1 strains are most commonly isolated from piglets younger than 8 weeks (46). In *S. pneumoniae*, the *cps14E*, *cps14G*, *cps14I* and *cps14J* encode the glycosyltransferases required for the synthesis of the type 14 tetrmeric repeating unit, showing that the *cps1E*, *cps1G* and *cps1I* genes encoded glycosyltransferases. The precise functions of these genes as well as the substrate specificities of the enzymes can be established. In *S. pneumoniae*, the *cps14E* gene was shown to encode a glucosyl-1-phosphate transferase catalyzing the transfer of glucose to a lipid carrier. Moreover, *cpsE-like* genes were found in *S. pneumoniae* serotypes 9N, 13, 14, 15B, 15C, 18F, 18A and 19F (60). *CpsE* mutants were constructed in the serotypes 9N, 13, 14 and 15B. All mutant strains lacked glucosyltransferase activity (60). Moreover, in all these *S. pneumoniae* serotypes, the *cpsE* gene seemed to be responsible for the addition of glucose to the lipid carrier. Based on these data, we suggest that in *S. suis* type 1, the *cps1E* gene may fulfill a similar function. The structure of the *S. suis* type 1 capsule is unknown, but it is composed of glucose, galactose, N-acetyl glucosamine, N-acetyl galactosamine and sialic acid in a ratio of 1: 2.4: 1: 1:1.4 (5). Therefore, a role of a *cpsE-like* glucosyltransferase activity can easily be envisaged. *CpsE-like* sequences were also found in serotypes 2, 1/2 and 14.

[00170] For polysaccharide biosynthesis in *S. pneumoniae* type 14, transfer of the second sugar of the repeating unit to the first lipid-linked sugar is performed by the gene products of *cps14F* and *cps14G* (20). Similar to Cps14F and Cps14G, the *S. suis* type 1 proteins Cps1F and Cps1G may act as one glycosyltransferase performing the same reaction. Cps14F and Cps14G of *S. pneumoniae* showed similarity to the N-terminal half and C-terminal half of the SpsK protein of *Sphingomonas* (20, 67), respectively. This suggests a combined function for both proteins. Moreover, *cps14F*- and *cps14G*-like sequences were found in several serotypes of *S. pneumoniae* and these genes always seemed to exist together (60). The same was observed for *S. suis* type 1. The *cps1F* and *cps1G* probes hybridized with type 1 and type 14 strains.

[00171] According to the similarity found between the *cps1H* gene and the *cps14H* gene of *S. pneumoniae* (20), *cps1H* is expected to encode a polysaccharide polymerase.

[00172] The protein encoded by the *cps1I* gene showed some similarity with the Cps14J protein of *S. pneumoniae* (19). The *cps14J* gene was shown to encode a β -1,4-galactosyltransferase activity, responsible for the addition of the fourth (*i.e.* last) sugar in the synthesis of the *S.*

pneumoniae serotype 14 polysaccharide. In *S. suis* type 2, the proteins encoded by the *cps2J* and *cps2K* genes showed similarity to the Cps14J protein. However, no significant homologies were found between Cps2J, Cps2K and Cps1I. In the N-terminal regions of Cps14J and Cps14I, two small conserved regions, DXS and DXDD, were identified (19). These regions seemed to be important for catalytic activity (13). At the same positions in the sequence, Cps2I contained the regions DXS and DXED.

[00173] In the region between Cps1G and Cps1H, three small Orfs were identified. Since the Orfs were expressed in three different reading frames, and did not contain potential start sites, expression is not expected. However, the three potential gene products encoded by this region showed some similarity with three successive regions of the C-terminal part of the EpsK (protein of *Streptococcus thermophilus* (27% identity, 40). The region related to the first 82 amino acids is lacking. The EpsK protein was suggested to play a role in the export of the exopolysaccharide by rendering the polymerized exopolysaccharide more hydrophobic through a lipid modification. These data could suggest that the sequences in the region between Cps1G and Cps1H originated from an *epsK*-like sequence. Hybridization experiments showed that this *epsK*-like region is also present in other serotype 1 strains as well as in serotype 14 strains (results not shown).

[00174] The function of most of the cloned serotype 9 genes can be established. Based on sequence similarity data, the *cps9E* and *cps9F* genes could be glycosyltransferases (61, 24, 63, 64, 65). Moreover, the *cps9G* and *cps9H* genes showed similarity to genes located in regions involved in polysaccharide biosynthesis, but the function of these genes is unknown (68).

[00175] Cross-hybridization experiments using the individual *cps2*, *cps1* and *cps9* genes as probes showed that the *cps9G* and *cps9H* probes specifically hybridized with serotype 9 strains. Therefore, these are useful as tools for the identification of *S. suis* type 9 strains both for diagnostic purposes as well as in epidemiological and transmission studies. We previously developed a PCR method which can be used to detect *S. suis* strains in nasal and tonsil swabs of pigs (62). The method was used to identify pathogenic (EF-positive) strains of *S. suis* serotype 2. Besides *S. suis* type 2 strains, serotype 9 strains are frequently isolated from organs of diseased pigs. However, until now, a rapid and sensitive diagnostic test was not available for type 9 strains. Therefore, the type 9-specific probes or the type 9-specific PCR is of great diagnostic value. The *cps1F*, *cps1G* and

cps1I probes hybridized with serotype 1 as well as with serotype 14 strains. In coagglutination tests, type 1 strains react with the anti-type 1 as well as with the anti-type 14 antisera (56). This suggests the presence of common epitopes between these serotypes. On the other hand, type 1 strains agglutinated only with anti-type 1 serum (56, 57), indicating that it is possible to detect differences between those serotypes.

[00176] The *cps2F*, *cps2G*, *cps2H*, *cps2I* and *cps2J* probes hybridized with serotypes 2 and 1/2 only. Serotype 34 showed a weak hybridizing signal with the *cps2G* probe. As shown in agglutination tests, type 1/2 strains react with sera directed against type 1 as well as with sera directed against type 2 strains (46). Therefore, type 1/2 shared antigens with both types 1 and 2. Based on the hybridization patterns of serotype 1/2 strains with the *cps1*- and *cps2*-specific genes, serotype 1/2 seemed to be more closely related to type 2 strains than to type 1 strains. In our current studies, we identify type-specific genes, primers or probes which are used for the discrimination of serotypes 1, 14 and 2 and 1/2 and others of the 35 serotypes yet known. Furthermore, type-specific genes, primers or probes can now easily be developed for yet unknown serotypes, once they become isolated.

Cloning and characterization of a further part of the *cps2* locus.

[00177] Based on the established sequence, 11 genes, designated *cps2L* to *cps2T*, *orf2U* and *orf2V*, were identified. A gene homologous to genes involved in the polymerization of the repeating oligosaccharide unit (*cps20*) as well as genes involved in the synthesis of sialic acid (*cps2P* to *cps2T*) were identified. Moreover, hybridization experiments showed that the genes involved in the sialic acid synthesis are present in *S. suis* serotypes 1, 2, 14, 27 and 1/2. The “*cps2M*” and “*cps2N*” regions showed similarity to proteins involved in the polysaccharide biosynthesis of other Gram-positive bacteria. However, these regions seemed to be truncated or were non-functional as the result of frame-shift or point mutations. At its 3'-end, the *cps2* locus contained two insertional elements (“*orf2U*” and “*orf2V*”), both of which seemed to be non-functional.

[00178] To clone the remaining part of the *cps2* locus, sequences of the 3'-end of pCPS26 (FIG. 1, part C) were used to identify a chromosomal fragment containing *cps2* sequences located further downstream. This fragment was cloned in pKUN19, resulting in pCPS29. Using a similar

approach, we subsequently isolated the plasmids pCPS30 and pCPS34 containing downstream *cps2* sequences (FIG. 1, part C).

Analysis of the *cps2* operon.

[00179] The complete nucleotide sequence of the cloned fragments was determined. Examination of the compiled sequence revealed the presence of: a sequence encoding the C-terminal part of Cps2K, six apparently functional genes (designated *cps2O-cps2T*) and the remnants of 5 different ancestral genes (designated “*cps2L*,” “*cps2M*,” “*cps2N*,” “*orf2U*” and “*orf2V*”). The latter genes seemed to be truncated or incomplete as the result of the presence of stop codons or frame-shift mutations (FIG. 1, part A). Neither potential promoter sequences nor potential stem-loop structures could be identified within the sequenced region. A ribosome-binding site precedes each ORF and the majority of the ORFs are very closely linked. Three intergenic gaps were found: one between “*cps2M*” and “*cps2N*” (176 nucleotides), one between *cps2O* and *cps2P* (525 nucleotides), and one between *cps2T* and “*orf2U*” (200 nucleotides). These and our above data show that *Orf2X* and *Cps2A-Orf2T* are part of a single operon.

[00180] A list of all loci and their properties is shown in Table 4. The “*cps2L*” region contained three potential ORFs of 103, 79 and 152 amino acids, respectively, which were only separated from each other by stop codons. Only the first ORF is preceded by a potential ribosomal binding site and contained a methionine start codon. This suggests that “*cps2L*” originates from an ancestral *cps2L* gene, which coded for a protein of 339 amino acids. The function of this hypothetical *cps2L* protein remains unclear so far: no significant homologies were found between *Cps2L* and proteins present in the data libraries. It is not clear whether the first ORF of the “*cps2L*” region is expressed into a protein of 103 amino acids. The “*cps2M*” region showed homology to the N-terminal 134 amino acids of the NeuA proteins of *Streptococcus agalactiae* and *Escherichia coli* (AB017355, 32). However, although the “*cps2M*” region contained a potential ribosome binding site, a methionine start codon was absent. Compared with the *S. agalactiae* sequence, the ATG start codon was replaced by a lysin encoding AAG codon. Moreover, the region homologous to the first 58 amino acids of the *S. agalactiae* NeuA (identity 77%) was separated from the region homologous to amino acids 59-134 of NeuA by a repeated DNA sequence of 100-bp (see herein). In addition, the

region homologous to amino acids 59 to 95 of NeuA (identity 32%) and the region homologous to the amino acids 96 to 134 of NeuA (identity 50%) were present in different reading frames. Therefore, the partial and truncated NeuA homologue is probably nonfunctional in *S. suis*. The “cps2N” region showed homology to CpsJ of *S. agalactiae* (accession no. AB017355). However, sequences homologous to the first 88 amino acids of CpsJ were lacking in *S. suis*. Moreover, the homologous region was present in two different reading frames. The protein encoded by the cps2O gene showed homology to proteins of several streptococci involved in the transport of the oligosaccharide repeating unit (accession no. AB017355), suggesting a similar function for Cps2O. The proteins encoded by the cps2P, cps2S and cps2T genes showed homology to the NeuB, NeuD and NeuA proteins of *S. agalactiae* and *E. coli* (accession no AB017355). Because the “cps2M” region also showed homology to NeuA of *E. coli*, the *S. suis* cps2 locus contains a functional neuA gene (cps2T) as well as a nonfunctional (“cps2M”) gene. The mutual homology between these two regions showed an identity of 77% at the amino acid level over amino acids 1-58 and 49% over the amino acids 59-134. Cps2Q and Cps2R showed homology to the N-terminal and C-terminal parts of the NeuC protein of *S. agalactiae* and *E. coli*, respectively. This suggests that the function of the *S. agalactiae* NeuC protein in *S. suis* is likely fulfilled by two different proteins. In *E. coli*, the neu genes are known to be involved in the synthesis of sialic acid. NeuNAc is synthesized from N-acetylmannosamine and phosphoenolpyruvate by NeuNAc synthetase. Subsequently, NeuNAc is converted to CMP-NeuNAc by the enzyme CMP-NeuNAc synthetase. CMP-NeuNAc is the substrate for the synthesis of polysaccharide. In *E. coli*, K1 NeuB is the NeuNAc synthetase, and NeuA is the CMP-NeuNAc synthetase. NeuC has been implicated in the NeuNAc synthesis, but its precise role is not known. The precise role of NeuD is not known. A role of the Cps2P-Cps2T proteins in the synthesis of sialic acid can easily be envisaged, since the capsule of *S. suis* serotype 2 is rich in sialic acid. In *S. agalactiae*, sialic acid has been shown to be critical to the virulence function of the type III capsule. Moreover, it has been suggested that the presence of sialic acid in the capsule of bacteria which can cause meningitis may be important for these bacteria to breach the blood-brain barrier. So far, however, the requirement of the sialic acid for virulence of *S. suis* remains unclear.

[00181] “Orf2U” and “Orf2V” showed homology to proteins located on two different insertional elements. “Orf2U” is homologous to IS1194 of *Streptococcus thermophilus*, whereas “Orf2V” showed homology to a putative transposase of *Streptococcus pneumoniae*. This putative transposase was recently found to be associated with the type 2 capsular locus of *S. pneumoniae*. Compared with the original insertional elements in *S. thermophilus* and *S. pneumoniae*, both “Orf2U” and “Orf2V” are likely to be non-functional due to frame shift mutations within their coding regions.

[00182] A striking observation was the presence of a sequence of 100 bp (FIG. 10) which was repeated three times within the cps2 operon. The sequence is highly conserved (between 94% and 98%) and was found in the intergenic regions between cps2G and cps2H, within “cps2M” and between cps2O and cps2P. No significant homologies were found between this 100-bp direct repeat sequence and sequences present in the data libraries, suggesting that the sequence is unique for *S. suis*.

Distribution of the cps2 sequences among the 35 *S. suis* serotypes.

[00183] To examine the presence of sialic acid encoding genes in other *S. suis* serotypes, we performed cross-hybridization experiments. DNA fragments of the individual cps2 genes were amplified by PCR, radiolabeled with 32P and hybridized to chromosomal DNA of the reference strains of the 35 different *S. suis* serotypes. As a positive control, we used a probe specific for *S. suis* 16S rRNA. The 16S rRNA probe hybridized with almost equal intensities to all serotypes tested (Table 4). The “cps2L” sequence hybridized with DNA of serotypes 1, 2, 14 and 1/2. The “cps2M”, cps2O, cps2P, cps2Q, cps2R, cps2S and cps2T genes hybridized with DNA of serotypes 1, 2, 14, 27 and 1/2. Because the cps2P-cps2T genes are most likely involved in the synthesis of sialic acid, these results suggest that sialic acid is also a part of the capsule in the *S. suis* serotypes 1, 2, 14, 27 and 1/2. This is in agreement with the finding that the serotypes 1, 2 and 1/2 possess a capsule that is rich in sialic acid. Although the chemical compositions of the capsules of serotypes 14 and 27 are unknown, recent agglutination studies using sialic acid-binding lectins suggested the presence of sialic acid in *S. suis* serotype 14, but not in serotype 27. In these studies, sialic acid was also detected in serotypes 15 and 16. Since the latter observation is not in agreement with our

hybridization studies, it might be that other genes, not homologous to the *cps2P-cps2T* genes, are responsible for the sialic acid synthesis in serotypes 15 and 16.

[00184] A probe based on “*cps2N*” sequences hybridized with DNA from serotypes 1, 2, 14 and 1/2. A probe specific for “*orf2U*” hybridized with serotypes 1, 2, 7, 14, 24, 27, 32, 34, and 1/2, whereas a probe specific for “*orf2V*” hybridized with many different serotypes. In addition, we prepared a probe specific for the 100-bp direct repeat sequence. This probe hybridized with the serotypes 1, 2, 13, 14, 22, 24, 27, 29, 32, 34 and 1/2 (Table 4). To analyze the number of copies of the direct repeat sequence within the *S. suis* serotype 2 chromosome, a Southern blot hybridization and analysis was performed. Therefore, chromosomal DNA of *S. suis* serotype 2 was digested with *Nco*I and hybridized with a 32P-labeled direct repeat sequence. Only one hybridizing fragment, containing the three direct repeats present on the *cps2* locus, was found (results not shown). This indicates that the 100-bp direct repeat sequence is only associated with the *cps2* locus. In *S. pneumoniae*, a 115-bp long repeated sequence was found to be associated with the capsular genes of serotypes 1, 3, 14 and 19F. In *S. pneumoniae*, this 115-bp sequence was also found in the vicinity of other genes involved in pneumococcal virulence (hyaluronidase and neuraminidase genes). A regulatory role of the 115-bp sequence in coordinate control of these virulence-related genes was suggested.

[00185] To study the role of the capsule in resistance to phagocytosis and in virulence, we constructed two isogenic mutants in which capsule synthesis was disturbed. In 10cpsB, the *cps2B* gene was disturbed by the insertion of an antibiotic-resistance gene, whereas in 10cpsEF, parts of the *cps2E* and *cps2F* genes were replaced. Both mutant strains seemed to be completely unencapsulated. Because the *cps2* genes seemed to be part of an operon, polar effects cannot be excluded. Therefore, these data did not give any information about the role of Cps2B, Cps2E or Cps2F in the polysaccharide biosynthesis. However, the results clearly show that the capsular polysaccharide of *S. suis* type 2 is a surface component with antiphagocytic activity. *In vitro* wild-type encapsulated bacteria are ingested by phagocytes at a very low frequency, whereas the mutant unencapsulated bacteria are efficiently ingested by porcine macrophages. Within 2 hours, over 99.6% of mutant bacteria were ingested and over 92% of the ingested bacteria were killed. Intracellularly, wild-type as well as mutant strains seemed to be killed with the same efficiency. This suggests that the loss of

capsular material is associated with loss of capacity to resist uptake by macrophages. This loss of resistance to *in vitro* phagocytosis was associated with a substantial attenuation of the virulence in germfree pigs. All pigs inoculated with the mutant strains survived the experiment and did not show any specific clinical signs of disease. Only some aspecific clinical signs of disease could be observed. Moreover, mutant bacteria could be reisolated from the pigs. This supports the idea that, as in other pathogenic Streptococci, the capsule of *S. suis* acts as an important virulence factor. Transposon mutants prepared by Charland impaired in the capsule production showed a reduced virulence in pigs and mice. To construct these mutants, the type 2 reference strain S735 was used. We previously showed that this strain is only weakly virulent for young pigs. Moreover, the insertion site of the transposon is unsolved so far.

As a further example herein a rapid PCT test for *Streptococcus suis* type 7 is described.

[00186] Recent epidemiological studies on *Streptococcus suis* infections in pigs indicated that, besides serotypes 1, 2 and 9, serotype 7 is also frequently associated with diseased animals. For the latter serotype, however, no rapid and sensitive diagnostic methods are available. This hampers prevention and control programs. Here we describe the development of a type-specific PCR test for the rapid and sensitive detection of *S. suis* serotype 7. The test is based on DNA sequences of capsular (cps) genes specific for serotype 7. These sequences could be identified by cross-hybridization of several individual cps genes with the chromosomal DNAs of 35 different *S. suis* serotypes.

[00187] *Streptococcus suis* is an important cause of meningitis, septicemia, arthritis and sudden death in young pigs [69, 70]. It can, however, also cause meningitis in man (71). Attempts to control the disease are still hampered by the lack of sufficient knowledge about the epidemiology of the disease and the lack of effective vaccines and sensitive diagnostics.

[00188] *S. suis* strains can be identified and classified by their morphological, biochemical and serological characteristics (70, 73, 74). Serological classification is based on the presence of specific antigenic determinants. Isolated and biochemically characterized *S. suis* cells are agglutinated with a panel of specific sera. These typing methods are very laborious and

time-consuming and can only be performed on isolated colonies. Moreover, it has been reported that nonspecific cross-reactions may occur among different types of *S. suis* (75, 76).

[00189] So far, 35 different serotypes have been described (7, 78, 79). *S. suis* serotype 2 is the most prevalent type isolated from diseased pigs, followed by serotypes 9 and 1. However, recently, serotype 7 strains were also frequently isolated from diseased pigs (80, 81, 82). This suggests that infections with *S. suis* serotype 7 strains seemed to be an increasing problem. Moreover, the virulence of *S. suis* serotype 7 strains was confirmed by experimental infection of young pigs (83).

[00190] Recently, rapid and sensitive PCR assays specific for serotypes 2 (and 1/2), 1 (and 14) and 9 were developed (84). These assays were based on the cps loci of *S. suis* serotypes 2, 1 and 9 (84, 85). However, until now, no rapid and sensitive diagnostic test was available for *S. suis* serotype 7. Herein we describe the development of a PCR test for the rapid and sensitive detection of *S. suis* serotype 7 strains. The test is based on DNA sequences which form a part of the cps locus of *S. suis* serotype 7. Compared with the serological serotyping methods, the PCR assay was a rapid, reliable and sensitive assay. Therefore, this test, in combination with the PCR tests which we previously developed for serotypes 1, 2 and 9, will undoubtedly contribute to a more rapid and reliable diagnosis of *S. suis* and may facilitate control and eradication programs.

Materials and Methods

Bacterial strains, growth conditions and serotyping.

[00191] The bacterial strains and plasmids used in this study are listed in Table 7. The *S. suis* reference strains were obtained from M. Gottschalk, Canada. *S. suis* strains were grown in Todd-Hewitt broth (code CM189, Oxoid), and plated on Columbia agar blood base (code CM331, Oxoid) containing 6% (v/v) horse blood. *E. coli* strains were grown in Luria broth (86) and plated on Luria broth containing 1.5% (w/v) agar. If required, ampicillin was added to the plates. The *S. suis* strains were serotyped by the slide agglutination test with serotype-specific antibodies (70).

DNA techniques.

[00192] Routine DNA manipulations and PCR reactions were performed as described by Sambrook et al. (88). Blotting and hybridization were performed as described previously (84, 86).

DNA sequence analysis.

[00193] DNA sequences were determined on a 373A DNA Sequencing system (Applied Biosystems, Warrington, GB). Samples were prepared by use of an ABI/PRISM dye terminator cycle sequencing ready reaction kit (Applied Biosystems). Custom-made sequencing primers were purchased from Life Technologies. Sequencing data were assembled and analyzed using the McMollyTetra program. The BLAST program was used to search for protein sequences homologous to the deduced amino acid sequences.

PCR.

[00194] The primers used for the cps7H PCR correspond to the positions 3334-3354 and 3585-3565 in the *S. suis* cps7 locus. The sequences were: 5'-AGCTCTAACACGAAATAAGGC-3' (SEQ ID NO:7) and 5'-GTCAAACACCCTGGATAGCCG-3' (SEQ ID NO:8).

[00195] The reaction mixtures contained 10 mM Tris-HCl, pH 8.3; 1.5 mM MgCl₂; 50 mM KCl; 0.2 mM of each of the four deoxynucleotide triphosphates; 1 microM of each of the primers and 1U of AmpliTaq Gold DNA polymerase (Perkin Elmer Applied Biosystems, New Jersey). DNA amplification was carried out in a Perkin Elmer 9600 thermal cycler and the program consisted of an incubation for 10 min at 95°C and 30 cycles of 1 min at 95°C, 2 min at 56°C and 2 min at 72°C.

Results and discussion

Cloning of the serotype 7-specific cps genes.

[00196] To isolate the type-specific cps genes of *S. suis* serotype 7, we used the cps9E gene of serotype 9 as a probe to identify chromosomal DNA fragments of type 7 containing homologous DNA sequences (84). A 1.6-kb PstI fragment was identified and cloned in pKUN19. This yielded pCPS7-1 (FIG. 11, part C). In turn, this fragment was used as a probe to identify an overlapping 2.7

kb ScaI-Clal fragment. pGEM7 containing the latter fragment was designated pCPS7-2 (FIG. 11, part C).

Analysis of the cloned cps 7 genes.

[00197] The complete nucleotide sequences of the inserts of pCPS7-1, pCPS7-2 were determined. Examination of the cps7 sequence revealed the presence of two complete and two incomplete open reading frames (ORFs) (FIG. 11, part C). All ORFs are preceded by a ribosome-binding site. In accord with the data obtained for the cps1, cps2 and cps9 genes of serotypes 1, 2 and 9, respectively, the type 7 ORFs are very closely linked to each other. The only significant intergenic gap was that found between cps7E and cps7F (443 nucleotides). No obvious promoter sequences or potential stem-loop structures were found in this region. This suggests that, as in serotypes 1, 2 and 9, the cps genes in serotype 7 form part of an operon.

[00198] An overview of the ORFs and their properties is shown in Table 8. As expected on the basis of the hybridization data (84), the Cps9E and Cps7E proteins showed a high similarity (identity 99%, Table 8). Based on sequence comparisons between Cps9E and Cps7E, the PstI fragment of pCPS7-1 lacks the region encoding the first 371 codons of Cps7E. The C-terminal part of the protein encoded by the cps7F gene showed some similarity with the BplG protein of *Bordetella pertussis* (88), as well as with the C-terminal part of *S. suis* Cps2E (85). Both BplG and Cps2E were suggested to have glycosyltransferase activity and are probably involved in the linkage of the first sugar to the lipid carrier (85, 88). The protein encoded by the cps7G gene showed similarity with the BlpF protein of *Bordetella pertussis* (88). BlpF is likely to be involved in the biosynthesis of an amino sugar, suggesting a similar function for Cps7G. The protein encoded by the cps7H gene showed similarity with the WbdN protein of *E. coli* (89) as well as with the N-terminal part of the Cps2K protein of *S. suis* (81). Both WbdN and Cps2K were suggested to have glycosyltransferase activity (85, 89).

Serotype 7-specific cps genes.

[00199] To determine whether the cloned fragments in pCPS7-1 and pCPS7-2 contained serotype 7-specific DNA sequences, cross-hybridization experiments were performed. DNA

fragments of the individual *cps7* genes were amplified by PCR, labeled with ³²P, and used to probe spot blots of chromosomal DNA of the reference strains of 35 different *S. suis* serotypes. The results are summarized in Table 9. As expected, based on the data obtained with the *cps9E* probe (84), the *cps7E* probe hybridized with chromosomal DNA of many different *S. suis* serotypes. The *cps7F* and *cps7G* probes showed hybridization with chromosomal DNA of *S. suis* serotypes 4, 5, 7, 17, and 23. However, the *cps7H* probe hybridized with chromosomal DNA of serotype 7 only, indicating that this gene is specific for serotype 7.

Type-specific PCR.

[00200] We tested whether we could use PCR instead of hybridization for the typing of the *S. suis* serotype 7 strains. For that purpose, we selected an oligonucleotide primer set within the *cps7H* gene with which an amplified fragment of 251-bp was expected. In addition, we included in our analysis several *S. suis* serotype 7 strains, other than the reference strain. These strains were obtained from different countries and were isolated from different organs (Table 7). The results show that indeed a fragment of about 250-bp was amplified with all type 7 strains used (FIG. 12, part B), whereas no PCR products were obtained with serotype 1, 2 and 9 strains (FIG. 12, part A). This suggests that the PCR test, as described here, is a rapid diagnostic tool for the identification of *S. suis* serotype 7 strains. Until now, such a diagnostic test was not available for serotype 7 strains. Together with the recently developed PCR assays for serotypes 1, 2, 1/2, 14 and 9, this assay may be an important diagnostic tool to detect pigs carrying serotype 2, 1/2, 1, 14, 9 and 7 strains and may facilitate control and eradication programs.

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<210> 10
<211> 239
<212> PRT
<213> *Streptococcus suis*
<220>
<221> misc_feature
<223> ORF2Z
<400> 10

Ser Leu Asp Ile Asp His Met Met Glu Val Met Glu Ala Ser Lys Ser
1 5 10 15

Ala Ala Gly Ser Ala Cys Pro Ser Pro Gln Ala Tyr Gln Ala Ala Phe
20 25 30

Glu Gly Ala Glu Asn Ile Ile Val Val Thr Ile Thr Gly Gly Leu Ser
35 40 45

Gly Ser Phe Asn Ala Ala Arg Val Ala Arg Asp Met Tyr Ile Glu Glu
50 55 60

His Pro Asn Val Asn Ile His Leu Ile Asp Ser Leu Ser Ala Ser Gly
65 70 75 80

Glu Met Asp Leu Leu Val His Gln Ile Asn Arg Leu Ile Ser Ala Gly
85 90 95

Leu Asp Phe Pro Gln Val Val Glu Ala Ile Thr His Tyr Arg Glu His
100 105 110

Ser Lys Leu Leu Phe Val Leu Ala Lys Val Asp Asn Leu Val Lys Asn
115 120 125

Gly Arg Leu Ser Lys Leu Val Gly Thr Val Val Gly Leu Leu Asn Ile
130 135 140

Arg Met Val Gly Glu Ala Ser Ala Glu Gly Lys Leu Glu Leu Leu Gln
145 150 155 160

Lys Ala Arg Gly His Lys Lys Ser Val Thr Ala Ala Phe Glu Glu Met
165 170 175

Lys Lys Ala Gly Tyr Asp Gly Gly Arg Ile Val Met Ala His Arg Asn
180 185 190

Asn	Ala	Lys	Phe	Phe	Gln	Gln	Phe	Ser	Glu	Leu	Val	Lys	Ala	Ser	Phe
195							200					205			
Pro	Thr	Ala	Val	Ile	Asp	Glu	Val	Ala	Thr	Ser	Gly	Leu	Cys	Ser	Phe
210					215					220					
Tyr	Ala	Glu	Glu	Gly	Gly	Leu	Leu	Met	Gly	Tyr	Glu	Val	Lys	Ala	
225					230			235							
<210> 11															
<211> 244															
<212> PRT															
<213> <i>Streptococcus suis</i>															
<220>															
<221> misc_feature															
<223> ORF2X															
<400> 11															
Met	Lys	Ile	Ile	Ile	Pro	Asn	Ala	Lys	Glu	Val	Asn	Thr	Asn	Leu	Glu
1					5				10				15		
Asn	Ala	Ser	Phe	Tyr	Leu	Leu	Ser	Asp	Arg	Ser	Lys	Pro	Val	Leu	Asp
				20				25				30			
Ala	Ile	Ser	Gln	Phe	Asp	Val	Lys	Lys	Met	Ala	Ala	Phe	Tyr	Lys	Leu
			35				40				45				
Asn	Glu	Ala	Lys	Ala	Glu	Leu	Glu	Ala	Asp	Arg	Trp	Tyr	Arg	Ile	Arg
	50				55					60					
Thr	Gly	Gln	Ala	Lys	Thr	Tyr	Pro	Ala	Trp	Gln	Leu	Tyr	Asp	Gly	Leu
	65				70				75			80			
Met	Tyr	Arg	Tyr	Met	Asp	Arg	Arg	Gly	Ile	Asp	Ser	Lys	Glu	Glu	Asn
				85				90				95			
Tyr	Leu	Arg	Asp	His	Val	Arg	Val	Ala	Thr	Ala	Leu	Tyr	Gly	Leu	Ile
				100				105				110			
His	Pro	Phe	Glu	Phe	Ile	Ser	Pro	His	Arg	Leu	Asp	Phe	Gln	Gly	Ser
					115			120				125			
Leu	Lys	Ile	Gly	Asn	Gln	Ser	Leu	Lys	Gln	Tyr	Trp	Arg	Pro	Tyr	Tyr
					130			135			140				

Asp	Gln	Glu	Val	Gly	Asp	Asp	Glu	Leu	Ile	Leu	Ser	Leu	Ala	Ser	Ser
145					150					155				160	
Glu	Phe	Glu	Gln	Val	Phe	Ser	Pro	Gln	Ile	Gln	Lys	Arg	Leu	Val	Lys
					165					170				175	
Ile	Leu	Phe	Met	Glu	Glu	Lys	Ala	Gly	Gln	Leu	Lys	Val	His	Ser	Thr
						180				185				190	
Ile	Ser	Lys	Lys	Gly	Arg	Gly	Arg	Leu	Leu	Ser	Trp	Leu	Ala	Lys	Asn
					195				200				205		
Asn	Ile	Gln	Glu	Leu	Ser	Asp	Ile	Gln	Asp	Phe	Lys	Val	Asp	Gly	Phe
						210			215			220			
Glu	Tyr	Cys	Thr	Ser	Glu	Ser	Thr	Ala	Asn	Gln	Leu	Thr	Phe	Ile	Arg
					225					230		235		240	
Ser	Ile	Lys	Met												

<210> 12

<211> 481

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS2A

<400> 12

Met	Lys	Lys	Arg	Ser	Gly	Arg	Ser	Lys	Ser	Ser	Lys	Phe	Lys	Leu	Val
1								5		10				15	

Asn	Phe	Ala	Leu	Leu	Gly	Leu	Tyr	Ser	Ile	Thr	Leu	Cys	Leu	Phe	Leu
								20		25			30		

Val	Thr	Met	Tyr	Arg	Tyr	Asn	Ile	Leu	Asp	Phe	Arg	Tyr	Leu	Asn	Tyr
											35		40	45	

Ile	Val	Thr	Leu	Leu	Leu	Val	Gly	Val	Ala	Val	Leu	Ala	Gly	Leu	Leu
								50		55			60		

Met	Trp	Arg	Lys	Lys	Ala	Arg	Ile	Phe	Thr	Ala	Leu	Leu	Leu	Val	Phe
65					70					75				80	
Ser	Leu	Val	Ile	Thr	Ser	Val	Gly	Ile	Tyr	Gly	Met	Gln	Glu	Val	Val
					85				90				95		
Lys	Phe	Ser	Thr	Arg	Leu	Asn	Ser	Asn	Ser	Thr	Phe	Ser	Glu	Tyr	Glu
					100				105				110		
Met	Ser	Ile	Leu	Val	Pro	Ala	Asn	Ser	Asp	Ile	Thr	Asp	Val	Arg	Gln
						115			120				125		
Leu	Thr	Ser	Ile	Leu	Ala	Pro	Ala	Glu	Tyr	Asp	Gln	Asp	Asn	Ile	Thr
						130			135				140		
Ala	Leu	Leu	Asp	Asp	Ile	Ser	Lys	Met	Glu	Ser	Thr	Gln	Leu	Ala	Thr
						145			150			155			160
Ser	Pro	Gly	Thr	Ser	Tyr	Leu	Thr	Ala	Tyr	Gln	Ser	Met	Leu	Asn	Gly
						165			170				175		
Glu	Ser	Gln	Ala	Met	Val	Phe	Asn	Gly	Val	Phe	Thr	Asn	Ile	Leu	Glu
					180				185				190		
Asn	Glu	Asp	Pro	Gly	Phe	Ser	Ser	Lys	Val	Lys	Lys	Ile	Tyr	Ser	Phe
						195			200				205		
Lys	Val	Thr	Gln	Thr	Val	Glu	Thr	Ala	Thr	Lys	Gln	Val	Ser	Gly	Asp
						210			215				220		
Ser	Phe	Asn	Ile	Tyr	Ile	Ser	Gly	Ile	Asp	Ala	Tyr	Gly	Pro	Ile	Ser
						225			230			235			240
Thr	Val	Ser	Arg	Ser	Asp	Val	Asn	Ile	Ile	Met	Thr	Val	Asn	Arg	Ala
						245			250				255		
Thr	His	Lys	Ile	Leu	Leu	Thr	Thr	Pro	Arg	Asp	Ser	Tyr	Val	Ala	
						260			265				270		
Phe	Ala	Asp	Gly	Gly	Gln	Asn	Gln	Tyr	Asp	Lys	Leu	Thr	His	Ala	Gly
						275			280				285		
Ile	Tyr	Gly	Val	Asn	Ala	Ser	Val	His	Thr	Leu	Glu	Asn	Phe	Tyr	Gly
						290			295				300		
Ile	Asp	Ile	Ser	Asn	Tyr	Val	Arg	Leu	Asn	Phe	Ile	Ser	Phe	Leu	Gln
						305			310			315			320
Leu	Ile	Asp	Leu	Val	Gly	Gly	Ile	Asp	Val	Tyr	Asn	Asp	Gln	Glu	Phe
						325			330				335		

Thr Ser Leu His Gly Asn Tyr His Phe Pro Val Gly Gln Val His Leu
340 345 350

Asn Ser Asp Gln Ala Leu Gly Phe Val Arg Glu Arg Tyr Ser Leu Thr
355 360 365

Gly Gly Asp Asn Asp Arg Gly Lys Asn Gln Glu Lys Val Ile Ala Ala
370 375 380

Leu Ile Lys Lys Met Ser Thr Pro Glu Asn Leu Lys Asn Tyr Gln Ala
385 390 395 400

Ile Leu Ser Gly Leu Glu Gly Ser Ile Gln Thr Asp Leu Ser Leu Glu
405 410 415

Thr Ile Met Ser Leu Val Asn Thr Gln Leu Glu Ser Gly Thr Gln Phe
420 425 430

Thr Val Glu Ser Gln Ala Leu Thr Gly Thr Gly Arg Ser Asp Leu Ser
435 440 445

Ser Tyr Ala Met Pro Gly Ser Gln Leu Tyr Met Met Glu Ile Asn Gln
450 455 460

Asp Ser Leu Glu Gln Ser Lys Ala Ala Ile Gln Ser Val Leu Val Glu
465 470 475 480

Lys

<210> 13

<211> 229

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2B

<400> 13

Met Asn Asn Gln Glu Val Asn Ala Ile Glu Ile Asp Val Leu Phe Leu
1 5 10 15

Leu Lys Thr Ile Trp Arg Lys Lys Phe Leu Ile Leu Leu Thr Ala Val
20 25 30

Leu Thr Ala Gly Leu Ala Phe Val Tyr Ser Ser Phe Leu Val Thr Pro
 35 40 45
 Gln Tyr Asp Ser Thr Thr Arg Ile Tyr Val Val Ser Gln Asn Val Glu
 50 55 60
 Ala Gly Ala Gly Leu Thr Asn Gln Glu Leu Gln Ala Gly Thr Tyr Leu
 65 70 75 80
 Ala Lys Asp Tyr Arg Glu Ile Ile Leu Ser Gln Asp Val Leu Thr Gln
 85 90 95
 Val Ala Thr Glu Leu Asn Leu Lys Glu Ser Leu Lys Glu Lys Ile Ser
 100 105 110
 Val Ser Ile Pro Val Asp Thr Arg Ile Val Ser Ile Ser Val Arg Asp
 115 120 125
 Ala Asp Pro Asn Glu Ala Ala Arg Ile Ala Asn Ser Leu Arg Thr Phe
 130 135 140
 Ala Val Gln Lys Val Val Glu Val Thr Lys Val Ser Asp Val Thr Thr
 145 150 155 160
 Leu Glu Glu Ala Val Pro Ala Glu Glu Pro Thr Thr Pro Asn Thr Lys
 165 170 175
 Arg Asn Ile Leu Leu Gly Leu Leu Ala Gly Gly Ile Leu Ala Thr Gly
 180 185 190
 Leu Val Leu Val Met Glu Val Leu Asp Asp Arg Val Lys Arg Pro Gln
 195 200 205
 Asp Ile Glu Glu Val Met Gly Leu Thr Leu Leu Gly Ile Val Pro Asp
 210 215 220
 Ser Lys Lys Leu Lys
 225

<210> 14
 <211> 225
 <212> PRT
 <213> *Streptococcus suis*
 <220>
 <221> misc_feature

<223> CPS2C

<400> 14

Met Ala Met Leu Glu Ile Ala Arg Thr Lys Arg Glu Gly Val Asn Lys
1 5 10 15

Thr Glu Glu Tyr Phe Asn Ala Ile Arg Thr Asn Ile Gln Leu Ser Gly
20 25 30

Ala Asp Ile Lys Val Val Gly Ile Thr Ser Val Lys Ser Asn Glu Gly
35 40 45

Lys Ser Thr Thr Ala Ala Ser Leu Ala Ile Ala Tyr Ala Arg Ser Gly
50 55 60

Tyr Lys Thr Val Leu Val Asp Ala Asp Ile Arg Asn Ser Val Met Pro
65 70 75 80

Gly Phe Phe Lys Pro Ile Thr Lys Ile Thr Gly Leu Thr Asp Tyr Leu
85 90 95

Ala Gly Thr Thr Asp Leu Ser Gln Gly Leu Cys Asp Thr Asp Ile Pro
100 105 110

Asn Leu Thr Val Ile Glu Ser Gly Lys Val Ser Pro Asn Pro Thr Ala
115 120 125

Leu Leu Gln Ser Lys Asn Phe Glu Asn Leu Leu Ala Thr Leu Arg Arg
130 135 140

Tyr Tyr Asp Tyr Val Ile Val Asp Cys Pro Pro Leu Gly Leu Val Ile
145 150 155 160

Asp Ala Ala Ile Ile Ala Gln Lys Cys Asp Ala Met Val Ala Val Val
165 170 175

Glu Ala Gly Asn Val Lys Cys Ser Ser Leu Lys Lys Val Lys Glu Gln
180 185 190

Leu Glu Gln Thr Gly Thr Pro Phe Leu Gly Val Ile Leu Asn Lys Tyr
195 200 205

Asp Ile Ala Thr Glu Lys Tyr Ser Glu Tyr Gly Asn Tyr Gly Lys Lys
210 215 220

Ala
225

<210> 15
<211> 243
<212> PRT
<213> *Streptococcus suis*
<220>
<221> misc_feature
<223> CPS2D
<400> 15

Met Ile Asp Ile His Ser His Ile Ile Phe Gly Val Asp Asp Gly Pro
1 5 10 15

Lys Thr Ile Glu Glu Ser Leu Ser Leu Ile Ser Glu Ala Tyr Arg Gln
20 25 30

Gly Val Arg Tyr Ile Val Ala Thr Ser His Arg Arg Lys Gly Met Phe
35 40 45

Glu Thr Pro Glu Lys Ile Ile Met Ile Asn Phe Leu Gln Leu Lys Glu
50 55 60

Ala Val Ala Glu Val Tyr Pro Glu Ile Arg Leu Cys Tyr Gly Ala Glu
65 70 75 80

Leu Tyr Tyr Ser Lys Asp Ile Leu Ser Lys Leu Glu Lys Lys Lys Val
85 90 95

Pro Thr Leu Asn Gly Ser Cys Tyr Ile Leu Leu Glu Phe Ser Thr Asp
100 105 110

Thr Pro Trp Lys Glu Ile Gln Glu Ala Val Asn Glu Met Thr Leu Leu
115 120 125

Gly Leu Thr Pro Val Leu Ala His Ile Glu Arg Tyr Asp Ala Leu Ala
130 135 140

Phe Gln Ser Glu Arg Val Glu Lys Leu Ile Asp Lys Gly Cys Tyr Thr
145 150 155 160

Gln Val Asn Ser Asn His Val Leu Lys Pro Ala Leu Ile Gly Glu Arg
165 170 175

Ala Lys Glu Phe Lys Lys Arg Thr Arg Tyr Phe Leu Glu Gln Asp Leu
180 185 190

Val His Cys Val Ala Ser Asp Met His Asn Leu Tyr Ser Arg Pro Pro
195 200 205

Phe Met Arg Glu Ala Tyr Gln Leu Val Lys Lys Glu Tyr Gly Glu Asp
210 215 220

Arg Ala Lys Ala Leu Phe Lys Lys Asn Pro Leu Leu Ile Leu Lys Asn
225 230 235 240

Gln Val Gln

<210> 16

<211> 459

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2E

<400> 16

Met Asn Ile Glu Ile Gly Tyr Arg Gln Thr Lys Leu Ala Leu Phe Asp
1 5 10 15

Met Ile Ala Val Thr Ile Ser Ala Ile Leu Thr Ser His Ile Pro Asn
20 25 30

Ala Asp Leu Asn Arg Ser Gly Ile Phe Ile Ile Met Met Val His Tyr
35 40 45

Phe Ala Phe Phe Ile Ser Arg Met Pro Val Glu Phe Glu Tyr Arg Gly
50 55 60

Asn Leu Ile Glu Phe Glu Lys Thr Phe Asn Tyr Ser Ile Ile Phe Val
65 70 75 80

Ile Phe Leu Met Ala Val Ser Phe Met Leu Glu Asn Asn Phe Ala Leu
85 90 95

Ser Arg Arg Gly Ala Val Tyr Phe Thr Leu Ile Asn Phe Val Leu Val
100 105 110

Tyr Leu Phe Asn Val Ile Ile Lys Gln Phe Lys Asp Ser Phe Leu Phe
115 120 125

Ser Thr Thr Tyr Gln Lys Lys Thr Ile Leu Ile Thr Thr Ala Glu Leu
 130 135 140

Trp Glu Asn Met Gln Val Leu Phe Glu Ser Asp Ile Leu Phe Gln Lys
 145 150 155 160

Asn Leu Val Ala Leu Val Ile Leu Gly Thr Glu Ile Asp Lys Ile Asn
 165 170 175

Leu Pro Leu Pro Leu Tyr Tyr Ser Val Glu Glu Ala Ile Gly Phe Ser
 180 185 190

Thr Arg Glu Val Val Asp Tyr Val Phe Ile Asn Leu Pro Ser Glu Tyr
 195 200 205

Phe Asp Leu Lys Gln Leu Val Ser Asp Phe Glu Leu Leu Gly Ile Asp
 210 215 220

Val Gly Val Asp Ile Asn Ser Phe Gly Phe Thr Val Leu Lys Asn Lys
 225 230 235 240

Lys Ile Gln Met Leu Gly Asp His Ser Ile Val Thr Phe Ser Thr Asn
 245 250 255

Phe Tyr Lys Pro Ser His Ile Trp Met Lys Arg Leu Leu Asp Ile Leu
 260 265 270

Gly Ala Val Val Gly Leu Ile Ile Ser Gly Ile Val Ser Ile Leu Leu
 275 280 285

Ile Pro Ile Ile Arg Arg Asp Gly Gly Pro Ala Ile Phe Ala Gln Lys
 290 295 300

Arg Val Gly Gln Asn Gly Arg Ile Phe Thr Phe Tyr Lys Phe Arg Ser
 305 310 315 320

Met Phe Val Asp Ala Glu Val Arg Lys Lys Glu Leu Met Ala Gln Asn
 325 330 335

Gln Met Gln Gly Gly Met Phe Lys Met Asp Asn Asp Pro Arg Ile Thr
 340 345 350

Pro Ile Gly His Phe Ile Arg Lys Thr Ser Leu Asp Glu Leu Pro Gln
 355 360 365

Phe Tyr Asn Val Leu Ile Gly Asp Met Ser Leu Val Gly Thr Arg Pro
 370 375 380

Pro Thr Val Asp Glu Phe Glu Lys Tyr Thr Pro Ser Gln Lys Arg Arg
 385 390 395 400

Leu Ser Phe Lys Pro Gly Ile Thr Gly Leu Trp Gln Val Ser Gly Arg
405 410 415

Ser Asp Ile Thr Asp Phe Asn Glu Val Val Arg Leu Asp Leu Thr Tyr
420 425 430

Ile Asp Asn Trp Thr Ile Trp Ser Asp Ile Lys Ile Leu Leu Lys Thr
435 440 445

Val Lys Val Val Leu Leu Arg Glu Gly Gly Gln
450 455

<210> 17

<211> 389

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS2F

<400> 17

Met Arg Thr Val Tyr Ile Ile Gly Ser Lys Gly Ile Pro Ala Lys Tyr
1 5 10 15

Gly Gly Phe Glu Thr Phe Val Glu Lys Leu Thr Glu Tyr Gln Lys Asp
20 25 30

Lys Ser Ile Asn Tyr Phe Val Ala Cys Thr Arg Glu Asn Ser Ala Lys
35 40 45

Ser Asp Ile Thr Gly Glu Val Phe Glu His Asn Gly Ala Thr Cys Phe
50 55 60

Asn Ile Asp Val Pro Asn Ile Gly Ser Ala Lys Ala Ile Leu Tyr Asp
65 70 75 80

Ile Met Ala Leu Lys Lys Ser Ile Glu Ile Ala Lys Asp Arg Asn Asp
85 90 95

Thr Ser Pro Ile Phe Tyr Ile Leu Ala Cys Arg Ile Gly Pro Phe Ile
100 105 110

Tyr Leu Phe Lys Lys Gln Ile Glu Ser Ile Gly Gly Gln Leu Phe Val
115 120 125

Asn	Pro	Asp	Gly	His	Glu	Trp	Leu	Arg	Glu	Lys	Trp	Ser	Tyr	Pro	Val
130					135						140				
Arg	Gln	Tyr	Trp	Lys	Phe	Ser	Glu	Ser	Leu	Met	Leu	Lys	Tyr	Ala	Asp
145					150					155				160	
Leu	Leu	Ile	Cys	Asp	Ser	Lys	Asn	Ile	Glu	Lys	Tyr	Ile	His	Glu	Asp
				165					170				175		
Tyr	Arg	Lys	Tyr	Ala	Pro	Glu	Thr	Ser	Tyr	Ile	Ala	Tyr	Gly	Thr	Asp
				180				185				190			
Leu	Asp	Lys	Ser	Arg	Leu	Ser	Pro	Thr	Asp	Ser	Val	Val	Arg	Glu	Trp
				195			200				205				
Tyr	Lys	Glu	Lys	Glu	Ile	Ser	Glu	Asn	Asp	Tyr	Tyr	Leu	Val	Val	Gly
				210		215				220					
Arg	Phe	Val	Pro	Glu	Asn	Asn	Tyr	Glu	Val	Met	Ile	Arg	Glu	Phe	Met
225					230				235				240		
Lys	Ser	Tyr	Ser	Arg	Lys	Asp	Phe	Val	Leu	Ile	Thr	Asn	Val	Glu	His
				245				250				255			
Asn	Ser	Phe	Tyr	Glu	Lys	Leu	Lys	Glu	Thr	Gly	Phe	Asp	Lys	Asp	
				260			265				270				
Lys	Arg	Ile	Lys	Phe	Val	Gly	Thr	Val	Tyr	Asn	Gln	Glu	Leu	Leu	Lys
				275			280				285				
Tyr	Ile	Arg	Glu	Asn	Ala	Phe	Ala	Tyr	Phe	His	Gly	His	Glu	Val	Gly
				290		295				300					
Gly	Thr	Asn	Pro	Ser	Leu	Leu	Glu	Ala	Leu	Ser	Ser	Thr	Lys	Leu	Asn
				305		310				315				320	
Leu	Leu	Leu	Asp	Val	Gly	Phe	Asn	Arg	Glu	Val	Gly	Glu	Gly	Ala	
				325				330				335			
Lys	Tyr	Trp	Asn	Lys	Asp	Asn	Leu	His	Arg	Val	Ile	Asp	Ser	Cys	Glu
				340			345				350				
Gln	Leu	Ser	Gln	Glu	Gln	Ile	Asn	Asp	Met	Asp	Ser	Leu	Ser	Thr	Lys
				355			360				365				
Gln	Val	Lys	Glu	Arg	Phe	Ser	Trp	Asp	Phe	Ile	Val	Asp	Glu	Tyr	Glu
				370		375				380					
Lys	Leu	Phe	Lys	Gly											
385															

<210> 18
<211> 385
<212> PRT
<213> *Streptococcus suis*
<220>
<221> misc_feature
<223> CPS2G
<400> 18

Met Lys Lys Ile Leu Tyr Leu His Ala Gly Ala Glu Leu Tyr Gly Ala
1 5 10 15

Asp Lys Val Leu Leu Glu Leu Ile Lys Gly Leu Asp Lys Asn Glu Phe
20 25 30

Glu Ala His Val Ile Leu Pro Asn Asp Gly Val Leu Val Pro Ala Leu
35 40 45

Arg Glu Val Gly Ala Gln Val Glu Val Ile Asn Tyr Pro Ile Leu Arg
50 55 60

Arg Lys Tyr Phe Asn Pro Lys Gly Ile Phe Asp Tyr Phe Ile Ser Tyr
65 70 75 80

His His Tyr Ser Lys Gln Ile Ala Gln Tyr Ala Ile Glu Asn Lys Val
85 90 95

Asp Ile Ile His Asn Asn Thr Thr Ala Val Leu Glu Gly Ile Tyr Leu
100 105 110

Lys Arg Lys Leu Lys Leu Pro Leu Leu Trp His Val His Glu Ile Ile
115 120 125

Val Lys Pro Lys Phe Ile Ser Asp Ser Ile Asn Phe Leu Met Gly Arg
130 135 140

Phe Ala Asp Lys Ile Val Thr Val Ser Gln Ala Val Ala Asn His Ile
145 150 155 160

Lys Gln Ser Pro His Ile Lys Asp Asp Gln Ile Ser Val Ile Tyr Asn
165 170 175

Gly Val Asp Asn Lys Val Phe Tyr Gln Ser Asp Ala Arg Ser Val Arg
180 185 190

Glu	Arg	Phe	Asp	Ile	Asp	Glu	Glu	Ala	Leu	Val	Ile	Gly	Met	Val	Gly
195						200						205			
Arg	Val	Asn	Ala	Trp	Lys	Gly	Gln	Gly	Asp	Phe	Leu	Glu	Ala	Val	Ala
210					215						220				
Pro	Ile	Leu	Glu	Gln	Asn	Pro	Lys	Ala	Ile	Ala	Phe	Ile	Ala	Gly	Ser
225					230				235				240		
Ala	Phe	Glu	Gly	Glu	Glu	Trp	Arg	Val	Val	Glu	Leu	Glu	Lys	Lys	Ile
	245					250						255			
Ser	Gln	Leu	Lys	Val	Ser	Ser	Gln	Val	Arg	Arg	Met	Asp	Tyr	Tyr	Ala
	260					265			270						
Asn	Thr	Thr	Glu	Leu	Tyr	Asn	Met	Phe	Asp	Ile	Phe	Val	Leu	Pro	Ser
	275					280				285					
Thr	Asn	Pro	Asp	Pro	Leu	Pro	Thr	Val	Val	Leu	Lys	Ala	Met	Ala	Cys
	290					295				300					
Gly	Lys	Pro	Val	Val	Gly	Tyr	Arg	His	Gly	Gly	Val	Cys	Glu	Met	Val
	305					310				315				320	
Lys	Glu	Gly	Val	Asn	Gly	Phe	Leu	Val	Thr	Pro	Asn	Ser	Pro	Leu	Asn
	325							330						335	
Leu	Ser	Lys	Val	Ile	Leu	Gln	Leu	Ser	Glu	Asn	Ile	Asn	Leu	Arg	Lys
	340							345						350	
Lys	Ile	Gly	Asn	Asn	Ser	Ile	Glu	Arg	Gln	Lys	Glu	His	Phe	Ser	Leu
	355						360					365			
Lys	Ser	Tyr	Val	Lys	Asn	Phe	Ser	Lys	Val	Tyr	Thr	Ser	Leu	Lys	Val
	370					375				380					
Tyr															
	385														
<210> 19															
<211> 456															
<212> PRT															
<213> Streptococcus suis															
<220>															
<221> misc_feature															

<223> cps2h

<400> 19

Met Lys Ile Ile Ser Phe Thr Met Val Asn Asn Glu Ser Glu Ile Ile
1 5 10 15

Glu Ser Phe Ile Arg Tyr Asn Tyr Asn Phe Ile Asp Glu Met Val Ile
20 25 30

Ile Asp Asn Gly Cys Thr Asp Asn Thr Met Gln Ile Ile Phe Asn Leu
35 40 45

Ile Lys Glu Gly Tyr Lys Ile Ser Val Tyr Asp Glu Ser Leu Glu Ala
50 55 60

Tyr Asn Gln Tyr Arg Leu Asp Asn Lys Tyr Leu Thr Lys Ile Ile Ala
65 70 75 80

Glu Lys Asn Pro Asp Leu Ile Ile Pro Leu Asp Ala Asp Glu Phe Leu
85 90 95

Thr Ala Asp Ser Asn Pro Arg Lys Leu Leu Glu Gln Leu Asp Leu Glu
100 105 110

Lys Ile His Tyr Val Asn Trp Gln Trp Phe Val Met Thr Lys Lys Asp
115 120 125

Asp Ile Asn Asp Ser Phe Ile Pro Arg Arg Met Gln Tyr Cys Phe Glu
130 135 140

Lys Pro Val Trp His His Ser Asp Gly Lys Pro Val Thr Lys Cys Ile
145 150 155 160

Ile Ser Ala Lys Tyr Tyr Lys Lys Met Asn Leu Lys Leu Ser Met Gly
165 170 175

His His Thr Val Phe Gly Asn Pro Asn Val Arg Ile Glu His His Asn
180 185 190

Asp Leu Lys Phe Ala His Tyr Arg Ala Ile Ser Gln Glu Gln Leu Ile
195 200 205

Tyr Lys Thr Ile Cys Tyr Thr Ile Arg Asp Ile Ala Thr Met Glu Asn
210 215 220

Asn Ile Glu Thr Ala Gln Arg Thr Asn Gln Met Ala Leu Ile Glu Ser
225 230 235 240

Gly Val Asp Met Trp Glu Thr Ala Arg Glu Ala Ser Tyr Ser Gly Tyr
245 250 255

Asp	Cys	Asn	Val	Ile	His	Ala	Pro	Ile	Asp	Leu	Ser	Phe	Cys	Lys	Glu
260								265					270		
Asn	Ile	Val	Ile	Lys	Tyr	Asn	Glu	Leu	Ser	Arg	Glu	Thr	Val	Ala	Glu
275						280				285					
Arg	Val	Met	Lys	Thr	Gly	Arg	Glu	Met	Ala	Val	Arg	Ala	Tyr	Asn	Val
290						295				300					
Glu	Arg	Lys	Gln	Lys	Glu	Lys	Lys	Phe	Leu	Lys	Pro	Ile	Ile	Phe	Val
305					310				315			320			
Leu	Asp	Gly	Leu	Lys	Gly	Asp	Glu	Tyr	Ile	His	Pro	Asn	Pro	Ser	Asn
	325						330				335				
His	Leu	Thr	Ile	Leu	Thr	Glu	Met	Tyr	Asn	Val	Arg	Gly	Leu	Leu	Thr
	340						345				350				
Asp	Asn	His	Gln	Ile	Lys	Phe	Leu	Lys	Val	Asn	Tyr	Arg	Leu	Ile	Ile
		355					360				365				
Thr	Pro	Asp	Phe	Ala	Lys	Phe	Leu	Pro	His	Glu	Phe	Ile	Val	Val	Pro
		370				375					380				
Asp	Thr	Leu	Asp	Ile	Glu	Gln	Val	Lys	Ser	Gln	Tyr	Val	Gly	Thr	Gly
		385			390				395			400			
Val	Asp	Leu	Ser	Lys	Ile	Ile	Ser	Leu	Lys	Glu	Tyr	Arg	Lys	Glu	Ile
		405						410				415			
Gly	Phe	Ile	Gly	Asn	Leu	Tyr	Ala	Leu	Leu	Gly	Phe	Val	Pro	Asn	Met
		420					425				430				
Leu	Asn	Arg	Ile	Tyr	Leu	Tyr	Ile	Gln	Arg	Asn	Gly	Ile	Ala	Asn	Thr
		435					440				445				
Ile	Ile	Lys	Ile	Lys	Ser	Arg	Leu								
		450			455										

<210> 20

<211> 410

<212> PRT

<213> Streptococcus suis.

<220>

<221> misc_feature

<223> CPS2I

<400> 20

Met Gln Ala Asp Arg Arg Lys Thr Phe Gly Lys Met Arg Ile Arg Ile
1 5 10 15

Asn Asn Leu Phe Phe Val Ala Ile Ala Phe Met Gly Ile Ile Ile Ser
20 25 30

Asn Ser Gln Val Val Leu Ala Ile Gly Lys Ala Ser Val Ile Gln Tyr
35 40 45

Leu Ser Tyr Leu Val Leu Ile Leu Cys Ile Val Asn Asp Leu Leu Lys
50 55 60

Asn Asn Lys His Ile Val Val Tyr Lys Leu Gly Tyr Leu Phe Leu Ile
65 70 75 80

Ile Phe Leu Phe Thr Ile Gly Ile Cys Gln Gln Ile Leu Pro Ile Thr
85 90 95

Thr Lys Ile Tyr Leu Ser Ile Ser Met Met Ile Ile Ser Val Leu Ala
100 105 110

Thr Leu Pro Ile Ser Leu Ile Lys Asp Ile Asp Asp Phe Arg Arg Ile
115 120 125

Ser Asn His Leu Leu Phe Ala Leu Phe Ile Thr Ser Ile Leu Gly Ile
130 135 140

Lys Met Gly Ala Thr Met Phe Thr Gly Ala Val Glu Gly Ile Gly Phe
145 150 155 160

Ser Gln Gly Phe Asn Gly Gly Leu Thr His Lys Asn Phe Phe Gly Ile
165 170 175

Thr Ile Leu Met Gly Phe Val Leu Thr Tyr Leu Ala Tyr Lys Tyr Gly
180 185 190

Ser Tyr Lys Arg Thr Asp Arg Phe Ile Leu Gly Leu Glu Leu Phe Leu
195 200 205

Ile Leu Ile Ser Asn Thr Arg Ser Val Tyr Leu Ile Leu Leu Phe
210 215 220

Leu Phe Leu Val Asn Leu Asp Lys Ile Lys Ile Glu Gln Arg Gln Trp
225 230 235 240

Ser Thr Leu Lys Tyr Ile Ser Met Leu Phe Cys Ala Ile Phe Leu Tyr
245 250 255

Tyr	Phe	Phe	Gly	Phe	Leu	Ile	Thr	His	Ser	Asp	Ser	Tyr	Ala	His	Arg
260								265					270		
Val	Asn	Gly	Leu	Ile	Asn	Phe	Phe	Glu	Tyr	Tyr	Arg	Asn	Asp	Trp	Phe
275								280					285		
His	Leu	Met	Phe	Gly	Ala	Ala	Asp	Leu	Ala	Tyr	Gly	Asp	Leu	Thr	Leu
290								295					300		
Asp	Tyr	Ala	Ile	Arg	Val	Arg	Arg	Val	Leu	Gly	Trp	Asn	Gly	Thr	Leu
305								310				315			320
Glu	Met	Pro	Leu	Leu	Ser	Ile	Met	Leu	Lys	Asn	Gly	Phe	Ile	Gly	Leu
									325			330			335
Val	Gly	Tyr	Gly	Ile	Val	Leu	Tyr	Lys	Leu	Tyr	Arg	Asn	Val	Arg	Ile
									340			345			350
Leu	Lys	Thr	Asp	Asn	Ile	Lys	Thr	Ile	Gly	Lys	Ser	Val	Phe	Ile	Ile
									355			360			365
Val	Val	Leu	Ser	Ala	Thr	Val	Glu	Asn	Tyr	Ile	Val	Asn	Leu	Ser	Phe
								370				375			380
Val	Phe	Met	Pro	Ile	Cys	Phe	Cys	Leu	Leu	Asn	Ser	Ile	Ser	Thr	Met
								385				390			400
Glu	Ser	Thr	Ile	Asn	Lys	Gln	Leu	Gln	Thr						
								405				410			

<210> 21

<211> 332

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS2J

<400> 21

Met	Glu	Lys	Val	Ser	Ile	Ile	Val	Pro	Ile	Phe	Asn	Thr	Glu	Lys	Tyr
1														15	

Leu	Arg	Glu	Cys	Leu	Asp	Ser	Ile	Ile	Ser	Gln	Ser	Tyr	Thr	Asn	Leu
													20		25

Glu Ile Leu Leu Ile Asp Asp Gly Ser Ser Asp Ser Ser Thr Asp Ile
 35 40 45

Cys Leu Glu Tyr Ala Glu Gln Asp Gly Arg Ile Lys Leu Phe Arg Leu
 50 55 60

Pro Asn Gly Gly Val Ser Asn Ala Arg Asn Tyr Gly Ile Lys Asn Ser
 65 70 75 80

Thr Ala Asn Tyr Ile Met Phe Val Asp Ser Asp Asp Ile Val Asp Gly
 85 90 95

Asn Ile Val Glu Ser Leu Tyr Thr Cys Leu Lys Glu Asn Asp Ser Asp
 100 105 110

Leu Ser Gly Gly Leu Leu Ala Thr Phe Asp Gly Asn Tyr Gln Glu Ser
 115 120 125

Glu Leu Gln Lys Cys Gln Ile Asp Leu Glu Glu Ile Lys Glu Val Arg
 130 135 140

Asp Leu Gly Asn Glu Asn Phe Pro Asn His Tyr Met Ser Gly Ile Phe
 145 150 155 160

Asn Ser Pro Cys Cys Lys Leu Tyr Lys Asn Ile Tyr Ile Asn Gln Gly
 165 170 175

Phe Asp Thr Glu Gln Trp Leu Gly Glu Asp Leu Leu Phe Asn Leu Asn
 180 185 190

Tyr Leu Lys Asn Ile Lys Lys Val Arg Tyr Val Asn Arg Asn Leu Tyr
 195 200 205

Phe Ala Arg Arg Ser Leu Gln Ser Thr Thr Asn Thr Phe Lys Tyr Asp
 210 215 220

Val Phe Ile Gln Leu Glu Asn Leu Glu Glu Lys Thr Phe Asp Leu Phe
 225 230 235 240

Val Lys Ile Phe Gly Gly Gln Tyr Glu Phe Ser Val Phe Lys Glu Thr
 245 250 255

Leu Gln Trp His Ile Ile Tyr Tyr Ser Leu Leu Met Phe Lys Asn Gly
 260 265 270

Asp Glu Ser Leu Pro Lys Lys Leu His Ile Phe Lys Tyr Leu Tyr Asn
 275 280 285

Arg His Ser Leu Asp Thr Leu Ser Ile Lys Arg Thr Ser Ser Val Phe
 290 295 300

Lys Arg Ile Cys Lys Leu Ile Val Ala Asn Asn Leu Phe Lys Ile Phe
305 310 315 320

Leu Asn Thr Leu Ile Arg Glu Glu Lys Asn Asn Asp
325 330

<210> 22

<211> 332

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2K

<400> 22

Met Ile Asn Ile Ser Ile Ile Val Pro Ile Tyr Asn Val Glu Gln Tyr
1 5 10 15

Leu Ser Lys Cys Ile Asn Ser Ile Val Asn Gln Thr Tyr Lys His Ile
20 25 30

Glu Ile Leu Leu Val Asn Asp Gly Ser Thr Asp Asn Ser Glu Glu Ile
35 40 45

Cys Leu Ala Tyr Ala Lys Lys Asp Ser Arg Ile Arg Tyr Phe Lys Lys
50 55 60

Glu Asn Gly Gly Leu Ser Asp Ala Arg Asn Tyr Gly Ile Ser Arg Ala
65 70 75 80

Lys Gly Asp Tyr Leu Ala Phe Ile Asp Ser Asp Asp Phe Ile His Ser
85 90 95

Glu Phe Ile Gln Arg Leu His Glu Ala Ile Glu Arg Glu Asn Ala Leu
100 105 110

Val Ala Val Ala Gly Tyr Asp Arg Val Asp Ala Ser Gly His Phe Leu
115 120 125

Thr Ala Glu Pro Leu Pro Thr Asn Gln Ala Val Leu Ser Gly Arg Asn
130 135 140

Val Cys Lys Lys Leu Leu Glu Ala Asp Gly His Arg Phe Val Val Ala
145 150 155 160

Trp	Asn	Lys	Leu	Tyr	Lys	Glu	Leu	Phe	Asp	Phe	Arg	Phe	Glu	Lys			
													165	170	175		
Gly	Lys	Ile	His	Glu	Asp	Glu	Tyr	Phe	Thr	Tyr	Arg	Leu	Leu	Tyr	Glu		
														180	185	190	
Leu	Glu	Lys	Val	Ala	Ile	Val	Lys	Glu	Cys	Leu	Tyr	Tyr	Tyr	Val	Asp		
														195	200	205	
Arg	Glu	Asn	Ser	Ile	Ile	Thr	Ser	Ser	Met	Thr	Asp	His	Arg	Phe	His		
														210	215	220	
Cys	Leu	Leu	Glu	Phe	Gln	Asn	Glu	Arg	Met	Asp	Phe	Tyr	Glu	Ser	Arg		
														225	230	235	240
Gly	Asp	Lys	Glu	Leu	Leu	Glu	Cys	Tyr	Arg	Ser	Phe	Leu	Ala	Phe			
														245	250	255	
Ala	Val	Leu	Phe	Leu	Gly	Lys	Tyr	Asn	His	Trp	Leu	Ser	Lys	Gln	Gln		
														260	265	270	
Lys	Lys	Leu	Gln	Thr	Leu	Phe	Arg	Ile	Val	Tyr	Lys	Gln	Leu	Lys	Gln		
														275	280	285	
Asn	Lys	Arg	Leu	Ala	Leu	Leu	Met	Asn	Ala	Tyr	Tyr	Leu	Val	Gly	Cys		
														290	295	300	
Leu	His	Leu	Asn	Phe	Ser	Val	Phe	Leu	Lys	Thr	Gly	Lys	Asp	Lys	Ile		
														305	310	315	320
Gln	Glu	Arg	Leu	Arg	Arg	Ser	Glu	Ser	Ser	Thr	Arg						
														325	330		

<210> 23
 <211> 467
 <212> PRT
 <213> *Streptococcus suis*
 <220>
 <221> misc_feature
 <223> CPS2O
 <220>
 <221> misc_feature

<222> (1)..(467)

<223> Xaa may be any amino acid

<400> 23

Met	Ser	Lys	Lys	Ser	Ile	Val	Val	Ser	Gly	Leu	Val	Tyr	Thr	Ile	Gly
1															15
Thr	Ile	Leu	Val	Gln	Gly	Leu	Ala	Phe	Ile	Thr	Leu	Pro	Ile	Tyr	Thr
				20				25						30	
Arg	Val	Ile	Ser	Gln	Glu	Val	Tyr	Gly	Gln	Phe	Ser	Leu	Tyr	Asn	Ser
				35			40							45	
Trp	Val	Gly	Leu	Val	Gly	Leu	Phe	Ile	Gly	Leu	Gln	Leu	Gly	Gly	Ala
				50			55							60	
Phe	Gly	Pro	Gly	Trp	Val	His	Phe	Arg	Glu	Lys	Phe	Asp	Asp	Phe	Val
				65			70			75				80	
Ser	Thr	Leu	Met	Val	Ser	Ser	Ile	Ala	Phe	Phe	Leu	Pro	Ile	Phe	Gly
				85				90						95	
Leu	Ser	Phe	Leu	Leu	Ser	Gln	Pro	Leu	Ser	Leu	Leu	Phe	Gly	Leu	Pro
				100				105						110	
Asp	Trp	Val	Val	Pro	Leu	Ile	Phe	Leu	Gln	Ser	Leu	Met	Ile	Val	Val
				115			120							125	
Gln	Gly	Phe	Phe	Thr	Thr	Tyr	Leu	Val	Gln	Arg	Gln	Gln	Ser	Met	Trp
				130			135							140	
Thr	Leu	Pro	Leu	Ser	Val	Leu	Ser	Ala	Val	Ile	Asn	Thr	Ala	Leu	Ser
				145			150			155				160	
Leu	Phe	Leu	Thr	Phe	Pro	Met	Glu	Asn	Asp	Phe	Ile	Ala	Arg	Val	Met
				165				170						175	
Ala	Asn	Pro	Ala	Thr	Thr	Gly	Val	Leu	Ala	Cys	Val	Ser	Xaa	Trp	Phe
				180			185							190	
Ser	Gln	Lys	Lys	Asn	Gly	Leu	His	Phe	Arg	Lys	Asp	Tyr	Leu	Arg	Tyr
				195			200							205	
Gly	Leu	Ser	Ile	Ser	Ile	Pro	Leu	Ile	Phe	His	Gly	Leu	Gly	His	Asn
				210			215							220	
Val	Leu	Asn	Gln	Phe	Asp	Arg	Ile	Met	Leu	Gly	Lys	Met	Leu	Thr	Leu
				225			230			235				240	

Ser Asp Val Ala Leu Tyr Ser Phe Gly Tyr Thr Leu Ala Ser Ile Leu
 245 250 255
 Gln Ile Val Phe Ser Ser Leu Asn Thr Val Trp Cys Pro Trp Tyr Phe
 260 265 270
 Glu Lys Lys Arg Gly Ala Asp Lys Asp Leu Leu Ser Tyr Val Arg Tyr
 275 280 285
 Tyr Leu Ala Ile Gly Leu Phe Val Thr Phe Gly Phe Leu Thr Ile Tyr
 290 295 300
 Pro Arg Leu Ala Met Leu Leu Gly Gly Ser Glu Tyr Arg Phe Ser Met
 305 310 315 320
 Gly Phe Ile Pro Met Ile Ile Val Gly Val Phe Phe Val Phe Leu Tyr
 325 330 335
 Ser Phe Pro Ala Asn Ile Gln Phe Tyr Ser Gly Asn Thr Lys Phe Leu
 340 345 350
 Pro Ile Gly Thr Phe Ile Ala Gly Val Leu Asn Ile Ser Val His Phe
 355 360 365
 Val Leu Ile Pro Thr Lys Asn Leu Trp Cys Cys Phe Ala Thr Thr Ala
 370 375 380
 Ser Tyr Leu Leu Leu Leu Val Leu His Tyr Phe Val Ala Lys Lys Lys
 385 390 395 400
 Tyr Ala Tyr Asp Glu Val Ala Ile Ser Thr Phe Val Lys Val Ile Ala
 405 410 415
 Leu Val Val Val Tyr Thr Gly Leu Met Thr Val Phe Val Gly Ser Ile
 420 425 430
 Trp Ile Arg Trp Ser Leu Gly Ile Ala Val Leu Val Val Tyr Ala Ile
 435 440 445
 Tyr Phe Arg Lys Glu Leu Thr Val Ala Leu Asn Thr Phe Arg Glu Lys
 450 455 460
 Arg Ser Lys
 465

<210> 24

<211> 338

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2P

<400> 24

Met Val Tyr Ile Ile Ala Glu Ile Gly Cys Asn His Asn Gly Asp Val
1 5 10 15

His Leu Ala Arg Lys Met Val Glu Val Ala Val Asp Cys Gly Val Asp
20 25 30

Ala Val Lys Phe Gln Thr Glu Lys Ala Asp Leu Leu Ile Ser Lys Tyr
35 40 45

Ala Pro Lys Ala Glu Tyr Gln Lys Ile Thr Thr Gly Glu Ser Asp Ser
50 55 60

Gln Leu Glu Met Thr Arg Arg Leu Glu Leu Ser Phe Glu Glu Tyr Leu
65 70 75 80

Asp Leu Arg Asp Tyr Cys Leu Glu Lys Gly Val Asp Val Phe Ser Thr
85 90 95

Pro Glu Asp Glu Glu Ser Leu Asp Phe Leu Ile Ser Thr Asp Met Pro
100 105 110

Val Tyr Lys Ile Pro Ser Gly Glu Ile Thr Asn Leu Pro Tyr Leu Glu
115 120 125

Lys Ile Gly Arg Gln Ala Lys Lys Val Ile Leu Ser Thr Gly Met Ala
130 135 140

Val Met Asp Glu Ile His Gln Ala Val Lys Ile Leu Gln Glu Asn Gly
145 150 155 160

Thr Thr Asp Ile Ser Ile Leu His Cys Thr Thr Glu Tyr Pro Thr Pro
165 170 175

Tyr Pro Ala Leu Asn Leu Asn Val Leu His Thr Leu Lys Lys Glu Phe
180 185 190

Pro Asn Leu Thr Ile Gly Tyr Ser Asp His Ser Val Gly Ser Glu Val
195 200 205

Pro Ile Ala Ala Ala Ala Met Gly Ala Glu Leu Ile Glu Lys His Phe
210 215 220

Thr Leu Asp Asn Glu Met Glu Gly Pro Asp His Lys Ala Ser Ala Thr
225 230 235 240
Pro Asp Ile Leu Ala Ala Leu Val Lys Gly Val Arg Ile Val Glu Gln
245 250 255
Ser Leu Gly Lys Phe Glu Lys Glu Pro Glu Glu Val Glu Val Arg Asn
260 265 270
Lys Ile Val Ala Glu Lys Ser Ile Val Ala Lys Lys Ala Ile Ala Lys
275 280 285
Gly Glu Val Phe Thr Glu Glu Asn Ile Thr Val Lys Arg Pro Gly Asn
290 295 300
Gly Ile Ser Pro Met Glu Trp Tyr Lys Val Leu Gly Gln Val Ser Glu
305 310 315 320
Gln Asp Phe Glu Glu Asp Gln Asn Ile Cys His Ser Ala Phe Glu Asn
325 330 335
Gln Met

<210> 25

<211> 170

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2Q

<400> 25

Met Lys Lys Ile Cys Phe Val Thr Gly Ser Arg Ala Glu Tyr Gly Ile
1 5 10 15

Met Arg Arg Leu Leu Ser Tyr Leu Gln Asp Asp Pro Glu Met Glu Leu
20 25 30

Asp Leu Val Val Ala Thr Met His Leu Glu Glu Lys Tyr Gly Met Thr
35 40 45

Val Lys Asp Ile Glu Ala Asp Lys Arg Arg Ile Val Lys Arg Ile Pro
50 55 60

Leu His Leu Thr Asp Thr Ser Lys Gln Thr Ile Val Lys Ser Leu Ala
65 70 75 80

Thr Leu Thr Glu Gln Leu Thr Val Leu Phe Glu Glu Val Gln Tyr Asp
85 90 95

Leu Val Leu Ile Leu Gly Asp Arg Tyr Glu Met Leu Pro Val Ala Asn
100 105 110

Ala Ala Leu Leu Tyr Asn Ile Pro Ile Cys His Ile His Gly Gly Glu
115 120 125

Lys Thr Met Gly Asn Phe Asp Glu Ser Ile Arg His Ala Ile Thr Lys
130 135 140

Met Ser His Leu His Leu Thr Ser Thr Asp Glu Phe Arg Asn Arg Val
145 150 155 160

Ile Gln Leu Gly Glu Asn Pro Thr Met Tyr
165 170

<210> 26

<211> 184

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS2R

<400> 26

Met Glu Leu Gly Ile Asp Phe Ala Glu Asp Tyr Tyr Val Val Leu Phe
1 5 10 15

His Pro Val Thr Leu Glu Asp Asn Thr Ala Glu Glu Gln Thr Gln Ala
20 25 30

Leu Leu Asp Ala Leu Lys Glu Asp Gly Ser Gln Cys Leu Ile Ile Gly
35 40 45

Ser Asn Ser Asp Thr His Ala Asp Lys Ile Met Glu Leu Met His Glu
50 55 60

Phe Val Lys Gln Asp Ser Asp Ser Tyr Ile Phe Thr Ser Leu Pro Thr
65 70 75 80

Arg Tyr Tyr His Ser Leu Val Lys His Ser Gln Gly Leu Ile Gly Asn
85 90 95

Ser Ser Ser Gly Leu Ile Glu Val Pro Ser Leu Gln Val Pro Thr Leu
100 105 110

Asn Ile Gly Asn Arg Gln Phe Gly Arg Leu Ser Gly Pro Ser Val Val
115 120 125

His Val Gly Thr Ser Lys Glu Ala Ile Val Gly Gly Leu Gly Gln Leu
130 135 140

Arg Asp Val Ile Asp Phe Thr Asn Pro Phe Glu Gln Pro Asp Ser Ala
145 150 155 160

Leu Gln Gly Tyr Arg Ala Ile Lys Glu Phe Leu Ser Val Gln Ala Ser
165 170 175

Thr Met Lys Glu Phe Tyr Asp Arg
180

<210> 27

<211> 208

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2S

<400> 27

Met Lys Lys Val Ala Phe Leu Gly Ala Gly Thr Phe Ser Asp Gly Val
1 5 10 15

Leu Pro Trp Leu Asp Arg Thr Arg Tyr Glu Leu Ile Gly Tyr Phe Glu
20 25 30

Asp Lys Pro Ile Ser Asp Tyr Arg Gly Tyr Pro Val Phe Gly Pro Leu
35 40 45

Gln Asp Val Leu Thr Tyr Leu Asp Asp Gly Lys Val Asp Ala Val Phe
50 55 60

Val Thr Ile Gly Asp Asn Val Lys Arg Lys Glu Ile Phe Asp Leu Leu
65 70 75 80

Ala Lys Asp His Tyr Asp Ala Leu Phe Asn Ile Ile Ser Glu Gln Ala
85 90 95

Asn Ile Phe Ser Pro Asp Ser Ile Lys Gly Arg Gly Val Phe Ile Gly
100 105 110

Phe Ser Ser Phe Val Gly Ala Asp Ser Tyr Val Tyr Asp Asn Cys Ile
115 120 125

Ile Asn Thr Gly Ala Ile Val Glu His His Thr Thr Val Glu Ala His
130 135 140

Cys Asn Ile Thr Pro Gly Val Thr Ile Asn Gly Leu Cys Arg Ile Gly
145 150 155 160

Glu Ser Thr Tyr Ile Gly Ser Gly Ser Thr Val Ile Gln Cys Ile Glu
165 170 175

Ile Ala Pro Tyr Thr Thr Leu Gly Ala Gly Thr Val Val Leu Lys Ser
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Leu Thr Glu Ser Gly Thr Tyr Val Gly Val Pro Ala Arg Lys Ile Lys
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<211> 410

<212> PRT

<213> *Streptococcus suis*

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<223> CPS2T

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Val	Val	Ser	Phe	Thr	Lys	Val	Asp	Lys	Ser	Pro	Thr	Leu	Phe	Ser	Thr
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Tyr	Ile	Ser	Ser	Lys	Gln	Ala	Tyr	Leu	Ala	Asp	Lys	Thr	Tyr	Phe	Ser
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Asp	Asp	His	Phe	Asp	Phe	Thr	Gly	Val	Ile	Gly	Arg	Ile	Tyr	Phe	Asp
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Tyr	Gln	Arg	Arg	Glu	Gln	Gln	Asn	Lys	Pro	Phe	Tyr	Lys	Arg	Glu	Leu
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Ser	Arg	Leu	Leu	Ala	Leu	Leu	Asp	Gly	Phe	Asp	Asn	Ile	Ser	Ile	
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Gly	Gly	Met	Thr	Ala	Ser	Thr	Ser	Leu	Glu	Asn	Gln	Gly	Leu	Phe	Leu
					275				280				285		
Ala	Thr	Pro	Ile	Lys	Lys	Val	Leu	Leu	Ser	Leu	Gly	Val	Asn	Asp	Leu
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Ile Thr Asp Tyr Pro Leu His Met Ile Glu Asp Thr Ile Arg Gln Leu
305 310 315 320

Met Glu Ser Leu Val Ser Lys Ala Glu Gln Val Glu Val Thr Thr Ile
325 330 335

Ala Tyr Thr Leu Phe Arg Asp Ser Val Ser Asn Glu Glu Thr Val Gln
340 345 350

Leu Asn Asp Val Ile Val Gln Ser Ala Ser Glu Leu Gly Ile Ser Val
355 360 365

Ile Asp Leu Asn Glu Val Val Glu Lys Glu Ala Met Leu Asp Tyr Gln
370 375 380

Tyr Thr Asn Asp Gly Leu His Phe Asn Gln Ile Gly Gln Glu Arg Val
385 390 395 400

Asn Gln Leu Ile Leu Thr Ser Leu Thr Arg
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<211> 6992

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<220>

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<213> Streptococcus suis

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<223> CPS1E

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Arg Gln Thr Lys Leu Ala Leu Phe Asp Met Ile Ala Val Ala Ile Ser
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Ala Ile Leu Thr Ser His Ile Pro Asn Ala Asp Leu Asn Arg Ser Gly
20 25 30

Ile Phe Ile Ile Met Met Val His Tyr Phe Ala Phe Phe Ile Ser Arg
35 40 45

Met Pro Val Glu Phe Glu Tyr Arg Gly Asn Leu Ile Glu Phe Glu Lys
50 55 60

Thr Phe Asn Tyr Ser Ile Ile Phe Ala Ile Phe Leu Thr Ala Val Ser
65 70 75 80

Phe Leu Leu Glu Asn Asn Phe Ala Leu Ser Arg Arg Gly Ala Val Tyr
85 90 95

Phe Thr Leu Ile Asn Phe Val Leu Val Tyr Leu Phe Asn Val Ile Ile
100 105 110

Lys Gln Phe Lys Asp Ser Phe Leu Phe Ser Thr Ile Tyr Gln Lys Lys
115 120 125

Thr Ile Leu Ile Thr Thr Ala Glu Arg Trp Glu Asn Met Gln Val Leu
 130 135 140

Phe Glu Ser His Lys Gln Ile Gln Lys Asn Leu Val Ala Leu Val Val
 145 150 155 160

Leu Gly Thr Glu Ile Asp Lys Ile Asn Leu Ser Leu Pro Leu Tyr Tyr
 165 170 175

Ser Val Glu Glu Ala Ile Glu Phe Ser Thr Arg Glu Val Val Asp His
 180 185 190

Val Phe Ile Asn Leu Pro Ser Glu Phe Leu Asp Val Lys Gln Phe Val
 195 200 205

Ser Asp Phe Glu Leu Leu Gly Ile Asp Val Ser Val Asp Ile Asn Ser
 210 215 220

Phe Gly Phe Thr Ala Leu Lys Asn Lys Lys Ile Gln Leu Leu Gly Asp
 225 230 235 240

His Ser Ile Val Thr Phe Ser Thr Asn Phe Tyr Lys Pro Ser His Ile
 245 250 255

Met Met Lys Arg Leu Leu Asp Ile Leu Gly Ala Val Val Gly Leu Ile
 260 265 270

Ile Cys Gly Ile Val Ser Ile Leu Leu Val Pro Ile Ile Arg Arg Asp
 275 280 285

Gly Gly Pro Ala Ile Phe Ala Gln Lys Arg Val Gly Gln Asn Gly Arg
 290 295 300

Ile Phe Thr Phe Tyr Lys Phe Arg Ser Met Tyr Val Asp Ala Glu Glu
 305 310 315 320

Arg Lys Lys Asp Leu Leu Ser Gln Asn Gln Met Gln Gly Trp Val Cys
 325 330 335

Phe Lys Met Gly Lys Thr Ile Leu Glu Leu Leu Gln Leu Asp Ile Ser
 340 345 350

Tyr Ala Lys Thr Ser Leu Asp Glu Leu Pro Gln Phe Tyr Asn Val Leu
 355 360 365

Ile Gly Asp Met Ser Leu Val Gly Thr Arg Pro Pro Thr Val Asp Glu
 370 375 380

Phe Glu Lys Tyr Thr Pro Gly Gln Lys Arg Arg Leu Ser Phe Lys Pro
 385 390 395 400

Gly Ile Thr Gly Leu Trp Gln Val Ser Gly Arg Ser Asn Ile Thr Asp
405 410 415

Phe Asp Asp Val Val Arg Leu Asp Leu Ala Tyr Ile Asp Asn Trp Thr
420 425 430

Ile Trp Ser Asp Ile Lys Ile Leu Leu Lys Thr Val Lys Val Val Leu
435 440 445

Leu Arg Glu Gly Ser Lys
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<211> 149

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<213> Streptococcus suis

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<223> CPS1F

<400> 31

Met Lys Val Cys Leu Val Gly Ser Ser Gly Gly His Leu Thr His Leu
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Tyr Leu Leu Lys Pro Phe Trp Lys Glu Glu Glu Arg Phe Trp Val Thr
20 25 30

Phe Asp Lys Glu Asp Ala Arg Ser Leu Leu Lys Asn Glu Lys Met Tyr
35 40 45

Pro Cys Tyr Phe Pro Thr Asn Arg Asn Leu Ile Asn Leu Val Lys Asn
50 55 60

Thr Phe Leu Ala Phe Lys Ile Leu Arg Asp Glu Lys Pro Asp Val Ile
65 70 75 80

Ile Ser Ser Gly Ala Ala Val Ala Val Pro Phe Phe Tyr Ile Gly Lys
85 90 95

Leu Phe Gly Ala Lys Thr Ile Tyr Ile Glu Val Phe Asp Arg Val Asn
100 105 110

Lys Ser Thr Leu Thr Gly Lys Leu Val Tyr Pro Val Thr Asp Ile Phe
115 120 125

Ile Val Gln Trp Glu Glu Met Lys Lys Val Tyr Pro Lys Ser Ile Asn
130 135 140

Leu Gly Ser Ile Phe
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<210> 32

<211> 164

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS1G

<400> 32

Met Ile Phe Val Thr Val Gly Thr His Glu Gln Gln Phe Asn Arg Leu
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Ile Lys Glu Ile Asp Leu Leu Lys Lys Asn Gly Ser Ile Thr Asp Glu
20 25 30

Ile Phe Ile Gln Thr Gly Tyr Ser Asp Tyr Ile Pro Glu Tyr Cys Lys
35 40 45

Tyr Lys Lys Phe Leu Ser Tyr Lys Glu Met Glu Gln Tyr Ile Asn Lys
50 55 60

Ser Glu Val Val Ile Cys His Gly Gly Pro Ala Thr Phe Met Asn Ser
65 70 75 80

Leu Ser Lys Gly Lys Lys Gln Leu Leu Phe Pro Arg Gln Lys Lys Tyr
85 90 95

Gly Glu His Val Asn Asp His Gln Val Glu Phe Val Arg Arg Ile Leu
100 105 110

Gln Asp Asn Asn Ile Leu Phe Ile Glu Asn Ile Asp Asp Leu Phe Glu
115 120 125

Lys Ile Ile Glu Val Ser Lys Gln Thr Asn Phe Thr Ser Asn Asn Asn
130 135 140

Phe Phe Cys Glu Arg Leu Lys Gln Ile Val Glu Lys Phe Asn Glu Asp
145 150 155 160

Gln Glu Asn Glu

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<211> 388

<212> PRT

<213> *Streptococcus suis*

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<221> misc_feature

<223> CPS1H

<400> 33

Met Phe Lys Leu Phe Lys Tyr Asp Pro Glu Tyr Phe Ile Phe Lys Tyr
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Phe Trp Leu Ile Ile Phe Ile Pro Glu Gln Lys Tyr Val Phe Leu Leu
20 25 30

Ile Phe Met Asn Leu Ile Leu Phe His Ile Lys Phe Leu Lys Thr Lys
35 40 45

Leu Ile Leu Lys Asn Glu Ile Leu Leu Phe Leu Leu Trp Ser Ile Leu
50 55 60

Cys Phe Val Ser Val Val Thr Ser Met Phe Val Glu Ile Asn Phe Glu
65 70 75 80

Arg Leu Phe Ala Asp Phe Thr Ala Pro Ile Ile Trp Ile Ile Ala Ile
85 90 95

Met Tyr Tyr Asn Leu Tyr Ser Phe Ile Asn Ile Asp Tyr Lys Lys Leu
100 105 110

Lys Asn Ser Ile Phe Phe Ser Phe Leu Val Leu Leu Gly Ile Ser Ala
115 120 125

Leu Tyr Ile Ile Gln Asn Gly Lys Asp Ile Val Phe Leu Asp Arg His
130 135 140

Leu Ile Gly Leu Asp Tyr Leu Ile Thr Gly Val Lys Thr Arg Leu Val
145 150 155 160

Gly Phe Met Asn Tyr Pro Thr Leu Asn Thr Thr Thr Ile Ile Val Ser
 165 170 175
 Ile Pro Leu Ile Phe Ala Leu Ile Lys Asn Lys Met Gln Gln Phe Phe
 180 185 190
 Phe Leu Cys Leu Ala Phe Ile Pro Ile Tyr Leu Ser Gly Ser Arg Ile
 195 200 205
 Gly Ser Leu Ser Leu Ala Ile Leu Ile Ile Cys Leu Leu Trp Arg Tyr
 210 215 220
 Ile Gly Gly Lys Phe Ala Trp Ile Lys Lys Leu Ile Val Ile Phe Val
 225 230 235 240
 Ile Leu Leu Ile Ile Leu Asn Thr Glu Leu Leu Tyr His Glu Ile Leu
 245 250 255
 Ala Val Tyr Asn Ser Arg Glu Ser Ser Asn Glu Ala Arg Phe Ile Ile
 260 265 270
 Tyr Gln Gly Ser Ile Asp Lys Val Leu Glu Asn Asn Ile Leu Phe Gly
 275 280 285
 Tyr Gly Ile Ser Glu Tyr Ser Val Thr Gly Thr Trp Leu Gly Ser His
 290 295 300
 Ser Gly Tyr Ile Ser Phe Phe Tyr Lys Ser Gly Ile Val Gly Leu Ile
 305 310 315 320
 Leu Leu Met Phe Ser Phe Phe Tyr Val Ile Lys Lys Ser Tyr Gly Val
 325 330 335
 Asn Gly Glu Thr Ala Leu Phe Tyr Phe Thr Ser Leu Ala Ile Phe Phe
 340 345 350
 Ile Tyr Glu Thr Ile Asp Pro Ile Ile Ile Ile Leu Val Leu Phe Phe
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 Thr Lys Asn Glu
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<210> 34

<211> 322

<212> PRT

<213> Streptococcus suis

<220>

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<223> CPS1I

<400> 34

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					20				25				30		
Leu	Glu	Val	Ile	Leu	Val	Asn	Asp	Gly	Ser	Thr	Asp	Asp	Ser	Glu	Lys
					35			40					45		
Ile	Cys	Leu	Asn	Tyr	Met	Lys	Asn	Asp	Gly	Arg	Ile	Lys	Tyr	Tyr	Lys
					50			55			60				
Lys	Ile	Asn	Gly	Gly	Leu	Ala	Asp	Ala	Arg	Asn	Phe	Gly	Leu	Glu	His
					65			70			75		80		
Ala	Thr	Gly	Lys	Tyr	Ile	Ala	Phe	Val	Asp	Ser	Asp	Asp	Tyr	Ile	Glu
					85			90					95		
Val	Ala	Met	Phe	Glu	Arg	Met	His	Asp	Asn	Ile	Thr	Glu	Tyr	Asn	Ala
					100			105				110			
Asp	Ile	Ala	Glu	Ile	Asp	Phe	Cys	Leu	Val	Asp	Glu	Asn	Gly	Tyr	Thr
					115			120			125				
Lys	Lys	Lys	Arg	Asn	Ser	Asn	Phe	His	Val	Leu	Thr	Arg	Glu	Glu	Thr
					130			135			140				
Val	Lys	Glu	Phe	Leu	Ser	Gly	Ser	Asn	Ile	Glu	Asn	Asn	Val	Trp	Cys
					145			150			155		160		
Lys	Leu	Tyr	Ser	Arg	Asp	Ile	Ile	Lys	Asp	Ile	Lys	Phe	Gln	Ile	Asn
					165			170			175				
Asn	Arg	Ser	Ile	Gly	Glu	Asp	Leu	Leu	Phe	Asn	Leu	Glu	Val	Leu	Asn
					180			185			190				
Asn	Val	Thr	Arg	Val	Val	Asp	Thr	Arg	Glu	Tyr	Tyr	Tyr	Asn	Tyr	
					195			200			205				
Val	Ile	Arg	Asn	Ser	Ser	Leu	Ile	Asn	Gln	Lys	Phe	Ser	Ile	Asn	Asn
					210			215			220				

Ile Asp Leu Val Thr Arg Leu Glu Asn Tyr Pro Phe Lys Leu Lys Arg
225 230 235 240
Glu Phe Ser His Tyr Phe Asp Ala Lys Val Ile Lys Glu Lys Val Lys
245 250 255
Cys Leu Asn Lys Met Tyr Ser Thr Asp Cys Leu Asp Asn Glu Phe Leu
260 265 270
Pro Ile Leu Glu Ser Tyr Arg Lys Glu Ile Arg Arg Tyr Pro Phe Ile
275 280 285
Lys Ala Lys Arg Tyr Leu Ser Arg Lys His Leu Val Thr Leu Tyr Leu
290 295 300
Met Lys Phe Ser Pro Lys Leu Tyr Val Met Leu Tyr Lys Lys Phe Gln
305 310 315 320
Lys Gln

<210> 35
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<220>
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<223> CPS1J
<400> 35

Met Asp Lys Ile Ser Val Ile Val Pro Val Tyr Asn Val Asp Lys Tyr
1 5 10 15
Leu Ser Ser Cys Ile Glu Ser Ile Ile Asn Gln Asn Tyr Lys Asn Ile
20 25 30
Glu Ile Leu Leu Ile Asp Asp Gly Ser Val Asp Asp Ser Ala Lys Ile
35 40 45
Cys Lys Glu Tyr Glu Lys Asp Lys Arg Val Lys Ile Phe Phe Thr Asn
50 55 60

His	Ser	Gly	Val	Ser	Asn	Ala	Arg	Asn	His	Gly	Ile	Lys	Arg	Ser	Thr
65				70					75				80		
Ala	Glu	Tyr	Ile	Met	Phe	Val	Asp	Ser	Asp	Asp	Val	Val	Asp	Ser	Arg
	85					90							95		
Leu	Val	Glu	Lys	Leu	Tyr	Phe	Asn	Ile	Ile	Lys	Ser	Arg	Ser	Asp	Leu
		100					105						110		
Ser	Gly	Cys	Leu	Tyr	Ala	Thr	Phe	Ser	Glu	Asn	Ile	Asn	Asn	Phe	Glu
		115				120						125			
Val	Asn	Asn	Pro	Asn	Ile	Asp	Phe	Glu	Ala	Ile	Asn	Thr	Val	Gln	Asp
		130				135					140				
Met	Gly	Glu	Lys	Asn	Phe	Met	Asn	Leu	Tyr	Ile	Asn	Asn	Ile	Phe	Ser
		145				150				155			160		
Thr	Pro	Val	Cys	Lys	Leu	Tyr	Lys	Lys	Arg	Tyr	Ile	Thr	Asp	Leu	Phe
			165					170				175			
Gln	Glu	Asn	Gln	Trp	Leu	Gly	Glu	Asp	Leu	Leu	Phe	Asn	Leu	His	Tyr
			180					185				190			
Leu	Lys	Asn	Ile	Asp	Arg	Val	Ser	Tyr	Leu	Thr	Glu	His	Leu	Tyr	Phe
			195				200				205				
Tyr	Arg	Arg	Gly	Ile	Leu	Ser	Thr	Val	Asn	Ser	Phe	Lys	Glu	Gly	Val
			210			215					220				
Phe	Leu	Gln	Leu	Glu	Asn	Leu	Gln	Lys	Gln	Val	Ile	Val	Leu	Phe	Lys
		225				230				235			240		
Gln	Ile	Tyr	Gly	Glu	Asp	Phe	Asp	Val	Ser	Ile	Val	Lys	Asp	Thr	Ile
			245					250				255			
Arg	Trp	Gln	Val	Phe	Tyr	Tyr	Ser	Leu	Leu	Met	Phe	Lys	Tyr	Gly	Lys
			260				265				270				
Gln	Ser	Ile	Phe	Asp	Lys	Phe	Leu	Ile	Phe	Arg	Asn	Leu	Tyr	Lys	Lys
			275				280				285				
Tyr	Tyr	Phe	Asn	Leu	Leu	Lys	Val	Ser	Asn	Lys	Asn	Ser	Leu	Ser	Lys
			290			295				300					
Asn	Phe	Cys	Ile	Arg	Ile	Val	Ser	Asn	Lys	Val	Phe	Lys	Lys	Ile	Leu
			305			310				315			320		
Trp	Leu														

<210> 36
<211> 278
<212> PRT
<213> *Streptococcus suis*
<220>
<221> misc_feature
<223> CPS1K
<400> 36

Met Asp Thr Ile Ser Lys Ile Ser Ile Ile Val Pro Ile Tyr Asn Val
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Glu Lys Tyr Leu Ser Lys Cys Ile Asp Ser Ile Val Asn Gln Thr Tyr
20 25 30

Lys His Ile Glu Ile Leu Leu Val Asn Asp Gly Ser Thr Asp Asn Ser
35 40 45

Glu Glu Ile Cys Leu Ala Tyr Ala Lys Lys Asp Ser Arg Ile Arg Tyr
50 55 60

Phe Lys Lys Glu Asn Gly Gly Leu Ser Asp Ala Arg Asn Tyr Gly Ile
65 70 75 80

Ser Arg Ala Lys Gly Asp Tyr Leu Ala Phe Ile Asp Ser Asp Asp Phe
85 90 95

Ile His Ser Glu Phe Ile Gln Arg Leu His Glu Ala Ile Glu Arg Glu
100 105 110

Asn Ala Leu Val Ala Val Ala Gly Tyr Asp Arg Val Asp Ala Ser Gly
115 120 125

His Phe Leu Thr Ala Glu Pro Leu Pro Thr Asn Gln Ala Val Leu Ser
130 135 140

Gly Arg Asn Val Cys Lys Lys Leu Leu Glu Ala Asp Gly His Arg Phe
145 150 155 160

Val Val Ala Cys Asn Lys Leu Tyr Lys Lys Glu Leu Phe Glu Asp Phe
165 170 175

Arg Phe Glu Lys Gly Lys Ile His Glu Asp Glu Tyr Phe Thr Tyr Arg
180 185 190

Leu Leu Tyr Glu Leu Glu Lys Val Ala Ile Val Lys Glu Cys Leu Tyr
195 200 205

Tyr Tyr Val Asp Arg Glu Asn Ser Ile Thr Thr Ser Ser Met Thr Asp
210 215 220

His Arg Phe His Cys Leu Leu Glu Phe Gln Asn Glu Arg Met Asp Phe
225 230 235 240

Tyr Glu Ser Arg Gly Asp Lys Glu Leu Leu Leu Glu Cys Tyr Arg Ser
245 250 255

Phe Leu Ala Phe Ala Val Leu Phe Leu Gly Lys Tyr Asn His Trp Leu
260 265 270

Ser Lys Gln Gln Lys Lys
275

<210> 37

<211> 4519

<212> DNA

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS9

<400> 37

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aagtttatcc tgaaatacga ttgtgctatg gtgctgaatt gtattatagt aaagatatat 180
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agttcagtag tgataactcct tggaaagaga ttcaagaagc agtgaacgaa gtgacgctac 300
ttgggctaac tcccgtactt gcccatatag aacgatatga cgccctagcg tttcatgcag 360
agagagtaga agagttaatt gacaagggat gctatactca ggtaaatagt aatcatgtgc 420
tgaagccac tttaatttgtt gatcgagcaa aagaattaa aaaacgtact cggtatttt 480
tagagcagga tttagtacat tgtgttgcta gcgatatgca taatttatct agtagacctc 540

cgtttatgag ggaggcttat aagttgctaa cagaggaatt tggcaaagat aaagcgaaaag 600
cgttgctaaa aaagaatcct cttatgctat taaaaaacca ggcgatttaa actggttact 660
ctagattgtg gagagaaaaa tggatttagg aactgttact gataaactgt tagaacgcaa 720
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<210> 38

<211> 215

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS9D

<400> 38

Ala Tyr Arg Gln Gly Val Arg Tyr Ile Val Ala Thr Ser His Arg Arg
1 5 10 15

Lys	Gly	Met	Phe	Glu	Thr	Pro	Glu	Lys	Val	Ile	Met	Thr	Asn	Phe	Leu
20								25						30	
Gln	Phe	Lys	Asp	Ala	Val	Ala	Glu	Val	Tyr	Pro	Glu	Ile	Arg	Leu	Cys
35							40					45			
Tyr	Gly	Ala	Glu	Leu	Tyr	Tyr	Ser	Lys	Asp	Ile	Leu	Ser	Lys	Leu	Glu
50						55					60				
Lys	Lys	Lys	Val	Pro	Thr	Leu	Asn	Gly	Ser	Arg	Tyr	Ile	Leu	Leu	Glu
65						70				75			80		
Phe	Ser	Ser	Asp	Thr	Pro	Trp	Lys	Glu	Ile	Gln	Glu	Ala	Val	Asn	Glu
85							90					95			
Val	Thr	Leu	Leu	Gly	Leu	Thr	Pro	Val	Leu	Ala	His	Ile	Glu	Arg	Tyr
100							105					110			
Asp	Ala	Leu	Ala	Phe	His	Ala	Glu	Arg	Val	Glu	Glu	Leu	Ile	Asp	Lys
115							120					125			
Gly	Cys	Tyr	Thr	Gln	Val	Asn	Ser	Asn	His	Val	Leu	Lys	Pro	Thr	Leu
130						135					140				
Ile	Gly	Asp	Arg	Ala	Lys	Glu	Phe	Lys	Lys	Arg	Thr	Arg	Tyr	Phe	Leu
145						150				155			160		
Glu	Gln	Asp	Leu	Val	His	Cys	Val	Ala	Ser	Asp	Met	His	Asn	Leu	Ser
165							170					175			
Ser	Arg	Pro	Pro	Phe	Met	Arg	Glu	Ala	Tyr	Lys	Leu	Leu	Thr	Glu	Glu
180							185					190			
Phe	Gly	Lys	Asp	Lys	Ala	Lys	Ala	Leu	Leu	Lys	Lys	Asn	Pro	Leu	Met
195						200						205			
Leu	Leu	Lys	Asn	Gln	Ala	Ile									
210						215									

<210> 39

<211> 608

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS9E

<400> 39

Met Asp Leu Gly Thr Val Thr Asp Lys Leu Leu Glu Arg Asn Ser Lys
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Arg Leu Ile Leu Val Cys Met Asp Thr Cys Leu Leu Ile Val Ser Met
20 25 30

Ile Leu Ser Arg Leu Phe Leu Asp Val Ile Ile Asp Ile Pro Asp Glu
35 40 45

Arg Phe Ile Leu Ala Val Leu Phe Val Ser Ile Leu Tyr Leu Ile Leu
50 55 60

Ser Phe Arg Leu Lys Val Phe Ser Leu Ile Thr Arg Tyr Thr Gly Tyr
65 70 75 80

Gln Ser Tyr Val Lys Ile Gly Leu Ser Leu Ile Ser Ala His Ser Leu
85 90 95

Phe Leu Ile Ile Ser Met Val Leu Trp Gln Ala Phe Ser Tyr Arg Phe
100 105 110

Ile Leu Val Ser Leu Phe Leu Ser Tyr Val Met Leu Ile Thr Pro Arg
115 120 125

Ile Val Trp Lys Val Leu His Glu Thr Arg Lys Asn Ala Ile Arg Lys
130 135 140

Lys Asp Ser Pro Leu Arg Ile Leu Val Val Gly Ala Gly Asp Gly Gly
145 150 155 160

Asn Ile Phe Ile Asn Thr Val Lys Asp Arg Lys Leu Asn Phe Glu Ile
165 170 175

Val Gly Ile Val Asp Arg Asp Pro Asn Lys Leu Gly Thr Phe Ile Arg
180 185 190

Thr Ala Lys Val Leu Gly Asn Arg Asn Asp Ile Pro Arg Leu Val Glu
195 200 205

Glu Leu Ala Val Asp Gln Val Thr Ile Ala Ile Pro Ser Leu Asn Gly
210 215 220

Lys Glu Arg Glu Lys Ile Val Glu Ile Cys Asn Thr Thr Gly Val Thr
225 230 235 240

Val Asn Asn Met Pro Ser Ile Glu Asp Ile Met Ala Gly Asn Met Ser
245 250 255

Val Ser Ala Phe Gln Glu Ile Asp Val Ala Asp Leu Leu Gly Arg Pro
 260 265 270
 Glu Val Val Leu Asp Gln Asp Glu Leu Asn Gln Phe Phe Gln Gly Lys
 275 280 285
 Thr Ile Leu Val Thr Gly Ala Gly Gly Ser Ile Gly Ser Glu Leu Cys
 290 295 300
 Arg Gln Ile Ala Lys Phe Thr Pro Lys Arg Leu Leu Leu Leu Gly His
 305 310 315 320
 Gly Glu Asn Ser Ile Tyr Leu Ile His Arg Glu Leu Leu Glu Lys Tyr
 325 330 335
 Gln Gly Lys Ile Glu Leu Val Pro Leu Ile Ala Asp Ile Gln Asp Arg
 340 345 350
 Glu Leu Ile Phe Ser Ile Met Ala Glu Tyr Gln Pro Asp Val Val Tyr
 355 360 365
 His Ala Ala Ala His Lys His Val Pro Leu Met Glu Tyr Asn Pro His
 370 375 380
 Glu Ala Val Lys Asn Asn Ile Phe Gly Thr Lys Asn Val Ala Glu Ala
 385 390 395 400
 Ala Lys Thr Ala Lys Val Ala Lys Phe Val Met Val Ser Thr Asp Lys
 405 410 415
 Ala Val Asn Pro Pro Asn Val Met Gly Ala Thr Lys Arg Val Ala Glu
 420 425 430
 Met Ile Val Thr Gly Leu Asn Glu Pro Gly Gln Thr Gln Phe Ala Ala
 435 440 445
 Val Arg Phe Gly Asn Val Leu Gly Ser Arg Gly Ser Val Val Pro Leu
 450 455 460
 Phe Lys Glu Gln Ile Arg Lys Gly Gly Pro Val Thr Val Thr Asp Phe
 465 470 475 480
 Arg Met Thr Arg Tyr Phe Met Thr Ile Pro Glu Ala Ser Arg Leu Val
 485 490 495
 Ile Gln Ala Gly His Leu Ala Lys Gly Gly Glu Ile Phe Val Leu Asp
 500 505 510
 Met Gly Glu Pro Val Gln Ile Leu Glu Leu Ala Arg Lys Val Ile Leu
 515 520 525

Leu Ser Gly His Thr Glu Glu Glu Ile Gly Ile Val Glu Ser Gly Ile
530 535 540

Arg Pro Gly Glu Lys Leu Tyr Glu Glu Leu Leu Ser Thr Glu Glu Arg
545 550 555 560

Val Ser Glu Gln Ile His Glu Lys Ile Phe Val Gly Arg Val Thr Asn
565 570 575

Lys Gln Ser Asp Ile Val Asn Ser Phe Ile Asn Gly Leu Leu Gln Lys
580 585 590

Asp Arg Asn Glu Leu Lys Asn Met Leu Ile Glu Phe Ala Lys Gln Glu
595 600 605

<210> 40

<211> 200

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS9F

<400> 40

Met Tyr Pro Ile Cys Lys Arg Ile Leu Ala Ile Ile Ile Ser Gly Ile
1 5 10 15

Ala Ile Val Val Leu Ser Pro Ile Leu Leu Leu Ile Ala Leu Ala Ile
20 25 30

Lys Leu Asp Ser Lys Gly Pro Val Leu Phe Lys Gln Lys Arg Val Gly
35 40 45

Lys Asn Lys Ser Tyr Phe Met Ile Tyr Lys Phe Arg Ser Met Tyr Val
50 55 60

Asp Ala Pro Ser Asp Met Pro Thr His Leu Leu Lys Asp Pro Lys Ala
65 70 75 80

Met Ile Thr Lys Val Gly Ala Phe Leu Arg Lys Thr Ser Leu Asp Glu
85 90 95

Leu Pro Gln Leu Phe Asn Ile Phe Lys Gly Glu Met Ala Ile Val Gly
100 105 110

Pro Arg Pro Ala Leu Trp Asn Gln Tyr Asp Leu Ile Glu Glu Arg Asp
115 120 125
Lys Tyr Gly Ala Asn Asp Ile Arg Pro Gly Leu Thr Gly Trp Ala Gln
130 135 140
Ile Asn Gly Arg Asp Glu Leu Glu Ile Asp Glu Lys Ser Lys Leu Asp
145 150 155 160
Gly Tyr Tyr Val Gln Asn Met Ser Leu Gly Leu Asp Ile Lys Cys Phe
165 170 175
Leu Gly Thr Phe Leu Ser Val Ala Arg Ser Glu Gly Val Val Glu Gly
180 185 190
Gly Thr Gly Gln Lys Gly Lys Gly
195 200

<210> 41

<211> 269

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS2G

<400> 41

Met Lys Phe Ser Val Leu Met Ser Val Tyr Glu Lys Glu Lys Pro Glu
1 5 10 15

Phe Leu Arg Glu Ser Leu Glu Ser Ile Leu Val Asn Gln Thr Met Ile
20 25 30

Pro Thr Glu Val Val Leu Val Glu Asp Gly Pro Leu Asn Gln Ser Leu
35 40 45

Tyr Ser Ile Leu Glu Glu Phe Lys Ser Arg Phe Ser Phe Phe Lys Thr
50 55 60

Ile Ala Leu Glu Lys Asn Ser Gly Leu Gly Ile Ala Leu Asn Glu Gly
65 70 75 80

Leu Lys His Cys Asn Tyr Glu Trp Val Cys Thr Lys Trp Ile Leu Met
85 90 95

Met Leu His Ile His Thr Arg Phe Glu Lys Gln Val Asn Phe Ile Lys
100 105 110

Gln Asn Pro Thr Ile Asp Ile Glu Ile Asp Glu Phe Leu Asn Ser Thr
115 120 125

Ser Glu Ile Val Ser His Lys Asn Val Pro Thr Gln His Asp Glu Ile
130 135 140

Leu Lys Met Ala Arg Arg Glu Lys Ser Met Cys His Met Thr Val Met
145 150 155 160

Phe Lys Lys Lys Ser Val Glu Arg Ala Gly Gly Tyr Gln Thr Leu Pro
165 170 175

Tyr Val Glu Asp Tyr Phe Leu Trp Val Arg Met Ile Ala Ser Gly Ser
180 185 190

Lys Phe Ala Asn Ile Asp Glu Thr Leu Val Leu Ala Arg Val Gly Asn
195 200 205

Gly Met Phe Asn Arg Arg Gly Asn Arg Glu Gln Ile Asn Ser Trp Thr
210 215 220

Leu Leu Ile Glu Phe Met Leu Ala Gln Gly Ile Val Thr Pro Leu Asp
225 230 235 240

Val Phe Ile Asn Gln Ile Tyr Ile Arg Val Phe Val Tyr Met Pro Thr
245 250 255

Trp Ile Lys Lys Leu Ile Tyr Gly Lys Ile Leu Arg Lys
260 265

<210> 42

<211> 143

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS9H

<400> 42

Met Ile Thr Val Leu Met Ala Thr Tyr Asn Gly Ser Pro Phe Ile Ile
1 5 10 15

Lys Gln Leu Asp Ser Ile Arg Asn Gln Ser Val Ser Ala Asp Lys Val
20 25 30

Ile Ile Trp Asp Asp Cys Ser Thr Asp Asp Thr Ile Lys Ile Ile Lys
35 40 45

Asp Tyr Ile Lys Lys Tyr Ser Leu Asp Ser Trp Val Val Ser Gln Asn
50 55 60

Lys Ser Asn Gln Gly His Tyr Gln Thr Phe Ile Asn Leu Thr Lys Leu
65 70 75 80

Val Gln Glu Gly Ile Val Phe Phe Ser Asp Gln Asp Asp Ile Trp Asp
85 90 95

Cys His Lys Ile Glu Thr Met Leu Pro Ile Phe Asp Arg Glu Asn Val
100 105 110

Ser Met Val Phe Cys Lys Ser Arg Leu Ile Asp Glu Asn Gly Asn Ile
115 120 125

Ile Ser Ser Pro Asp Thr Ser Asp Arg Ile Asn Thr Tyr Ser Leu
130 135 140

<210> 43

<211> 3738

<212> DNA

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS7

<400> 43

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ttatggtttc aacagataaa gctgttaatc cgccaaatgt catggagcg actaaacgtg 180

ttgcagaaat gattgtaaca ggtttaaacg agccaggtca gactcaattt gcggcagtcc 240
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gaaaaggtgg acctgttacg gttaccgact ttaggatgac tcgttatttc atgacgattc 360
ctgaggcaag tcgttgggtt atccaagctg gacatttggc aaaaggtgga gaaatcttg 420
tcttggatat gggtgagcca gtacaaatcc tggaaattggc aagaaaaagtt atcttggtaa 480
gcggacatac agaggaagaa atcgggattg tagaatctgg aatcagacca ggcgagaaac 540
tctacgagga attgttatca acagaagaac gtgtcagcga acagattcat gaaaaaatat 600
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tccaaaaaga tagaaatgaa ttaaaagata tggtgattga atttgcaaaa caagaataag 720
aaagtaaaaaa atattttac tttccttagag tttaaacgat gtttaagttc taggaaggaa 780
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tgaatgcaac ctctcgctcg ttactaagca ggagatagta aagttgcttg aaagagagtt 960
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actttggaat tatttctgat tttaaagtaa tgattgatac agtaattaaa gtaataaaat 1860
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gactagaaaa atgatgtaca attacggcta tccagggtgt ttgactttca tgtatgatgc 3600
agacaaaatg ggttaattc agataaaaga tataaagaaa aataacgatt atgogatatt 3660
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<210> 44

<211> 238

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS7E

<400> 44

Ala Ala His Lys His Val Pro Leu Met Glu Tyr Asn Pro His Glu Ala
1 5 10 15

Val Lys Asn Asn Ile Phe Gly Thr Lys Asn Val Ala Glu Ala Ala Lys
20 25 30

Thr Ala Lys Val Ala Lys Phe Val Met Val Ser Thr Asp Lys Ala Val
35 40 45

Asn Pro Pro Asn Val Met Gly Ala Thr Lys Arg Val Ala Glu Met Ile
50 55 60

Val Thr Gly Leu Asn Glu Pro Gly Gln Thr Gln Phe Ala Ala Val Arg
65 70 75 80

Phe Gly Asn Val Leu Gly Ser Arg Gly Ser Val Val Pro Leu Phe Lys
85 90 95

Glu Gln Ile Arg Lys Gly Gly Pro Val Thr Val Thr Asp Phe Arg Met
100 105 110

Thr Arg Tyr Phe Met Thr Ile Pro Glu Ala Ser Arg Leu Val Ile Gln
115 120 125

Ala Gly His Leu Ala Lys Gly Gly Glu Ile Phe Val Leu Asp Met Gly
130 135 140

Glu Pro Val Gln Ile Leu Glu Leu Ala Arg Lys Val Ile Leu Leu Ser
145 150 155 160

Gly His Thr Glu Glu Glu Ile Gly Ile Val Glu Ser Gly Ile Arg Pro
165 170 175

Gly Glu Lys Leu Tyr Glu Glu Leu Leu Ser Thr Glu Glu Arg Val Ser
180 185 190

Glu Gln Ile His Glu Lys Ile Phe Val Gly Arg Val Thr Asn Lys Gln
195 200 205

Ser Asp Ile Val Asn Ser Phe Ile Asn Gly Leu Leu Gln Lys Asp Arg
210 215 220

Asn Glu Leu Lys Asp Met Leu Ile Glu Phe Ala Lys Gln Glu
225 230 235

<210> 45

<211> 232

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS7F

<400> 45

Met Thr Arg Val Glu Leu Ile Thr Arg Glu Phe Phe Lys Lys Asn Glu
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Ala Thr Ser Lys Tyr Phe Gln Lys Ile Glu Ser Arg Arg Gly Glu Leu
20 25 30

Phe Ile Lys Phe Phe Met Asp Lys Leu Leu Ala Leu Ile Leu Leu Leu
35 40 45

Leu Leu Ser Pro Val Ile Ile Ile Leu Ala Ile Trp Ile Lys Leu Asp
50 55 60

Ser Lys Gly Pro Ile Phe Tyr Arg Gln Glu Arg Val Thr Arg Tyr Gly
65 70 75 80

Arg Ile Phe Arg Ile Phe Lys Phe Arg Thr Met Ile Ser Asp Ala Asp
85 90 95

Lys Val Gly Ser Leu Val Thr Val Gly Gln Asp Asn Arg Ile Thr Lys
100 105 110

Val Gly His Ile Ile Arg Lys Tyr Arg Leu Asp Glu Val Pro Gln Leu
115 120 125

Phe Asn Val Leu Met Gly Asp Met Ser Phe Val Gly Val Arg Pro Glu
130 135 140

Val Gln Lys Tyr Val Asn Gln Tyr Thr Asp Glu Met Phe Ala Thr Leu
145 150 155 160

Leu Leu Pro Ala Gly Ile Thr Ser Pro Ala Ser Ile Ala Tyr Lys Asp
165 170 175

Glu Asp Ile Val Leu Glu Glu Tyr Cys Ser Gln Gly Tyr Ser Pro Asp
180 185 190

Glu Ala Tyr Val Gln Lys Val Leu Pro Glu Lys Met Lys Tyr Asn Leu
195 200 205

Glu Tyr Ile Arg Asn Phe Gly Ile Ile Ser Asp Phe Lys Val Met Ile
210 215 220

Asp Thr Val Ile Lys Val Ile Lys
225 230

<210> 46

<211> 404

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> CPS7G

<400> 46

Met Thr Lys Arg Gln Asn Ile Pro Phe Ser Pro Pro Asp Ile Thr Gln
1 5 10 15

Ala Glu Ile Asp Glu Val Ile Asp Thr Leu Lys Ser Gly Trp Ile Thr
20 25 30

Thr Gly Pro Lys Thr Lys Glu Leu Glu Arg Arg Leu Ser Val Phe Thr
35 40 45

Gly Thr Asn Lys Thr Val Cys Leu Asn Ser Ala Thr Ala Gly Leu Glu
50 55 60

Leu Val Leu Arg Ile Leu Gly Val Gly Pro Gly Asp Glu Val Ile Val
65 70 75 80

Pro Ala Met Thr Tyr Thr Ala Ser Cys Ser Val Ile Thr His Val Gly
85 90 95

Ala Thr Pro Val Met Val Asp Ile Gln Lys Asn Ser Phe Glu Met Glu
100 105 110

Tyr Asp Ala Leu Glu Lys Ala Ile Thr Pro Lys Thr Lys Val Ile Ile
115 120 125

Pro Val Asp Leu Ala Gly Ile Pro Cys Asp Tyr Asp Lys Ile Tyr Thr
130 135 140

Ile Val Glu Asn Lys Arg Ser Leu Tyr Val Ala Ser Asp Asn Lys Trp
145 150 155 160

Gln Lys Leu Phe Gly Arg Val Ile Ile Leu Ser Asp Ser Ala His Ser
165 170 175

Leu Gly Ala Ser Tyr Lys Gly Lys Pro Ala Gly Ser Leu Ala Asp Phe
180 185 190

Thr Ser Phe Ser Phe His Ala Val Lys Asn Phe Thr Thr Ala Glu Gly
195 200 205

Gly Ser Val Thr Trp Arg Ser His Pro Asp Leu Asp Asp Glu Glu Met
210 215 220

Tyr Lys Glu Phe Gln Ile Tyr Ser Leu His Gly Gln Thr Lys Asp Ala
225 230 235 240

Leu Ala Lys Thr Gln Leu Gly Ser Trp Glu Tyr Asp Ile Val Ile Pro
245 250 255

Gly Tyr Lys Cys Asn Met Thr Asp Ile Met Ala Gly Ile Gly Leu Val
260 265 270

Gln Leu Glu Arg Tyr Pro Ser Leu Leu Asn Arg Arg Arg Glu Ile Ile
 275 280 285
 Glu Lys Tyr Asn Ala Gly Phe Glu Gly Thr Ser Ile Lys Pro Leu Val
 290 295 300
 His Leu Thr Glu Asp Lys Gln Ser Ser Met His Leu Tyr Ile Thr His
 305 310 315 320
 Leu Gln Gly Tyr Thr Leu Glu Gln Arg Asn Glu Val Ile Gln Lys Met
 325 330 335
 Ala Glu Ala Gly Ile Ala Cys Asn Val His Tyr Lys Pro Leu Pro Leu
 340 345 350
 Leu Thr Ala Tyr Lys Asn Leu Gly Phe Glu Met Lys Asp Phe Pro Asn
 355 360 365
 Ala Tyr Gln Tyr Phe Glu Asn Glu Val Thr Leu Pro Leu His Thr Asn
 370 375 380
 Leu Ser Asp Glu Asp Val Glu Tyr Val Ile Glu Met Phe Leu Lys Ile
 385 390 395 400
 Val Ser Arg Asp

<210> 47

<211> 210

<212> PRT

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> CPS7H

<400> 47

Met Val Glu Arg Asp Met Val Glu Arg Asp Thr Leu Val Ser Ile Ile
 1 5 10 15

Met Pro Ser Trp Asn Thr Ala Lys Tyr Ile Ser Glu Ser Ile Gln Ser
 20 25 30

Val Leu Asp Gln Thr His Gln Asn Trp Glu Leu Ile Ile Val Asp Asp
 35 40 45

Cys	Ser	Asn	Asp	Glu	Thr	Glu	Lys	Val	Val	Ser	His	Phe	Lys	Asp	Ser
50						55					60				
Arg	Ile	Lys	Phe	Phe	Lys	Asn	Ser	Asn	Asn	Leu	Gly	Ala	Ala	Leu	Thr
65					70					75				80	
Arg	Asn	Lys	Ala	Leu	Arg	Lys	Ala	Arg	Gly	Arg	Trp	Ile	Ala	Phe	Leu
					85				90				95		
Asp	Ser	Asp	Asp	Leu	Trp	His	Pro	Ser	Lys	Leu	Glu	Lys	Gln	Leu	Glu
					100			105				110			
Phe	Met	Lys	Asn	Asn	Gly	Tyr	Ser	Phe	Thr	Tyr	His	Asn	Phe	Glu	Lys
					115			120			125				
Ile	Asp	Glu	Ser	Ser	Gln	Ser	Leu	Arg	Val	Leu	Val	Ser	Gly	Pro	Ala
					130			135			140				
Ile	Val	Thr	Arg	Lys	Met	Met	Tyr	Asn	Tyr	Gly	Tyr	Pro	Gly	Cys	Leu
					145			150			155			160	
Thr	Phe	Met	Tyr	Asp	Ala	Asp	Lys	Met	Gly	Leu	Ile	Gln	Ile	Lys	Asp
					165			170				175			
Ile	Lys	Lys	Asn	Asn	Asp	Tyr	Ala	Ile	Leu	Leu	Gln	Leu	Cys	Lys	Lys
					180				185			190			
Tyr	Asp	Cys	Tyr	Leu	Leu	Asn	Glu	Ser	Leu	Ala	Ser	Tyr	Arg	Ile	Arg
					195			200			205				
Lys	Lys														
	210														

<210> 48
<211> 101
<212> DNA
<213> *Streptococcus suis*
<220>
<221> misc_feature
<222> (1)..(101)
<223> N may be any nucleotide
<220>

<221> misc_feature

<223> 100 base pair repeat between CPS2G and CPS2H

<400> 48

aagggcacct ctataaactc ccaaaaattgc gaatttggag ttacgaaagc cttgttaaat 60
caancatttt aaattttaga aaatttagttt ttagagctcc c 101

<210> 49

<211> 101

<212> DNA

<213> *Streptococcus suis*

<220>

<221> misc_feature

<222> (1)..(101)

<223> N may be any nucleotide

<220>

<221> misc_feature

<223> 100 base pair repeat within CPS2M

<400> 49

ggcgccaccc ctataaattc ccaaaaattgc gaatttcgag ttacgaaagc cttgttaaat 60
caancatctt aaattttaga aaatttagttt ttagaggtcc c 101

<210> 50

<211> 101

<212> DNA

<213> *Streptococcus suis*

<220>

<221> misc_feature

<223> 100 base pair repeat between CPS2O and CPS2P
 <400> 50
 aagggcacct ctataaactc ccaaaattgc gaatttcgag ttacgaaagc cttgttaat 60
 caaacatttt aaattttaga aaatttagttt ttagaggtcc c 101
 <210> 51
 <211> 120
 <212> PRT
 <213> *Streptococcus suis*
 <220>
 <221> misc_feature
 <223> N-terminal part of CPS2J
 <400> 51
 Met Ala Lys Val Ser Ile Ile Val Pro Ile Phe Asn Thr Glu Lys Tyr
 1 5 10 15
 Leu Arg Glu Cys Leu Asp Ser Ile Ile Ser Gln Ser Tyr Thr Asn Leu
 20 25 30
 Glu Ile Leu Leu Ile Asp Asp Gly Ser Ser Asp Ser Ser Thr Asp Ile
 35 40 45
 Cys Leu Glu Tyr Ala Glu Gln Asp Gly Arg Ile Lys Leu Phe Arg Leu
 50 55 60
 Pro Asn Gly Gly Val Ser Asn Ala Arg Asn Tyr Gly Ile Lys Asn Ser
 65 70 75 80
 Thr Ala Asn Tyr Ile Met Phe Val Asp Ser Asp Asp Ile Val Asp Gly
 85 90 95
 Asn Ile Val Glu Ser Leu Tyr Thr Cys Leu Lys Glu Asn Asp Ser Asp
 100 105 110
 Leu Ser Gly Gly Leu Leu Ala Thr
 115 120

<210> 52
 <211> 120
 <212> PRT
 <213> *Streptococcus suis*
 <220>
 <221> misc_feature
 <223> N-terminal part of CPS2K
 <220>
 <221> misc_feature
 <222> (1)..(120)
 <223> Xaa may be any amino acid
 <400> 52

Met	Ile	Asn	Ile	Ser	Ile	Ile	Val	Pro	Ile	Tyr	Asn	Val	Glu	Gln	Tyr
1				5					10				15		

Leu	Ser	Lys	Cys	Ile	Asn	Ser	Ile	Val	Asn	Gln	Thr	Tyr	Lys	His	Ile
			20					25					30		

Glu	Leu	Leu	Val	Asn	Asp	Gly	Ser	Ser	Thr	Asp	Asn	Ser	Glu	Glu	Ile
				35			40				45				

Cys	Leu	Ala	Tyr	Ala	Lys	Lys	Asp	Ser	Arg	Ile	Arg	Tyr	Phe	Lys	Lys
					50			55			60				

Glu	Asn	Gly	Gly	Leu	Ser	Asp	Ala	Arg	Asn	Tyr	Gly	Ile	Ser	Arg	Ala
				65			70		75				80		

Lys	Gly	Asp	Tyr	Leu	Ala	Phe	Ile	Asp	Ser	Asp	Asp	Phe	Ile	His	Ser
					85			90					95		

Glu	Phe	Ile	Gln	Arg	Leu	Xaa	His	Glu	Ala	Ile	Glu	Arg	Glu	Asn	Ala
					100			105				110			

Leu	Xaa	Xaa	Val	Ala	Val	Ala	Gly								
				115			120								

<210> 53
 <211> 419

<212> PRT

<213> Streptococcus suis

<220>

<221> misc_feature

<223> ORF2Y

<400> 53

Met Lys Lys Tyr Gln Val Ile Ile Gln Asp Ile Leu Thr Gly Ile Glu
1 5 10 15

Glu His Arg Phe Lys Arg Gly Glu Lys Leu Pro Ser Ile Arg Gln Leu
20 25 30

Arg Glu Gln Tyr His Cys Ser Lys Asp Thr Val Gln Lys Ala Met Leu
35 40 45

Glu Leu Lys Tyr Gln Asn Lys Ile Tyr Ala Val Glu Lys Ser Gly Tyr
50 55 60

Tyr Ile Leu Glu Asp Arg Asp Phe Gln Asp His Thr Cys Arg Ala Gln
65 70 75 80

Ser Tyr Arg Leu Ser Arg Ile Thr Tyr Glu Asp Phe Arg Ile Cys Leu
85 90 95

Lys Glu Ser Leu Ile Gly Arg Glu Asn Tyr Leu Phe Asn Tyr Tyr His
100 105 110

Gln Gln Glu Gly Leu Ala Glu Leu Ile Ser Ser Val Gln Ser Leu Leu
115 120 125

Met Asp Tyr His Val Tyr Thr Lys Lys Asp Gln Leu Val Ile Thr Ala
130 135 140

Gly Ser Gln Gln Ala Leu Tyr Ile Leu Thr Gln Met Glu Thr Leu Ala
145 150 155 160

Gly Lys Thr Glu Ile Leu Ile Glu Asn Pro Thr Tyr Ser Arg Met Ile
165 170 175

Glu Leu Ile Arg His Gln Gly Ile Pro Tyr Gln Thr Ile Glu Arg Asn
180 185 190

Leu Asp Gly Ile Asp Leu Glu Glu Leu Glu Ser Ile Phe Gln Thr Gly
195 200 205

Lys Ile Lys Phe Phe Tyr Thr Ile Pro Arg Leu His Asn Pro Leu Gly
210 215 220

Ser Thr Tyr Asp Ile Ala Thr Lys Thr Ala Ile Val Lys Leu Ala Lys
225 230 235 240

Gln Tyr Asp Val Tyr Ile Ile Glu Asp Asp Tyr Leu Ala Asp Phe Asp
245 250 255

Ser Ser His Ser Leu Pro Leu His Tyr Leu Asp Thr Asp Asn Arg Val
260 265 270

Ile Tyr Ile Lys Ser Phe Thr Pro Thr Leu Phe Pro Ala Leu Arg Ile
275 280 285

Gly Ala Ile Ser Leu Pro Asn Gln Leu Arg Asp Ile Phe Ile Lys His
290 295 300

Lys Ser Leu Ile Asp Tyr Asp Thr Asn Leu Ile Met Gln Lys Ala Leu
305 310 315 320

Ser Leu Tyr Ile Asp Asn Gly Met Phe Ala Arg Asn Thr Gln His Leu
325 330 335

His His Ile Tyr His Ala Gln Trp Asn Lys Ile Lys Asp Cys Leu Glu
340 345 350

Lys Tyr Ala Leu Asn Ile Pro Tyr Arg Ile Pro Lys Gly Ser Val Thr
355 360 365

Phe Gln Leu Ser Lys Gly Ile Leu Ser Pro Ser Ile Gln His Met Phe
370 375 380

Gly Lys Cys Tyr Tyr Phe Ser Gly Gln Lys Ala Asp Phe Leu Gln Ile
385 390 395 400

Phe Phe Glu Gln Asp Phe Ala Asp Lys Leu Glu Gln Phe Val Arg Tyr
405 410 415

Leu Asn Glu

TABLE 1. Bacterial strains and plasmids

strain/plasmid	relevant characteristics	source/reference
Strain		
<i>E. coli</i>		
CC118	PhoA ⁻	(28)
XL2 blue	Stratagene	
<i>E. coli</i>		
XL2 blue	Stratagene	
<i>S. suis</i>		
10	virulent serotype 2 strain	(49)
3	serotype 2	(63)
17	serotype 2	(63)
735	reference strain serotype 2	(63)
T15	serotype 2	(63)
6555	reference strain serotype 1	(63)
6388	serotype 1	(63)
6290	serotype 1	(63)
5637	serotype 1	(63)
5673	serotype 1/2	(63)
5679	serotype 1/2	(63)
5928	serotype 1/2	(63)
5934	serotype 1/2	(63)
5209	reference strains serotype 1/2	(63)
5218	reference strain serotype 9	(63)

strain/plasmid	relevant characteristics	source/reference
5973	serotype 9	(63)
6437	serotype 9	(63)
6207	serotype 9	(63)
reference strains	serotypes 1-34	(9, 56, 14)
<i>S. suis</i>		
10	virulent serotype 2 strain	(51)
10cpsB	isogenic cpsB mutant of strain 10	this work
10cpsEF	isogenic cpsEF mutant of strain 10	this work
Plasmid		
pKUN19	replication functions pUC, Amp ^R	(23)
pGEM7zf (+)	replication functions pUC, Amp ^R	Promega Corp.
pIC19R	replication functions pUC, Amp ^R	(29)
pIC20R	replication functions pUC, Amp ^R	(29)
pIC-spc	pIC19R containing spc ^R gene of pDL282	labcollection
pDL282	replication functions of pBR 322 and pVT736-1, Amp ^R , Spc ^R	(43)
pPHOS2	pIC-spc containing the truncated <i>phoA</i> gene of pPHO7 as a <i>PstI-BamHI</i> fragment	this work
pPHO7	contains truncated <i>phoA</i> gene	(15)
pPHOS7	pPHOS2 containing chromosomal <i>S. suis</i> DNA	this work
pCPS6	pKUN19 containing 6 kb <i>HindIII</i> fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS7	pKUN19 containing 3,5 kb <i>EcoRI-HindIII</i> fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS11	pCPS7 in which 0.4 kb <i>PstI-BamHI</i> fragment of <i>cpsB</i> gene is replaced by Spc ^R gene of pIC-spc	this work (Fig. 1)

strain/plasmid	relevant characteristics	source/reference
pCPS17	pKUN19 containing 3.1 kb <i>Kpn</i> I fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS18	pKUN19 containing 1.8 kb <i>Sna</i> BI fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS20	pKUN19 containing 3.3 kb <i>Xba</i> I- <i>Hind</i> III fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS23	pGEM7Zf (+) containing 1.5 kb <i>Mlu</i> I fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS25	pIC20R containing 2.5 kb <i>Kpn</i> I- <i>Sal</i> I fragment of pCPS17	this work (Fig. 1)
pCPS26	pKUN19 containing 3.0 kb <i>Hind</i> III fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS27	pCPS25 containing 2.3 kb <i>Xba</i> I (blunt) - <i>Cl</i> I fragment of pCPS20	this work (Fig. 1)
pCPS28	pCPS27 containing the 1.2 kb <i>Pst</i> I- <i>Xho</i> I <i>Spc</i> ^R gene of pIC-spc	this work (Fig. 1)
pCPS29	pKUN19 containing 2.2 kb <i>Sac</i> I- <i>Pst</i> I fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS1-1	pKUN19 containing 5 kb <i>Eco</i> RV fragment of <i>cps</i> operon of type 1	this work (Fig. 1)
pCPS1-2	pKUN19 containing 2.2 kb <i>Hind</i> III fragment of <i>cps</i> operon of type 1	this work (Fig. 1)
pCPS9-1	pKUN19 containing 1 kb <i>Hind</i> III- <i>Xba</i> I fragment of <i>cps</i> operon of serotype 9	this work (Fig. 1)
pCPS9-2	pKUN19 containing 4.0 kb <i>Xba</i> I- <i>Xba</i> I fragment of <i>cps</i> operon of serotype 9	this work (Fig. 1)

Amp^R: ampicillin resistant

Spc^R: spectinomycin resistant

cps: capsular polysaccharide

Table 2. Properties of Orfs in the cps locus of *S. suis* serotype 2 and similarities to gene products of other bacteria

ORF	nucleotide position in sequence	number of amino acids	GC%	proposed function of gene product ¹	similar gene product (% identity)
Orf2Z	1- 719	240	44	Unknown	<i>B. subtilis</i> YitS (26%)
Orf2Y	2079- 822	419	38	Transcription regulation	<i>B. subtilis</i> YcxD (39%)
Orf2X	2202- 2934	244	39	Unknown	<i>H. influenzae</i> YAAA (24%)
Cps2A	3041- 4484	481	39	Regulation	<i>S. pneumoniae</i> Cps19fA (58%)
Cps2B	4504- 5191	229	40	Chain length determination	<i>S. pneumoniae</i> type 3 Orf1 (58%)
Cps2C	5203- 5878	225	40	Chain length determination/ Export	<i>S. pneumoniae</i> Cps23fD (63%)
Cps2D	5919- 6648	243	38	Unknown	<i>S. pneumoniae</i> CpsB (62%)
Cps2E	6675- 8052	459	33	Glycosyltransferase	<i>S. pneumoniae</i> Cps14E (56%)
Cps2F	8089- 9256	389	32	Glycosyltransferase	<i>S. pneumoniae</i> Cps23fT
Cps2G	9262-10417	385	36	Glycosyltransferase	<i>S. thermophilus</i> EpsF (25%)
Cps2H	10808-12176	457	31	Glycosyltransferase	<i>S. mutans</i> RGPEC _N (29%)
Cps2I	12213-13443	410	29	CP polymerase	<i>S. pneumoniae</i> Cps23fI (48%)
Cps2J	13583-14579	332	29	Glycosyltransferase	<i>S. pneumoniae</i> Cps14J (31%)
Cps2K	14574-15576	334	37	Glycosyltransferase	<i>S. pneumoniae</i> Cps14J (40%)
“Cps2L”	15618-16635	103	37	Unknown	-
“Cps2M”	16811-17322	-	38	-	<i>S. agalactiae</i> CpsF _N (77%)
					<i>E. coli</i> NeuA _N (47%)

ORF	nucleotide position in sequence	number of amino acids	GC%	proposed function of gene product ¹	similar gene product (% identity)
“Cps2N”	17559-18342	-	39	-	<i>S. agalactiae</i> CpsJ (43%)
Cps2O	18401-19802	476	40	Repeat unit transporter	<i>S. agalactiae</i> CpsK (41%)
Cps2P	20327-21341	338	39	Sialic acid synthesis	<i>S. agalactiae</i> NeuB (80%)
					<i>E. coli</i> NeuB (59%)
Cps2Q	21355-21865	170	42	Sialic acid synthesis	<i>S. agalactiae</i> NeuC ^N (61%)
					<i>E. coli</i> NeuC ^N (54%)
Cps2R	21933-22483	184	40	Sialic acid synthesis	<i>S. agalactiae</i> NeuC ^C (55%)
					<i>E. coli</i> NeuC ^C (40%)
Cps2S	22501-23125	208	42	Sialic acid synthesis	<i>E. coli</i> NeuD (32%)
Cps2T	23136-24366	395	40	CMP-NeuNAc synthetase	<i>S. agalactiae</i> CpsF (49%)
					<i>E. coli</i> NeuA (34%)
“Orf2U”	24566-25488	168	42	Transposase	<i>S. thermophilus</i> IS1194 (51%)
“Orf2V”	25691-26281	116	37	Transposase	<i>S. pneumoniae</i> orf1 (85%)

¹ Predicted by sequence similarity

N Similarity refers to the amino-terminal part of the gene product

C Similarity refers to the carboxy-terminal part of the gene product

ORFs between “ ” are truncated or non-functional as the result of frame-shift or point mutations

TABLE 3. Properties of ORFs in the cps gene of *S. suis* serotypes 1 and 9 and similarities to gene products of other bacteria

ORF	nucleotide position in sequence	G + C%	number of amino acids	predicted mol. mass	predicted pI	proposed function of gene product ¹	similar gene product (% identity)	reference/ accession nr.
Cps1E ²	1-1363	34%	454	52.2	8.0	Glucosyltransferase	<i>Streptococcus suis</i> Cps2E (86%)	(26)
Cps1F	1374-1821	33%	149	17.3	8.2	Unknown	<i>Streptococcus pneumoniae</i> Cps14E (48%)	(12)
Cps1G	1823-2315	25%	164	19.5	7.5	Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14F (83%)	(14)
Cps1H	3035-4202	24%	389	45.5	8.4	CP polymerase	<i>Streptococcus pneumoniae</i> Cps14G (50%)	(14)
Cps1I	4917-					Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14H (30%)	(14)
							<i>Lactococcus lactis</i> EpsG (31%)	(29)
Cps1J							<i>Streptococcus thermophilus</i> EpsI (33%)	(28)
							<i>Streptococcus pneumoniae</i> Cps14J (%)	(13)

ORF	nucleotide position in sequence	G + C%	number of amino acids	predicted mol. mass (kDa)	predicted pI	proposed function ¹ of gene product ¹	similar gene product (% identity)	reference/ accession nr.
Cps1K ³		37%	278	32.5	7.8	Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14J (44%)	(13)
Cps9D ²	1-646	37%	215	24.9	8.1	Unknown	<i>Streptococcus suis</i> Cps2D (89%)	(26)
Cps9E	680-					Glycosyltransferase	<i>Staphylococcus aureus</i> Cap1D (27%)	(18)
Cps9F		36%	200	22.3	8.2	Glycosyltransferase	<i>Staphylococcus aureus</i> Cap5M (52%)	(17)
Cps9G		35%	269	31.5	8.0	Unknown	<i>Actinobacillus actinomycetemcomitans</i> (43%)	(AB002668_4)
							<i>Haemophilus influenzae</i> Lsg (43%)	(O05081)
Cps9H ³		30%	143	16.5	7.2	Unknown	<i>Yersinia enterolitica</i> RfbB (28%)	(33)

¹ Predicted by sequence similarity

² N-terminal part of protein is lacking

³ C-terminal part of protein is lacking

Table 4. Hybridization of serotype 2 *cps* genes and neighboring sequences with chromosomal DNA of serotypes

serotypes	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	1/2
DNA probes																																			
<i>orf2Z</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>orf2Y</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>orf2X</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2A</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2B</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2C</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2D</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2E</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2F</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2G</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2H</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2I</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2J</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2K</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
“ <i>cps2L</i> ”	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
“ <i>cps2M</i> ”	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2N</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2O</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2P</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2Q</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2R</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				
<i>cps2S</i>	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				

serotypes	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	1/2
<i>cps2T</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-		
“ <i>orf2U</i> ”	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-		
“ <i>orf2V</i> ”	+	+	±	±	-	±	-	-	-	-	-	-	-	-	+	+	±	-	±	+	-	-	-	-	-	-	-	-	-	-	-	-	-		
<i>100-bp repeat</i>	+	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-		
<i>16SrRNA</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+		

Table 5. Hybridization of serotypes 1 and 9 *cps* genes with chromosomal DNA of other *S. suis* serotypes

Serotype	DNA probes							<i>cps9H</i>	<i>cps9G</i>	<i>cps9F</i>	<i>cps9E</i>	<i>cps9H</i>	<i>cps9I</i>	<i>cps9G</i>	<i>cps9F</i>	<i>cps9E</i>	-
	<i>cps1E</i>	<i>cps1F</i>	<i>cps1G</i>	<i>cps1H</i>	<i>cps1I</i>	<i>cps1H</i>	<i>cps1G</i>										
1	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-
2	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
8	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
11	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
12	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
15	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Serotype	<i>cpsIE</i>	<i>cpsIF</i>	<i>cpsIG</i>	<i>cpsIH</i>	<i>cpsII</i>	<i>cps9E</i>	<i>cps9F</i>	<i>cps9G</i>	<i>cps9H</i>	<i>16rRNA</i>
16	-	-	-	-	-	-	-	-	-	+
17	-	-	-	-	-	-	-	-	-	+
18	-	-	-	-	-	-	-	-	-	+
19	-	-	-	-	-	-	-	-	-	-
20	-	-	-	-	-	-	-	-	-	-
21	-	-	-	-	-	±	-	-	-	-
22	-	-	-	-	-	+	-	-	-	-
23	-	-	-	-	-	+	-	-	-	-
24	-	-	-	-	-	+	-	-	-	-
25	-	-	-	-	-	-	-	-	-	-
26	-	-	-	-	-	-	-	-	-	-
27	-	-	-	-	-	-	-	-	-	-
28	-	-	-	-	-	-	-	-	-	-
29	-	-	-	-	-	-	-	-	-	-
30	-	-	-	-	-	-	-	-	-	-
31	-	-	-	-	-	-	-	-	-	-
32	-	-	-	-	-	-	-	-	-	-
33	-	-	-	-	-	-	-	-	-	-
34	-	-	-	-	-	-	-	-	-	-
½	-	-	-	-	-	-	-	-	-	-

TABLE 6. Virulence of wild-type and capsular mutant *S. suis* strains in germfree pigs

<i>S. suis</i> strains ¹	pigs/ group (n)	mortality ² (%)	morbidity ³ (%)	clinical index of the group		fever index ⁷	leucocyte index ⁸	isolation of <i>S. suis</i> in pigs (n) per group in
				spec symptoms ⁵	non-spec. symptoms ⁶			
10	4	100	100	11	88	43	44	2
10cpsB	4	0	0	0	10	1	3	3
10cpsEF	4	0	0	0	0	1	0	2

¹ strain 10 in the wild-type strain, strains 10cpsB and 10cpsEF are isogenic capsular mutant strains

² piglets which died spontaneously or had to be killed for animal welfare reasons

³ only considering pigs with specific symptoms

⁴ clinical index: % of observations which matched the described criteria

⁵ specific symptoms: ataxia, lameness on at least one joint, stiffness

⁶ non-specific symptoms: inappetance, depression

⁷ % of observations in the experimental group with a body temperature > 40°C

⁸ % of blood samples in the group in which number of granulocytes > 10¹⁰/1

Table 7. Bacterial strains and plasmids

strain/plasmid	relevant characteristics
Strain	
<i>E. coli</i>	
XL2 blue	
<i>S. suis</i>	serotypes 1-34
reference strains	
5667	serotype 7, tonsil (1993)
7037	serotype 7, organs (1994)
7044	serotype 7, brains (1994)
7068	serotype 7 (1994)
7646	serotype 7 (1994)
7744	serotype 7, lungs (1996)
7759	serotype 7, joints (1996)
8169	serotype 7 (1997)
15913	serotype 7, meninges (1998)
Plasmid	
pKUN19	replication functions pUC, Amp ^R
pGEM7zf (+)	replication functions pUC, Amp ^R
pCPS9-1	pKUN19 containing 1 kb <i>Hind</i> III- <i>Xba</i> I fragment of <i>cps</i> operon of serotype 9
pCPS9-2	pKUN19 containing 4.09 kb <i>Xba</i> I- <i>Xba</i> I fragment of <i>cps</i> operon of serotype 9
pCPS7-1	pKUN19 containing 1.6-kb <i>Pst</i> I fragment of <i>cps</i> operon of type 7
pCPS7-2	pGEM7 containing 2.7-kb <i>Scal</i> - <i>Clai</i> fragment of <i>cps</i> operon of type 7

Amp^R: ampicillin resistant
cps: capsular polysaccharide

Table 8. Properties of Orfs in the *cps* genes of *S. suis* serotype 7 and similarities to gene products of other bacteria

Orf	nucleotide position in sequence	proposed function of gene product	similar gene product (% identity)
Cps7E	1-719	Glycosyltransferase	<i>Streptococcus suis</i> Cps9E (99%)
Cps7F	1164-1863	Glycosyltransferase	<i>Bordetella pertussis</i> BplG ¹ (43%)
			<i>Streptococcus suis</i> Cps2E ¹ (33%)
Cps7G	1872-3086	Biosynthesis amino sugar	<i>Bordetella pertussis</i> BplF (48%)
Cps7H	3104-3737	Glycosyltransferase	<i>Escherichia coli</i> WbdN (35%)
			<i>Streptococcus suis</i> Cps2K ² (31%)

¹ similarity refers to the C-terminal part of the gene product

² similarity refers to the N-terminal part of the gene product

Table 9. Hybridization of serotype 7 *cps* probes with chromosomal DNA of *S. suis* serotypes

serotypes	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	1/2
DNA probes																																			
<i>cps7E</i>	-	+	+	+	-	+	-	+	+	+	+	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
<i>cps7F</i>	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
<i>cps7G</i>	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
<i>cps7H</i>	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
<i>16SrRNA</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+				

REFERENCES

1. Arends, J. P., and H. C. Zanen. 1988. Meningitis caused by *Streptococcus suis* in humans. *Rev. Infect. Dis.* 10:131-137.
2. Arrecubieta, C., E. Garcia, and R. Lopez. 1995. Sequence and transcriptional analysis of a DNA region involved in the production of capsular polysaccharide in *Streptococcus pneumoniae* type 3. *Gene* 167:1-7
3. Arrecubieta, C., R. Lopez, and E. Garcia. 1994. Molecular characterization of *cap3A*, a gene from the operon required for the synthesis of the capsule of *Streptococcus pneumoniae* type 3: sequencing of mutations responsible for the unencapsulated phenotype and localization of the capsular cluster on the pneumococcal chromosome. *J. Bacteriol.* 176:6375-6383.
4. Clifton-Hadley, F.A. 1983. *Streptococcus suis* type 2 infections. *Br. Vet. J.* 139:1-5.
5. Charland, N., J. Harel, M. Kobisch, S. Lacasse, and M. Gottschalk. 1998. *Streptococcus suis* serotype 2 mutants deficient in capsular expression. *Microbiol.* 144:325-332.
6. Cross, A. S. 1990. The biological significance of bacterial encapsulation. *Curr. Top. Microbiol. Immunol.* 150:87-95.
7. Elliott, S. D. and J. Y. Tai. 1978. The type-specific polysaccharide of *Streptococcus suis*. *J. Exp. Med.* 148:1699-1704.
8. Feder, I., M. M. Chengappa, B. Fenwick, M. Rider and J. Staats. 1994. Partial characterization of *Streptococcus suis* type 2 hemolysin. *J. Clin. Microbiol.* 32:1256-1260.
9. Gottschalk, M., R. Higgins, M. Jacques, M. Beaudoin, and J. Henrichsen. 1991. Characterization of six new capsular types (23 through 28) of *Streptococcus suis*. *J. Clin. Microbiol.* 29:2590-2594.
10. Gottschalk, M., S. Lacouture, and J. D. Dubreuil. 1995. Characterization of *Streptococcus suis* type 2 haemolysin. *Microbiology* 141:189-195.
11. Gottschalk, M., A. Lebrun, M. Jacques, and R. Higgins. 1990. Haemagglutination properties of *Streptococcus suis*. *J. Clin. Microbiol.* 28:2156-2158.
12. Guidolin, A., J. M. Morona, R. Morona, D. Hansman, and J. C. Paton. 1994. Nucleotide sequence analysis of genes essential for capsular polysaccharide biosynthesis in *Streptococcus pneumoniae* type 19F. 1994. *Infect. Immun.* 62:5384-5396.
13. Guitierrez, C., and J. C. Devedjian. 1989. Plasmid facilitating *in vitro* construction of PhoA fusions in *Escherichia coli*. *Nucl. Acid. Res.* 17:3999.
14. Higgins, R., M. Gottschalk, M. Boudreau, A. Lebrun, and J. Henrichsen. 1995. Description of six new capsular types (28 through 34) of *Streptococcus suis*. *J. Vet. Diagn. Invest.* 7:405-406

15. Jacobs, A. A., P. L. W. Loeffen, A. J. G. van den Berg, and P. K. Storm. 1994. Identification, purification and characterization of a thiol-activated hemolysin (suilysin) of *Streptococcus suis*. *Infect. Immun.* 62:1742-1748.
16. Jacques, M., M. Gottschalk, B. Foiry B. and R. Higgins. 1990. Ultrastructural study of surface components of *Streptococcus suis*. *J. Bacteriol.* 172:2833-2838.
17. Klein P., M. Kanehisa and C. DeLisi. 1985. The detection and classification of membrane spanning proteins. *Biochim. Biophys. Acta.* 851:468-476.
18. Kolkman, M. A. B., D. A. Morrison, B. A. M. van der Zeijst, and P. J. M. Nuijten. 1996. The capsule polysaccharide synthesis locus of *Streptococcus pneumoniae* serotype 14: identification of the glycosyl transferase gene *cps14E*. *J. Bacteriol.* 178: 3736-3541.
19. Kolkman, M. A. B., W. Wakarchuk, P. J. M. Nuijten, and B. A. M. van der Zeijst. 1997. Capsular polysaccharide synthesis in *Streptococcus pneumoniae* serotype 14: molecular analysis of the complete *cps* locus and identification of genes encoding glycosyltransferases required for the biosynthesis of the tetrasaccharide subunit. *Mol. Microbiol.* 26:197-208.
20. Kolkman, M. A. B., B. A. M. van der Zeijst and P. J. M. Nuijten. 1997. Functional analysis of glycosyltransferases encoded by the capsular polysaccharide biosynthesis locus of *Streptococcus pneumoniae* serotype 14. *J. Biol. Chem.* 272:19502-19508.
21. Konings, R. N. H., E. J. M. Verhoeven, and B. P H. Peeters. 1987. pKUN vectors for the separate production of both DNA strands of recombinant plasmids. *Methods Enzymol.* 153:12-34.
22. Korolik, V., B. N. Fry, M. R. Alderton, B. A. M. van der Zeijst, and P. J. Coloe. 1997. Expression of *Campylobacter hyoilei* lipo-oligosaccharide (LOS) antigens in *Escherichia coli*. *Microbiol.* 143:3481-3489.
23. Leij, P. C. J., R. van Furth, and T. L. van Zwet. 1986. In vitro determination of phagocytosis and intracellular killing of polymorphonuclear and mononuclear phagocytes. In *Handbook of Experimental Immunology*, vol. 2. *Cellular Immunology*, pp. 46.1-46.21. Edited by D. M. Weir, L. A. Herzenberg, C. Blackwell and L. A. Herzenberg. Blackwell Scientific Publications, Oxford.
24. Lin, W. S., T. Cunneen, and C. Y. Lee. 1994. Sequence analysis and molecular characterization of genes required for the biosynthesis of type 1 capsular polysaccharide in *Staphylococcus aureus*. *J. Bacteriol.* 176:7005-7016.

25. Liu, D., A. M. Haase, L. Lindqvist, A. A. Lindberg, and P. R. Reeves. 1993. Glycosyl transferases of O-antigen biosynthesis in *Salmonella enteritica*: Identification and characterization of transferase genes of group B, C2, and E1. *J. Bacteriol.* 175:3408-3413.
26. Manoil, C., and J. Beckwith. 1985. A transposon probe for protein export signals. *Proc. Natl. Acad. Sci. USA* 82:8129-8133.
27. Marsh, J. L., N. Erfle, and E. J. Wykes. 1984. The pIC plasmid and phage vectors with versatile cloning sites for recombinant selection by insertional inactivation. *Gene* 32:481-485.
28. Miller, J. 1972. Experiments in Molecular Genetics. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory.
29. Morona, J. K., K. Morona, and J. C. Paton. 1997. Characterization of the locus encoding the *Streptococcus pneumoniae* type 19F capsular polysaccharide biosynthesis pathway. *Mol. Microbiol.* 23:761-763.
30. Muñoz, R., M. Mollerach, K. Lopez and E. Garcia. 1997. Molecular organization of the genes required for the synthesis of type 1 capsular polysaccharide of *Streptococcus pneumoniae*; formation of binary encapsulated pneumococci and identification of cryptic dTDP-rhamnose biosynthesis genes. *Mol. Microbiol.* 25:79-92.
31. Pearce B. J., Y. B. Yin, and H. R. Masure. 1993. Genetic identification of exported proteins in *Streptococcus pneumoniae*. *Mol. Microbiol.* 9:1037-1050.
32. Roberts, I. S. 1996. The biochemistry and genetics of capsular polysaccharide production in bacteria. *Ann. Rev. Microbiol.* 50:285-315.
33. Rossbach, S., D. A. Kulpa, U. Rossbach, and F. J. de Bruin. 1994. Molecular and genetic characterization of the rhizopine catabolism (mocABRC) genes of *Rhizobium meliloti* L5-30. *Mol. Gen. Genet.* 245:11-24.
34. Rubens, C. E., L. M. Heggen, R. F. Haft, and R. M. Wessels. 1993. Identification of *cpsD*, a gene essential for type III capsule expression in group B streptococci. *Mol. Microbiol.* 8:843-855.
35. Rubens, C. E., L. M. R. Wessels, L. M. Heggen, and D. L. Kasper. 1987. Transposon mutagenesis of type III group B Streptococcus: correlation of capsule expression with virulence. *Proc. Natl. Acad. Sci. USA* 84:7208-7212.
36. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. Molecular cloning. A laboratory manual. Second edition. Cold Spring Harbor Laboratory Press. Cold Spring Harbor, New York.

37. Smith, H. E., U. Vecht, H. J. Wisselink, N. Stockhove-Zurwieden, Y. Biermann, and M. A. Smits. 1996. Mutants of *Streptococcus suis* types 1 and 2 impaired in expression of muramidase-released protein and extracellular protein induce disease in newborn germfree pigs. *Infect Immun.* 64:4409-4412.
38. Smith, H. E., H. J. Wisselink, U. Vecht, A. L. J. Gielkens and M. A. Smits. 1995. High-efficiency transformation and gene inactivation in *Streptococcus suis* type 2. *Microbiol.* 141:181-188.
39. Sreenivasan, P. K., D. L. LeBlanc, L. N. Lee, and P. Fives-Taylor. 1991. Transformation of *Actinobacillus actinomycetemcomitans* by electroporation, utilizing constructed shuttle plasmids. *Infect. Immun.* 59:4621-4627.
40. Stringele F., J. R. Neeser, and B. Mollet. 1996. Identification and characterization of the *eps* (exopolysaccharide) gene cluster from *Streptococcus thermophilus* Sfi6. *J. Bacteriol.* 178:1680-1690.
41. Stockhove-Zurwieden, N., U. Vecht, H. J. Wisselink, H. van Lieshout, and H. E. Smith. 1996. Comparative studies on the pathogenicity of different *Streptococcus suis* serotype 1 strains. In Proceedings of the 14th IPVS Congress. pp. 299.
42. van Kranenburg, R., J. D. Marugg, I. I. van Swam, N. J. Willem and W. M. de Vos. 1997. Molecular characterization of the plasmid-encoded *eps* gene cluster essential for exopolysaccharide biosynthesis in *Lactococcus lactis*. *Mol. Microbiol.* 24:387-397.
43. van Leengoed, L. A., E. M. Kamp, and J. M. A. Pol. 1989. Toxicity of *Haemophilus pleuropneumoniae* to porcine lung macrophages. *Vet. Microbiol.* 19:337-349.
44. van Leengoed, L. A. M. G., U. Vecht, and E. R. M. Verheyen. 1987. *Streptococcus suis* type 2 infections in pigs in The Netherlands (part two). *Vet Quart.* 9, 111-117.
45. Vecht, U., J. P. Arends, E. J. van der Molen, and L. A. M. G. van Leengoed. 1989. Differences in virulence between two strains of *Streptococcus suis* type 2 after experimentally induced infection of newborn germfree pigs. *Am. J. Vet. Res.* 50:1037-1043.
46. Vecht, U., L. A. M. G. van Leengoed, and E. R. M. Verheyen. 1985. *Streptococcus suis* infections in pigs in The Netherlands (part one). *Vet. Quart.* 7:315-321
47. Vecht, U., H. J. Wisselink, M. L. Jellema, and H. E. Smith. 1991. Identification of two proteins associated with virulence of *Streptococcus suis* type 2. *Infect. Immun.* 59:3156-3162.

48. Vecht, U., H. J. Wisselink, N. Stockhove-Zurwieden, and H. E. Smith. 1996. Characterization of virulence of the *Streptococcus suis* serotype 2 reference strain Henrichsen S. 735 in newborn gnotobiotic pigs. *Vet. Microbiol.* 51:125-136.

49. Vecht, U., H. J. Wisselink, J. E. van Dijk, and H. E. Smith. 1992. Virulence of *Streptococcus suis* type 2 strains in newborn germfree pigs depends on phenotype. *Infect. Immun.* 60:550-556.

50. Wagenaar, F., G. L. Kok, J. M. Broekhuijsen-Davies, and J. M. A. Pol. 1993. Rapid cold fixation of tissue samples by microwave irradiation for use in electron microscopy. *Histochemical J.* 25:719-725.

51. Wessels, M. R. and M. S. Bronze. 1994. Critical role of the group A streptococcal capsule in pharyngeal colonization and infection in mice. *Proc. Natl. Acad. Sci. USA* 91:12238-12242.

52. Wessels, M. R., A. E. Moses, J. B. Goldberg, and T. J. DiCesare. 1991. Hyaluronic acid capsule is a virulence factor for mucoid group A streptococci. *Proc. Natl. Acad. Sci. USA*. 88:8317-8321.

53. Yamane, K., M. Kumamano, and K. Kurita. 1996. The 25°-36° region of the *Bacillus subtilis* chromosome: determination of the sequence of a 146 kb segment and identification of 113 genes. *Microbiol.* 142:3047-3056.

54. Butler, J. C., R. F. Breiman, H. B. Lipman, J. Hofmann, and R. R. Facklam. 1995. Serotype distribution of *Streptococcus pneumoniae* infections among preschool children in the United States, 1978-1994: implications for development of a conjugate vaccine. *J. Infect. Dis.* 171:885-889.

55. Charland, N., M. Jacques, S. Lacoutre and N. Gottschalk. 1997. Characterization and protective activity of a monoclonal antibody against a capsular epitope shared by *Streptococcus suis* serotypes 1, 2 and 1/2. *Microbiol.* 143:3607-3614.

56. Gottschalk, M., R. Higgins, M. Jacques, K. R. Mittal, and J. Henrichsen. Description of 14 new capsular types of *Streptococcus suis*. *J. Clin. Microbiol.* 27:2633-2636.

57. Heath, P. J., B. W. Hunt, and J. P. Duff. 1996. *Streptococcus suis* serotype 14 as a cause of pig disease in the UK. *Vet. Rec.* 2:450-451.

58. Hommez, J., L. A. Devrieze, J. Henrichsen, and F. Castryck. 1986. Identification and characterization of *Streptococcus suis*. *Vet. Microbiol.* 16:349-355.

59. Killper-Balz, R., and K. H. Schleifer. 1987. *Streptococcus suis* sp. nov. nom. rev. *Int. J. Syst. Bacteriol.* 37:160-162.

60. Kolkman, M. A. B., B. A. M. van der Zeijst, and P. J. M. Nuijten. 1998. Diversity of capsular polysaccharide synthesis gene clusters in *Streptococcus pneumoniae*. Submitted for publication.

61. Lee, J. C., S. Xu, A. Albus, and P. J. Livolsi. 1994. Genetic analysis of type 5 capsular polysaccharide expression by *Staphylococcus aureus*. *J. Bacteriol.* 176:4883-4889.

62. Reek, F. H., M. A. Smits, E. M. Kamp, and H. E. Smith. 1995. Use of multiscreen plates for the preparation of bacterial DNA suitable for PCR. *BioTechniques* 19:282-285.

63. Sau, S., N. Bhasin, E. R. Wann, J. C. Lee, T. J. Foster, and C. Y. Lee. 1997. The *Staphylococcus aureus* allelic genetic loci for serotype 5 and 8 capsule expression contain the type-specific genes flanked by common genes. *Microbiol.* 143: 2395-2405.

64. Sau, S., and C. Y. Lee. 1996. Cloning of type 8 capsule genes and analysis of gene clusters for the production of different capsular polysaccharides in *Staphylococcus aureus*. *J. Bacteriol.* 178:2118-2126.

65. Sau, S., and C. Y. Lee. 1997. Molecular characterization and transcriptional analysis of type 8 capsule genes in *Staphylococcus aureus*. *J. Bacteriol.* 179:1614-1621.

66. Smith, H. E., M. Rijnsburger, N. Stockhofe-Zurwieden, H. J. Wisselink, U. Vecht, and M. A. Smits. 1997. Virulent strains of *Streptococcus suis* serotype 2 and highly virulent strains of *Streptococcus suis* serotype 1 can be recognized by a unique ribotype profile. *J. Clin. Microbiol.* 35:1049-1053.

67. Yamazaki, M., L. Thorne, M. Mikolajczak, R. W. Armentrout, and T. J. Pollock. 1996. Linkage of genes essential for synthesis of a polysaccharide capsule in *Sphingomonas* strain S88. *J. Bacteriol.* 178:2676-2687.

68. Zhang, L., A. Al-Hendy, P. Toivanen, and M. Skurnik. 1993. Genetic organization and sequence of the *rfb* gene cluster of *Yersinia enterolitica* serotype 0:3: similarities to the dTDP-L-rhamnose biosynthesis pathway of *Salmonella* and to the bacterial polysaccharide transport systems. *Mol. Microbiol.* 9:309-321.

69. Clifton-Hadley, F.A. (1983). *Streptococcus suis* type 2 infections. *Br. Vet. J.* 139, 1-5.

70. Vecht, U., van Leengoed, L. A. M. G. and Verheyen, E. R. M. (1985). *Streptococcus suis* infections in pigs in The Netherlands (part one). *Vet. Quart.* 7, 315-321.

71. Arends, J. P. and Zanen, H. C. (1988). Meningitis caused by *Streptococcus suis* in humans. *Rev. Infect. Dis.* 10, 131-137.

72. Hommez, J., Devrieze, L.A., Henrichsen, J. and Castryck, F. (1986). Identification and characterization of *Streptococcus suis*. *Vet. Microbiol.* 16, 349-355.

73. Killper-Balz, R. and Schleifer, K. H. (1987). *Streptococcus suis* sp. nov. nom.rev. Int. J. Syst. Bacteriol. 37, 160-162.

74. Gottschalk, M., Higgins, R. and Jacques, M. (1993). Production of capsular material by *Streptococcus suis* serotype 2 under different conditions. Can. J. Vet. Res. 57, 49-52.

75. Higgins, R. and Gottschalk, M. (1990). Update on *Streptococcus suis* identification. J. Vet. Diagn. Invest. 2, 249-252.

76. Gottschalk, M., Higgins, R., Jacques, M., Beaudoin, M. and Henrichsen, J. (1991). Characterization of six new capsular types (23 through 28) of *Streptococcus suis*. J. Clin. Microbiol. 29, 2590-2594.

77. Gottschalk, M., Higgins, R., Jacques, M., Mittal, K. R. and Henrichsen, J. (1989). Description of 14 new capsular types of *Streptococcus suis*. J. Clin. Microbiol. 27, 2633-2636.

78. Higgins, R., Gottschalk, M., Boudreau, M., Lebrun, A. and Henrichsen, J. (1995). Description of six new capsular types (28 through 34) of *Streptococcus suis*. J. Vet. Diagn. Invest. 7, 405-406

79. Aarestrup, F. M., Jorsal, S. E. and Jensen, N. E. (1998). Serological characterization and antimicrobial susceptibility of *Streptococcus suis* isolates from diagnostic samples in Denmark during 1995 and 1996. Vet. Microbiol. 15, 59-66.

80. MacLennan, M., Foster, G., Dick, K., Smith, W. J. and Nielsen, B. (1996). *Streptococcus suis* serotypes 7, 8 and 14 from diseased pigs in Scotland. Vet Rec. 139, 423-424.

81. Sihvonen, L., Kurl, D. N. and Henrichsen, J. (1988). *Streptococcus suis* isolates from pigs in Finland. Acta Vet. Scand. 29, 9-13.

82. Boetner, A. G., Binder, M. and Bille-Hansen, V. (1987). *Streptococcus suis* infections in Danish pigs and experimental infection with *Streptococcus suis* serotype 7. Acta Path. Microbiol. Immunol. Scand. Sect. B, 95, 233-239.

83. Smith, H. E., Veenbergen, V., van der Velde, J., Damman, M., Wisselink, H. J. and Smits, M. A. (1999). The cps genes of *Streptococcus suis* serotypes 1, 2 and 9: development of rapid serotype-specific PCR assays. J. Clin. Microbiol. submitted

84. Smith, H. E., Damman, M., van der Velde, J., Wagenaar, F., Wisselink, H. J., Stockhose-Zurwieden, N. and Smits, M. A. (1999). Identification and characterization of the cps locus of *Streptococcus suis* serotype 2: the capsule protects against phagocytosis and is an important virulence factor. Infect. Immun. 67, 1750-1756.

85. Miller, J. (1972). Experiments in Molecular Genetics. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY.
86. Sambrook, J., E. F. Fritsch, and T. Maniatis. (1989). Molecular cloning: a laboratory manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY.
87. Allen, A. and Maskell, D. (1996). The identification, cloning and mutagenesis of a genetic locus required for lipopolysaccharide biosynthesis in *Bordetella pertussis*. Mol. Microbiol. 19, 37-52.
88. Wang, L. and Reeves, P. R. (1998). Organization of *Escherichia coli* O157 O antigen gene cluster and identification of its specific genes. Infect. Immun. 66, 3545-3551.
89. Wisselink, H. J., Reek, F. H., Vecht, U., Stockhove-Zurwieden, N., Smits, M. A. and Smith, H. E. (1999). Detection of virulent strains of *Streptococcus suis* type 2 and highly virulent strains of *Streptococcus suis* type 1 in tonsillar specimens of pigs by PCR. Vet. Microbiol. 67, 143-157.
90. Konings, R. N. H., Verhoeven, E. J. M. and Peeters, B. P. H. (1987). pKUN vectors for the separate production of both DNA strands of recombinant plasmids. Methods Enzymol. 153, 12-34.

ABSTRACT OF THE DISCLOSURE

The invention relates to *Streptococcus suis* infection in pigs, vaccines directed against those infections and tests for diagnosing *Streptococcus suis* infections. The invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* or a gene or gene fragment derived thereof. The invention further provides a nucleic acid probe or primer allowing species or serotype-specific detection of *Streptococcus suis*. The invention also provides a *Streptococcus suis* antigen and vaccine derived thereof.